



**US Army Corps
of Engineers®**
Alaska District

DRAFT Feasibility Report and Environmental Assessment

Homer Navigation Improvements Homer, Alaska Appendix P: Fieldwork Summary



May 2026

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Attachment 1. Aquatic Resource Permit Documentation
Attachment 2. Sediment Analysis Plan and Chemical Analysis Results

ACRONYMS AND ABBREVIATIONS

°C	Degrees Celsius
ADEC	Alaska Department of Environmental Conservation
ADF&G	Alaska Department of Fish and Game
ARP	Aquatic Resource Permit
BUPA	Beneficial Use Placement Area
CPUE	Catch-per-unit-effort
CTD	Conductivity, Temperature, and Depth
DWDS	Deep Water Disposal Site
eDNA	Environmental DNA
GPS	Global Positioning System
HTT	Homer Trawl Transect
IFR/EA	Integrated Feasibility Report and Environmental Assessment
MLLW	Mean Lower Low Water
NCBI	National Center for Biotechnology Information
NMFS	National Marine Fisheries Service
PSS	Practical Salinity Scale
RBA	Revised Harbor Basin
SRA	Sequence Read Archive
USACE	U.S. Army Corps of Engineers

APPENDIX P - FIELDWORK SUMMARY

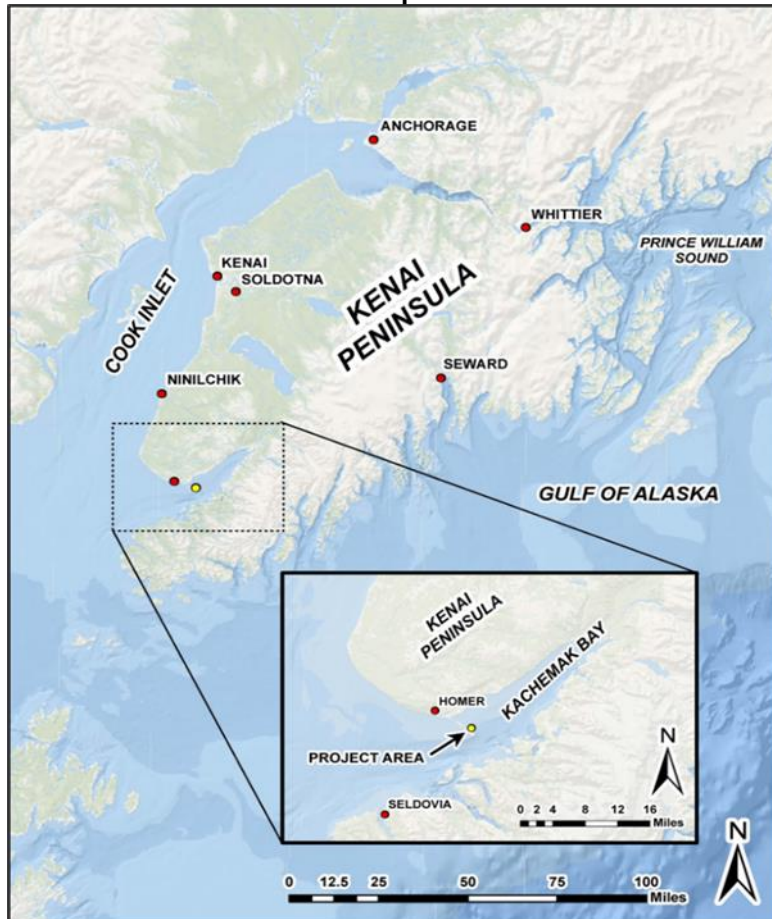
P.1 Introduction

This document is the Fieldwork Summary for fieldwork completed by U.S. Army Corps of Engineers (USACE) in 2024 through 2025 for the Homer Navigation Improvements, AK, Integrated Feasibility Report and Environmental Assessment (IFR/EA), authorized under Section 204 of the Flood Control Act of 1948. This feasibility study was requested by the City of Homer (the non-Federal sponsor), in response to navigation inefficiencies within Homer Port and Harbor (herein Homer Harbor).

P.1.1 Project Location

The Homer Harbor is located in Homer, Alaska at the end of the Homer Spit (Figure P-1). The Homer Spit is a stretch of land that extends into Kachemak Bay, which is located on the southern tip of the Kenai Peninsula and drains into the Gulf of Alaska by way of Cook Inlet.

Figure P-1. Location of Homer with respect to Cook Inlet and Kachemak Bay.



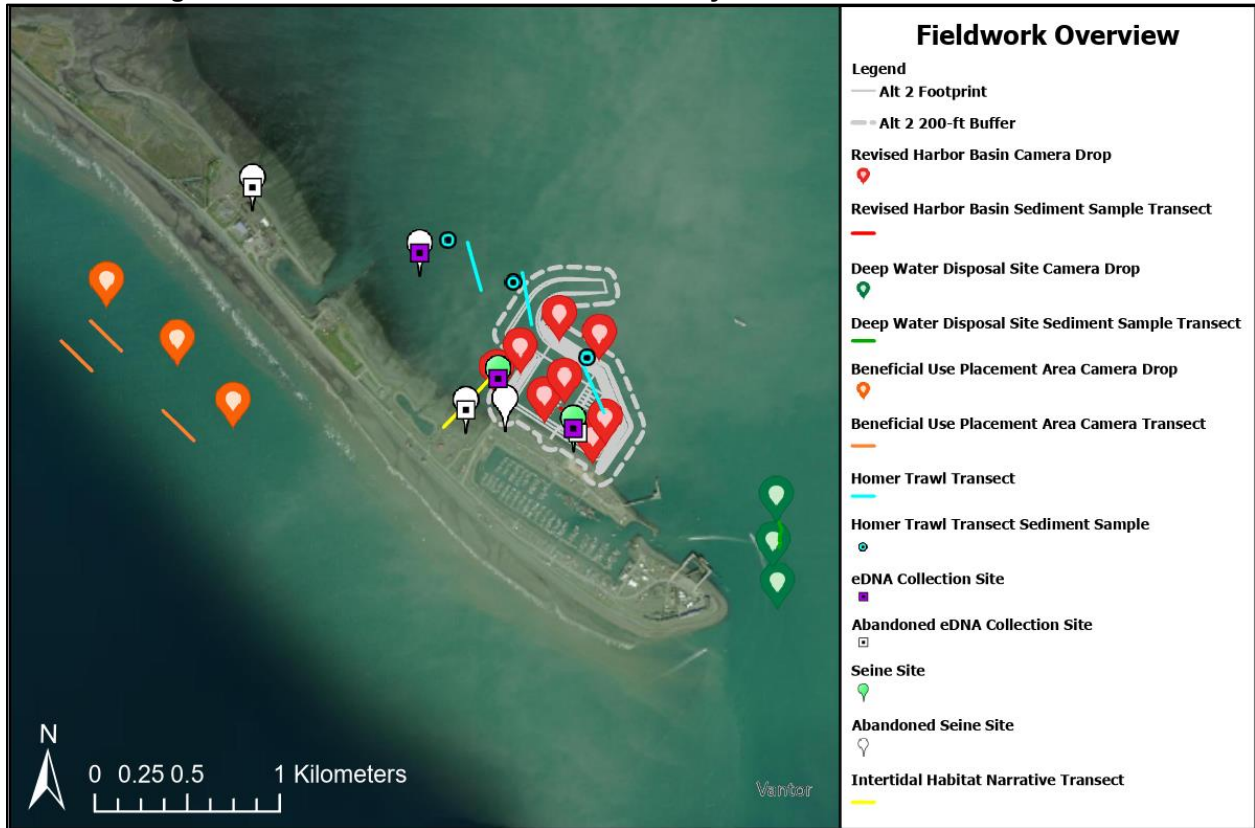
P.2 Environmental Fieldwork

Biologists from USACE Alaska District collected baseline ecological data from areas of interest (Figure P-2) in Kachemak Bay near the Homer Harbor in the years 2024 and 2025. Surveys included the use of both traditional survey techniques and contemporary molecular analysis (Figure P-3). The data collected is intended to supplement the environmental analysis of the Homer Navigation Improvements, AK, IFR/EA.

Figure P-2. Fieldwork area of interest for environmental surveys conducted in 2024 and 2025.



Figure P-3. Overview of environmental surveys conducted in 2024 and 2025.



The Fieldwork Overview figure has been updated to show the Alternative 2 footprint and its buffer for spatial context. It does not, however, include the survey sectors for the Steller's eider surveys mentioned in Section P.2.5.

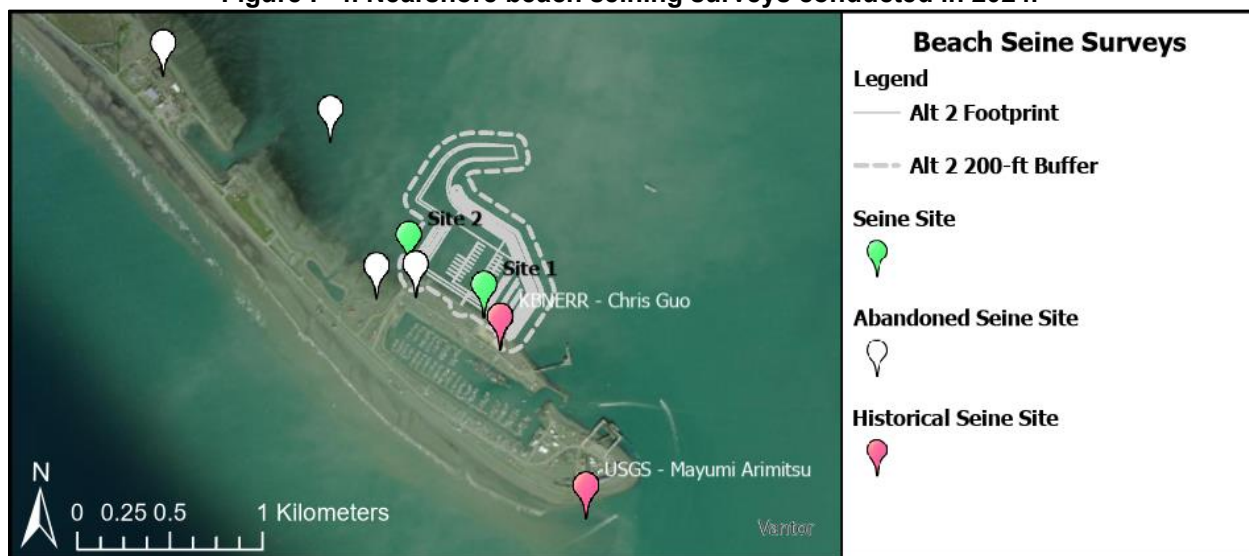
P.2.1 General Data Collection

A Global Positioning System (GPS) tablet was used to collect location data in the field, navigate between stations, take georeferenced photos, import and export geometry from ArcGIS Pro, and take notes during fieldwork for the feasibility study. Maps and charts of the study area were downloaded prior to the start fieldwork efforts in order to project location data in the appropriate spatial reference.

P.2.2 Nearshore Beach Seining Surveys

To assess the nearshore fish and invertebrate community near the Homer Harbor, beach seining was conducted from April 2024 to September 2024 (Figure P-4). Seine sites were resampled on a monthly basis. All beach seining activities were conducted under Alaska Department of Fish and Game (ADF&G) Aquatic Resource Permit (ARP) No. CF-24-005 and ARP No. CF-25-029. The Nearshore Fish Study Plans, ARPs, and Collection Reports for this study are located in Attachment 1. USACE will coordinate with the National Marine Fisheries Service (NMFS) to upload data into the *Nearshore Fish Atlas of Alaska*, to make data publicly available.

Figure P-4. Nearshore beach seining surveys conducted in 2024.



P.2.2.1 Seining Methodology

Beach seines were set during daylight hours within the project area at two sites (Figure P-4). Nearshore fish and invertebrates were sampled at low tide in waters less than 0.5 meters deep. Tidal variation is extreme in Kachemak Bay and Cook Inlet. In areas with a large tide differential, beach seines are typically set at low tide to both standardize the sampling protocol and to sample the richest habitat. Seine hauls were made at two sites for six sampling events, for a total of 12 sets (Table P-1).

Table P-1. Nearshore beach seining site and set information.

Location			Date						# of Sets
Site	Latitude (°N)	Longitude (°W)	9-Apr-24	8-May-24	6-Jun-24	22-Jul-22	20-Aug-24	19-Sep-24	
1	59.6099	-151.4294	X	X	X	X	X	X	6
2	59.6097	-151.4332	X	X	X	X	X	X	6
Number of Sets			2	2	2	2	2	2	12

The seine length was 37 meters long with variable mesh, tapering from 5 meters deep at the center to 1 meter deep at the ends. The two end panels were each 10 meters long with 32-millimeter stretch mesh. The two intermediate panels were each 4 meters long with 6-millimeter square mesh. The bunt was 9 meters long with 3.2-millimeter square mesh. The seine was equipped with a leadline and a floatline so that the bottom edge of the net would maintain contact with the bottom substrate and the top would float on the water surface. Two braided nylon lines, approximately 25–30 meters long, were attached to the seine—one per end. The seine was set as a round haul from the bow of a small jet-powered landing craft boat due to shallow water conditions. One line was held on the beach while the net was paid out from the boat in a semi-circle to a second position on the beach, about 18 meters from the starting point. The net was brought onto shore by pulling on both lines concurrently from shore, trapping fish and other organisms within the net. The captured fish and other organisms were quickly transferred to aerated buckets of brackish site water for identification and measurement.

Beach seining was attempted further north of the project footprint and during high tide at the start of the fieldwork season in 2024. The sites further north and along the high tide shoreline were abandoned due to the presence of boulders and boat operation constraints. The seine net would catch the boulders and prevent a smooth retrieval that increased the potential for catch loss. The area further north and near the high tide shoreline also presented a more gradual slope. Due to safety concerns with the gradual slope, potential for larger rock, and relatively fast changing tide at these initial sites, USACE adjusted site locations for the rest of the nearshore beach seine surveys.

Beach seining was not conducted in 2025 due to the timing of the low tides. The monthly low tides that were required for beach seining occurred at night (e.g., 2:00 a.m.), which was not feasible from a logistical perspective (i.e., working in the dark and processing fish at night without adequate lighting), or from a safety perspective (working in the dark, at night, from a small vessel with no deck lights, in an area that is foul with boulders and rocks).

P.2.2.2 Seining Catch

Catch was sorted and counted, and individuals were identified to the lowest taxon. Data collected included the number of species caught, the average total length to the nearest millimeter of 10 individuals within each age cohort, and any incidental notes. Fish life stages (i.e., young-of-the-year, juvenile, and adult) were assigned in the field based on phenotypic characteristics using Johnson et al. (2015) as a key.

A total of 321 individuals were caught across all sites and dates, representing at least 29 taxa (Table P-2 and Table P-3). Life-stage cohorts were combined to represent a single catch metric in these tables, and the sampling sites correspond with Figure P-4. Total catch of fishes and invertebrates captured in Kachemak Bay near Homer Harbor by site is depicted in Table P-2. Total catch and catch-per-unit-effort (CPUE; unit = set) of fishes and invertebrates captured by trip date is depicted in Table P-3. Variation in catch was observed by site and by sampling date, though species composition was relatively consistent between both sites. The greatest diversity of species occurred at Site 2, with 27 taxa represented in total compared to the 19 taxa that were caught at Site 1 (Table P-2). The greatest number of organisms were caught in August (116 individuals) followed by September (104 individuals; Table P-3). There were only four individuals caught in April, which was the least number of individuals caught across all sampling dates (Table P-3). See Table P-1 for beach seine sit and set information.

Table P-2. Total catch by site for nearshore beach seining.

Common Name	Scientific Name	Site 1	Site 2
Arctic Shanny	<i>Stichaeus punctatus</i>	-	1
Buffalo Sculpin	<i>Enophrys bison</i>	-	2
Capelin	<i>Mallotus villosus</i>		1
Chinook Salmon	<i>Oncorhynchus tshawytscha</i>	-	1
Crescent Gunnel	<i>Pholis laeta</i>	9	8
Graceful Decorator Crab	<i>Oregonia gracilis</i>	9	1
Great Sculpin	<i>Myoxocephalus polyacanthocephalus</i>	12	10
Helmet Crab	<i>Telmessus cheiragonus</i>	5	9
Kelp Greenling	<i>Hexagrammos decagrammus</i>	10	17
Masked Greenling	<i>Hexagrammos yoctograms</i>	28	29
Pacific Halibut	<i>Hippoglossus stenolepis</i>	1	4
Pacific Staghorn Sculpin	<i>Leptocottus armatus</i>	1	1
Padded Sculpin	<i>Artedius fenestralis</i>	2	
Ribbon Snailfish	<i>Liparis cyclopus</i>	1	1
Rock Greenling	<i>Hexagrammos lagocephalus</i>	8	9
Rock Sole	<i>Lepidopsetta bilineata</i>	2	10
Saffron Cod	<i>Eleginus gracilis</i>	2	3
Sharpnose sculpin	<i>Clinocottus acuticeps</i>	1	
Silverspotted Sculpin	<i>Blepsias cirrhosus</i>	8	8
Snake Prickleback	<i>Lumpenus sagitta</i>	-	33
Sturgeon Poacher	<i>Podothecus accipenserinus</i>	-	1
Surf Smelt	<i>Hypomesus pretiosus</i>	-	4
Three-spine Stickleback	<i>Gasterosteus aculeatus</i>	-	1
Tube-nose Poacher	<i>Pallasina barbata</i>	-	1
Unidentified Flatfish	-	-	12
Unidentified Sculpin	-	1	13
Unidentified Shrimp	-	31	3
Unidentified Snailfish	-	1	2
Whitespotted Greenling	<i>Hexagrammos stelleri</i>	1	3
Total Catch By Site		133	188

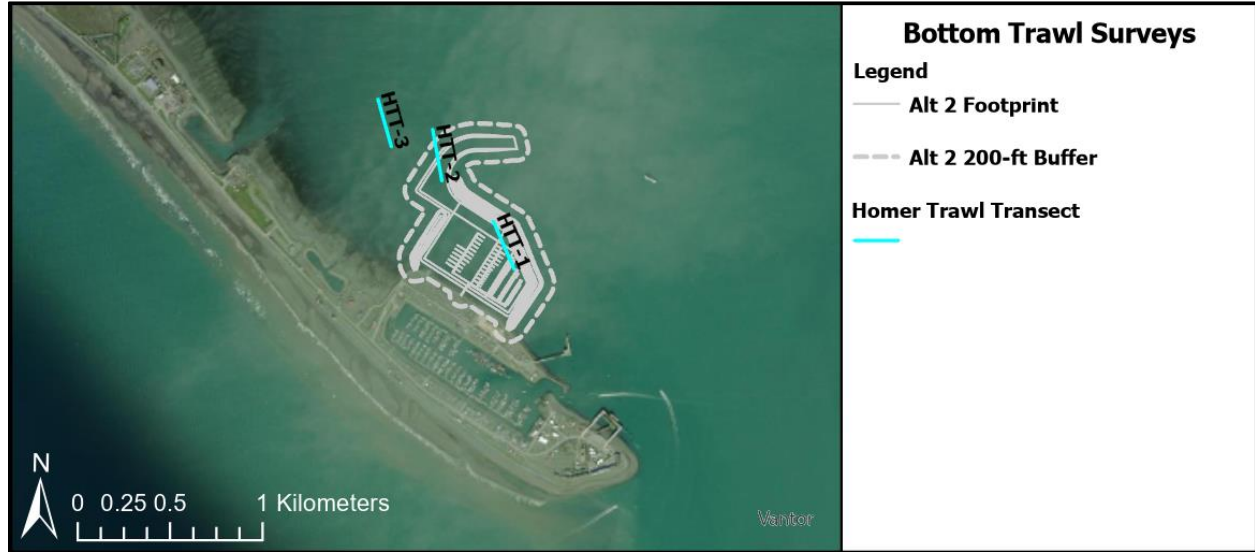
Table P-3. Total catch by date for nearshore beach seining.

Common Name	Scientific Name	9-Apr-24		8-May-24		6-Jun-24		22-Jul-24		20-Aug-24		19-Sep-24	
		Catch	CPUE	Catch	CPUE	Catch	CPUE	Catch	CPUE	Catch	CPUE	Catch	CPUE
Arctic Shanny	<i>Stichaeus punctatus</i>	-	-	-	-	-	-	1	0.5	-	-	-	-
Buffalo Sculpin	<i>Enophrys bison</i>	-	-	-	-	-	-	-	-	2	1	-	-
Capelin	<i>Mallotus villosus</i>	1	0.5	-	-	-	-	-	-	-	-	-	-
Chinook Salmon	<i>Oncorhynchus tshawytscha</i>	-	-	-	-	1	0.5	-	-	-	-	-	-
Crescent Gunnel	<i>Pholis laeta</i>	-	-	1	0.5	1	0.5	-	-	8	4	7	3.5
Graceful Decorator Crab	<i>Oregonia gracilis</i>	-	-	6	3	1	0.5	-	-	1	0.5	2	1
Great Sculpin	<i>Myoxocephalus polyacanthocephalus</i>	1	0.5	-	-	1	0.5	-	-	-	-	20	10
Helmet Crab	<i>Telmessus cheiragonus</i>	-	-	1	0.5	5	2.5	2	1	2	0	4	2
Kelp Greenling	<i>Hexagrammos decagrammus</i>	-	-	1	0.5	6	3	2	1	14	7	4	2
Masked Greenling	<i>Hexagrammos octogrammus</i>	-	-	-	-	-	-	32	16	19	9.5	6	3
Pacific Halibut	<i>Hippoglossus stenolepis</i>	-	-	-	-	-	-	2	1	-	-	3	1.5
Pacific Staghorn Sculpin	<i>Leptocottus armatus</i>	1	0.5	-	-	-	-	-	-	-	-	1	0.5
Padded Sculpin	<i>Artedius fenestralis</i>	-	-	-	-	-	-	-	-	2	-	-	-
Ribbon Snailfish	<i>Liparis cyclopus</i>	-	-	-	-	-	-	-	-	-	-	2	1
Rock Greenling	<i>Hexagrammos lagocephalus</i>	-	-	1	0.5	8	4	4	2	1	0.5	3	1.5
Rock Sole	<i>Lepidopsetta bilineata</i>	-	-	2	1	-	-	5	2.5	3	1.5	2	1
Saffron Cod	<i>Eleginus gracilis</i>	-	-	-	-	-	-	2	1	3	1.5	-	-
Sharpnose sculpin	<i>Clinocottus acuticeps</i>	1	0.5	-	-	-	-	-	-	-	-	-	-
Silverspotted Sculpin	<i>Blepsias cirrhosus</i>	-	-	-	-	-	-	-	-	6	3	10	5
Snake Prickleback	<i>Lumpenus sagitta</i>	-	-	-	-	-	-	-	-	33	16.5	-	-
Sturgeon Poacher	<i>Podothecus accipenserinus</i>	-	-	-	-	-	-	1	0.5	-	-	-	-
Surf Smelt	<i>Hypomesus pretiosus</i>	-	-	-	-	-	-	-	-	-	-	4	2
Three-spine Stickleback	<i>Gasterosteus aculeatus</i>	-	-	-	-	-	-	-	-	1	0.5	-	-
Tubenose Poacher	<i>Pallasina barbata</i>	-	-	-	-	-	-	-	-	-	-	1	0.5
Unidentified Flatfish	-	-	-	-	-	-	-	-	-	12	6	-	-
Unidentified Sculpin	-	-	-	-	-	1	0.5	1	0.5	5	2.5	7	3.5
Unidentified Shrimp	-	-	-	-	-	2	1	2	1	2	1	28	14
Unidentified Snailfish	-	-	-	-	-	-	-	1	0.5	2	1	-	-
Whitespotted Greenling	<i>Hexagrammos stelleri</i>	-	-	2	1	1	0.5	1	0.5	-	-	-	-
Total Catch by Date		4	-	14	-	27	-	56	-	116	-	104	-

P.2.3 Bottom Trawling Surveys

To assess the fish community occupying areas of deep water habitat near the Homer Harbor, bottom trawling was conducted from April 2024 to September 2024, and January 2025 to March 2025 (Figure P-5). Bottom trawl transects were resampled on a monthly basis. All bottom trawl activities were conducted under ADF&G ARP No. CF-24-005 and ARP No. CF-25-029 (Attachment 1).

Figure P-5. Bottom trawl surveys conducted in 2024 and 2025.



P.2.3.1 Trawling Methodology

Bottom trawling occurred during daylight hours along three transects (Figure P-5) for six sampling events in 2024 and three sampling events in 2025 (Table P-4), for a total of 27 hauls. The trawl net was 5.2 meters long, which included a 1.7-meter-long cod end of 3.2-millimeter stretch mesh protected by an outer skirt of 29-millimeter stretch mesh. The mouth of the net was 2.6 meters wide, 1.2 meters deep, and connected to two weighted otter doors (33 centimeters by 61 centimeters) on each end. The doors were attached to a 6.3-meter-long bridle of 1.3-centimeter braided line, which was then attached to the City of Homer Harbormaster’s tugboat with a 1.6-centimeter polypropylene tow line. Minimum scope ratio of the tow line was kept to at least 3 to 4:1. Once on the bottom, the trawl was towed for 5 minutes. Overall, the trawl was fished at speeds ranging from 2 to 3 knots over the seafloor, resulting in an average transect distance of 0.25 km. Tow direction was determined by the tugboat operator based on tide, currents, wind, and sea state. After tows for 5 minutes, the net was hauled back onto the stern of the tugboat by hand. Fish and invertebrates caught in the net were removed and immediately measured before being returned to site waters.

Table P-4. Bottom trawl transect and haul information.

Location			Date									# of Hauls	
Site	Latitude (°N)	Longitude (°W)	2024						2025				
			9-Apr	8-May	6-Jun	22-Jul	20-Aug	19-Sep	15-Jan	12-Feb	26-Mar		
1	59.6110	-151.4211	X	X	X	X	X	X	X	X	X	X	9
2	59.6152	-151.4276	X	X	X	X	X	X	X	X	X	X	9
3	59.6167	-151.4327	X	X	X	X	X	X	X	X	X	X	9
Number of Sets			3	3	3	3	3	3	3	3	3	3	27

P.2.3.2 Trawling Catch

Similar to beach seining, trawl catch was sorted and counted, and individuals were identified to the lowest taxon. Data collected included the number of species caught, the average total length to the nearest millimeter of 10 individuals within each age cohort, and any incidental notes. Fish life stages (i.e., young-of-the-year, juvenile, and adult) were assigned in the field based on phenotypic characteristics using Johnson et al. (2015) as a key.

A total of 213 individuals were caught across all transects and dates, representing at least 26 taxa (Table P-5 and Table P-6). Life-stage cohorts were combined to represent a single catch metric in these tables, and the bottom trawl sites correspond with Figure P-5. Total catch of fishes and invertebrates captured by transect via method of bottom trawling in Kachemak Bay near Homer Harbor is depicted in Table P-5. Total catch and CPUE of fishes and invertebrates captured by trip date is depicted in Table P-6. See Table P-4 for transect and haul information. Transect 2 had the greatest total number of individuals caught between the three transects, with 102 individuals. Transect 1 had the greatest diversity of taxa caught (19 taxa), comprised of 55 individuals (Table P-5). The summer sampling events in July, August, and September had the greatest numbers of catch across all sampling events, while March and April had the fewest number of individuals caught (Table P-5).

Table P-5. Total catch by transect bottom trawling.

Common Name	Scientific Name	Transect 1	Transect 2	Transect 3
Graceful Decorator Crab	<i>Oregonia gracilis</i>	3	10	5
Helmet Crab	<i>Telmessus cheiragonus</i>	1	1	1
Hermit Crab	<i>Paguroidea</i> sp.	-	2	-
Lyre Crab	<i>Hyas coarctatus</i>	1	1	1
Pacific Cod	<i>Gadus macrocephalus</i>	3	-	-
Pacific Halibut	<i>Hippoglossus stenolepis</i>	-	-	1
Pacific Herring	<i>Clupea pallasii</i>	1	-	-
Purple Sea Star	<i>Pisaster ochraceus</i>	3	1	1
Rock Greenling	<i>Hexagrammos lagocephalus</i>	-	1	-
Rock Sole	<i>Lepidopsetta bilineata</i>	4	6	3
Saffron Cod	<i>Eleginus gracilis</i>	1	2	3
Sea Pen	<i>Ptiloisarcus gurneyi</i>	21	4	-
Silverspotted Sculpin	<i>Blepsias cirrhosus</i>	1	-	1
Snake Prickleback	<i>Lumpenus sagitta</i>	4	1	-
Starry Flounder	<i>Platichthys stellatus</i>	1	-	-
Tanner Crab	<i>Chionoecetes bairdi</i>	1	-	1
Unidentified Anemone	-	1	-	-
Unidentified Arthropod	-	1	5	-
Unidentified Crab	-	-	11	1
Unidentified Flatfish	-	1	1	2
Unidentified Greenling	-	-	1	-
Unidentified Sculpin	-	2	-	-
Unidentified Sea Star	-	3	-	-
Unidentified Shrimp	-	-	54	34
Unidentified Snailfish	-	-	1	2
Yellowfin Sole	<i>Limanda aspera</i>	2	-	-
Total Catch by Transect		55	102	56

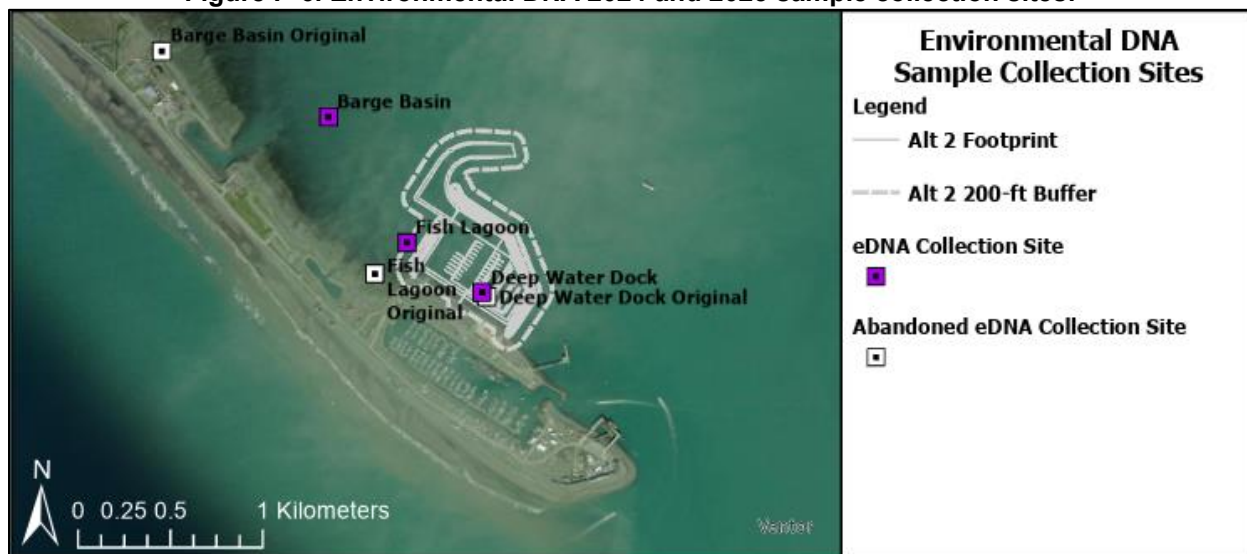
Table P-6. Total catch by trip date for bottom trawling.

Common Name	Scientific Name	10-Apr-24		8-May-24		6-Jun-24		22-Jul-24		20-Aug-24		19-Sep-24		15-Jan-25		12-Feb-25		26-Mar-25	
		Catch	CPUE	Catch	CPUE	Catch	CPUE	Catch	Catch	Catch	CPUE	Catch	CPUE	Catch	CPUE	Catch	CPUE	Catch	CPUE
Graceful Decorator Crab	<i>Oregonia gracilis</i>	-	-	-	-	9	1	-	-	-	-	9	1	-	-	-	-	-	-
Helmet Crab	<i>Telmessus cheiragonus</i>	-	-	1	0.11	1	0.11	2	0.22	-	-	-	-	-	-	-	-	-	-
Hermit Crab	<i>Paguroidea</i> sp.	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	0.11	-	-
Lyre Crab	<i>Hyas coarctatus</i>	-	-	-	-	1	0.11	-	-	1	0.11	1	0.11	-	-	-	-	-	-
Pacific Cod	<i>Gadus macrocephalus</i>	-	-	-	-	-	-	1	0.11	-	-	-	-	2	0.22	-	-	-	-
Pacific Halibut	<i>Hippoglossus stenolepis</i>	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	0.11	-	-
Pacific Herring	<i>Hippoglossus stenolepis</i>	-	-	-	-	-	-	-	-	1	0.11	-	-	-	-	-	-	-	-
Purple Sea Star	<i>Clupea pallasii</i>	-	-	-	-	3	0.33	-	-	-	-	1	0.11	-	-	1	0.11	-	-
Rock Greenling	<i>Hexagrammos lagocephalus</i>	-	-	-	-	-	-	-	-	1	0.11	-	-	-	-	-	-	-	-
Rock Sole	<i>Lepidopsetta bilineata</i>	-	-	1	0.11	4	0.44	2	0.22	2	0.22	-	-	2	0.22	1	0.11	1	0.11
Saffron Cod	<i>Eleginus gracilis</i>	-	-	-	-	-	-	1	0.11	5	0.56	-	-	-	-	-	-	-	-
Sea Pen	<i>Ptiloisarcus gurneyi</i>	-	-	-	-	-	-	-	-	-	-	5	0.56	-	-	19	2.11	1	0.11
Silverspotted Sculpin	<i>Blepsias cirrhosus</i>	-	-	-	-	-	-	1	0.11	1	0.11	-	-	-	-	-	-	-	-
Snake Prickleback	<i>Lumpenus sagitta</i>	-	-	-	-	-	-	-	-	4	0.44	1	0.11	-	-	-	-	-	-
Starry Flounder	<i>Platichthys stellatus</i>	-	-	-	-	-	-	-	-	1	0.11	-	-	-	-	-	-	-	-
Tanner Crab	<i>Chionoecetes bairdi</i>	-	-	-	-	-	-	-	-	-	-	2	0.22	-	-	-	-	-	-
Unidentified Anemone	-	-	-	-	-	-	-	-	-	-	-	1	0.11	-	-	-	-	-	-
Unidentified Arthropod	-	5	0.56	-	-	-	-	1	0.11	-	-	-	-	-	-	-	-	-	-
Unidentified Crab	-	1	0.11	5	0.56	-	-	-	-	6	0.67	-	-	-	-	-	-	-	-
Unidentified Flatfish	-	-	-	-	-	-	-	-	-	2	0.22	1	0.11	1	0.11	-	-	-	-
Unidentified Greenling	-	-	-	-	-	-	-	-	-	1	0.11	-	-	-	-	-	-	-	-
Unidentified Sculpin	-	-	-	-	-	-	-	-	-	-	-	-	-	2	0.22	-	-	1	0.11
Unidentified Sea Star	-	-	-	-	-	-	-	-	-	1	0.11	1	0.11	-	-	-	-	-	-
Unidentified Shrimp	-	2	0.22	3	0.33	3	0.33	26	2.89	-	-	52	5.78	2	0.22	-	-	-	-
Unidentified Snailfish	-	-	-	-	-	-	-	-	-	3	0.33	-	-	-	-	-	-	-	-
Yellowfin Sole	<i>Limanda aspera</i>	-	-	2	0.22	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Total Catch by Date		8	-	12	-	21	-	34	-	29	-	74	-	9	-	23	-	3	-

P.2.4 Environmental DNA Sample Collection and Analysis

An emerging method that improves detection of many aquatic species is environmental DNA (eDNA); which determines the presence of a species based on the collection, extraction, and amplification of DNA from the environment (Ficetola et al. 2008; Laramie et al. 2015). Sources of eDNA can include sloughed cells, feces, and bodily secretions. eDNA can be obtained from various environmental samples and reveals important information about present and past biodiversity within an ecosystem (Shu et al. 2020). For fish species, investigations into their distribution and ecology are often hindered by the challenge and complexity of working in these aquatic ecosystems (Goldberg et al. 2011). However, detection of eDNA within aquatic ecosystems is a reliable and cost-effective method for determining the distribution of various fish species (Jerde et al. 2011; Thomsen et al. 2011; Shu et al. 2020). USACE collected eDNA April through September 2024 and January through February 2025 to supplement data collected from nearshore beach seining and bottom trawling surveys (Figure P-6).

Figure P-6. Environmental DNA 2024 and 2025 sample collection sites.



P.2.4.1 Sample Methodology

For eDNA collection, 1-Liter water samples were collected at eDNA collection sites (Figure P-6), two of which overlap with beach seine Site 1 and Site 2 (Figure P-4), i.e., the Deep Water Dock and Fish Lagoon eDNA collection sites, respectively. At each sampling location, three replicate samples were taken to detect any potential inconsistencies in filtering techniques or downstream metagenomic library preparation. Collected water samples were filtered through a sterilized glass Buchner funnel that contained a 47-millimeter diameter cellulose nitrate filter with a 0.45-micrometer pore size. A vacuum pump was used to filter samples following procedures outlined in Protocol #1 of Laramie et al. (2015). Cross-contamination was mitigated by disinfecting surfaces and instruments with 10% bleach solution and then rinsing with distilled water between processing replicates from different field sites. In addition to the three replicate

samples collected at each field site, a negative control containing only distilled water was filtered to ensure our methods were effective in mitigating cross-contamination.

The eDNA collection sites were collocated with USACE beach seine sites. Thus, when the beach seine sites were adjusted, the eDNA collections sites were adjusted as well.

P.2.4.2 Molecular Analysis

Samples were sent to the U.S. Army Engineer Research and Development Center's Genetics Reconnaissance Team (Environmental Processes Branch, Environmental Laboratory) for DNA extraction, sequencing, and bioinformatic analysis (Table P-7).

Table P-7. Sample matrix for eDNA.

Tag Number	Replicate	Date	Location	Latitude (°N)	Longitude (°W)	Accession Number (MiFish-U)	Accession Number (AKOncCytB)
HOM-1	1	9-Apr-24	Deep Water Dock (Original)	59.6088	-151.4224	SAMN53858102	SAMN53858182
HOM-2	2	9-Apr-24	Deep Water Dock (Original)	59.6088	-151.4224	SAMN53858103	SAMN53858183
HOM-3	3	9-Apr-24	Deep Water Dock (Original)	59.6088	-151.4224	SAMN53858104	-
HOM-4	1	9-Apr-24	Fish Lagoon (Original)	59.6098	-151.4332	SAMN53858105	SAMN53858184
HOM-5	2	9-Apr-24	Fish Lagoon (Original)	59.6098	-151.4332	SAMN53858106	-
HOM-6	3	9-Apr-24	Fish Lagoon (Original)	59.6098	-151.4332	SAMN53858107	-
HOM-7	1	9-Apr-24	Barge Basin (Original)	59.6204	-151.4541	SAMN53858108	SAMN53858185
HOM-8	2	9-Apr-24	Barge Basin (Original)	59.6204	-151.4541	SAMN53858109	-
HOM-9	3	9-Apr-24	Barge Basin (Original)	59.6204	-151.4541	SAMN53858110	SAMN53858186
HOM-10	-	9-Apr-24	N/A: Method Blank	-	-	SAMN53858111	SAMN53858187
HOM-11	1	8-May-24	Deep Water Dock	59.6090	-151.4229	SAMN53858112	SAMN53858188
HOM-12	2	8-May-24	Deep Water Dock	59.6090	-151.4229	SAMN53858113	SAMN53858189
HOM-13	3	8-May-24	Deep Water Dock	59.6090	-151.4229	SAMN53858114	SAMN53858190
HOM-14	1	8-May-24	Fish Lagoon	59.6113	-151.4302	SAMN53858115	-
HOM-15	2	8-May-24	Fish Lagoon	59.6113	-151.4302	SAMN53858116	-
HOM-16	3	8-May-24	Fish Lagoon	59.6113	-151.4302	SAMN53858117	SAMN53858191
HOM-17	1	8-May-24	Barge Basin	59.6174	-151.4380	SAMN53858118	SAMN53858192
HOM-18	2	8-May-24	Barge Basin	59.6174	-151.4380	SAMN53858119	-
HOM-19	3	8-May-24	Barge Basin	59.6174	-151.4380	SAMN53858120	-
HOM-20		8-May-24	N/A: Method Blank	-	-	SAMN53858121	-
HOM-21	1	6-Jun-24	Barge Basin	59.6174	-151.4380	SAMN53858122	-
HOM-22	2	6-Jun-24	Barge Basin	59.6174	-151.4380	SAMN53858123	SAMN53858193
HOM-23	3	6-Jun-24	Barge Basin	59.6174	-151.4380	SAMN53858124	SAMN53858194
HOM-24	1	6-Jun-24	Fish Lagoon	59.6113	-151.4302	SAMN53858125	SAMN53858195
HOM-25	2	6-Jun-24	Fish Lagoon	59.6113	-151.4302	SAMN53858126	SAMN53858196
HOM-26	3	6-Jun-24	Fish Lagoon	59.6113	-151.4302	SAMN53858196	SAMN53858197
HOM-27	1	6-Jun-24	Deep Water Dock	59.6090	-151.4229	SAMN53858127	SAMN53858198
HOM-28	2	6-Jun-24	Deep Water Dock	59.6090	-151.4229	SAMN53858128	SAMN53858199

APPENDIX P: FIELDWORK SUMMARY
HOMER NAVIGATION IMPROVEMENTS

MAY 2026

Tag Number	Replicate	Date	Location	Latitude (°N)	Longitude (°W)	Accession Number (MiFish-U)	Accession Number (AKOncCytB)
HOM-29	3	6-Jun-24	Deep Water Dock	59.6090	-151.4229	SAMN53858129	-
HOM-30	-	6-Jun-24	N/A: Method Blank	-	-	SAMN53858131	-
HOM-31	1	22-Jul-24	Deep Water Dock	59.6090	-151.4229	SAMN53858132	SAMN53858200
HOM-32	2	22-Jul-24	Deep Water Dock	59.6090	-151.4229	SAMN53858133	SAMN53858201
HOM-33	3	22-Jul-24	Deep Water Dock	59.6090	-151.4229	SAMN53858134	-
HOM-34	1	22-Jul-24	Fish Lagoon	59.6113	-151.4302	SAMN53858135	SAMN53858202
HOM-35	2	22-Jul-24	Fish Lagoon	59.6113	-151.4302	SAMN53858136	SAMN53858203
HOM-36	3	22-Jul-24	Fish Lagoon	59.6113	-151.4302	SAMN53858137	SAMN53858204
HOM-37	1	22-Jul-24	Barge Basin	59.6174	-151.4380	SAMN53858138	SAMN53858205
HOM-38	2	22-Jul-24	Barge Basin	59.6174	-151.4380	SAMN53858139	SAMN53858206
HOM-39	3	22-Jul-24	Barge Basin	59.6174	-151.4380	SAMN53858140	SAMN53858207
HOM-40	-	22-Jul-24	N/A: Method Blank	-	-	SAMN53858141	-
HOM-41	1	20-Aug-24	Deep Water Dock	59.6090	-151.4229	SAMN53858142	-
HOM-42	2	20-Aug-24	Deep Water Dock	59.6090	-151.4229	SAMN53858143	SAMN53858208
HOM-43	3	20-Aug-24	Deep Water Dock	59.6090	-151.4229	SAMN53858144	SAMN53858209
HOM-44	1	20-Aug-24	Fish Lagoon	59.6113	-151.4302	SAMN53858145	SAMN53858210
HOM-45	2	20-Aug-24	Fish Lagoon	59.6113	-151.4302	SAMN53858146	SAMN53858211
HOM-46	3	20-Aug-24	Fish Lagoon	59.6113	-151.4302	SAMN53858147	SAMN53858212
HOM-47	1	20-Aug-24	Barge Basin	59.6174	-151.4380	SAMN53858148	-
HOM-48	2	20-Aug-24	Barge Basin	59.6174	-151.4380	SAMN53858149	SAMN53858213
HOM-49	3	20-Aug-24	Barge Basin	59.6174	-151.4380	SAMN53858150	SAMN53858214
HOM-50	-	20-Aug-24	N/A: Method Blank	-	-	SAMN53858151	SAMN53858215
HOM-51	1	19-Sep-24	Deep Water Dock	59.6090	-151.4229	SAMN53858152	SAMN53858216
HOM-52	2	19-Sep-24	Deep Water Dock	59.6090	-151.4229	SAMN53858153	SAMN53858217
HOM-53	3	19-Sep-24	Deep Water Dock	59.6090	-151.4229	SAMN53858154	-
HOM-54	1	19-Sep-24	Fish Lagoon	59.6113	-151.4302	SAMN53858155	SAMN53858218
HOM-55	2	19-Sep-24	Fish Lagoon	59.6113	-151.4302	SAMN53858156	SAMN53858219
HOM-56	3	19-Sep-24	Fish Lagoon	59.6113	-151.4302	SAMN53858157	SAMN53858220
HOM-57	1	19-Sep-24	Barge Basin	59.6174	-151.4380	SAMN53858158	-
HOM-58	2	19-Sep-24	Barge Basin	59.6174	-151.4380	SAMN53858159	-
HOM-59	3	19-Sep-24	Barge Basin	59.6174	-151.4380	SAMN53858160	SAMN53858221
HOM-60	-	19-Sep-24	N/A: Method Blank	-	-	SAMN53858161	-
HOM-61	1	16-Jan-25	Deep Water Dock	59.6090	-151.4229	SAMN53858162	-
HOM-62	2	16-Jan-25	Deep Water Dock	59.6090	-151.4229	SAMN53858163	-
HOM-63	3	16-Jan-25	Deep Water Dock	59.6090	-151.4229	SAMN53858164	-
HOM-64	1	16-Jan-25	Fish Lagoon	59.6113	-151.4302	SAMN53858165	-
HOM-65	2	16-Jan-25	Fish Lagoon	59.6113	-151.4302	SAMN53858166	-
HOM-66	3	16-Jan-25	Fish Lagoon	59.6113	-151.4302	SAMN53858167	SAMN53858222
HOM-67	1	16-Jan-25	Barge Basin	59.6174	-151.4380	SAMN53858168	SAMN53858223
HOM-68	2	16-Jan-25	Barge Basin	59.6174	-151.4380	SAMN53858169	-
HOM-69	3	16-Jan-25	Barge Basin	59.6174	-151.4380	SAMN53858170	SAMN53858224
HOM-70	-	16-Jan-25	N/A: Method Blank	-	-	SAMN53858171	SAMN53858225
HOM-71	1	13-Feb-25	Deep Water Dock	59.6090	-151.4229	SAMN53858172	-
HOM-72	2	13-Feb-25	Deep Water Dock	59.6090	-151.4229	SAMN53858173	-
HOM-73	3	13-Feb-25	Deep Water Dock	59.6090	-151.4229	SAMN53858174	SAMN53858226
HOM-74	1	13-Feb-25	Fishing Lagoon	59.6113	-151.4302	SAMN53858175	SAMN53858227
HOM-75	2	13-Feb-25	Fish Lagoon	59.6113	-151.4302	SAMN53858176	-
HOM-76	3	13-Feb-25	Fish Lagoon	59.6113	-151.4302	SAMN53858177	-

Tag Number	Replicate	Date	Location	Latitude (°N)	Longitude (°W)	Accession Number (MiFish-U)	Accession Number (AKOncCytB)
HOM-77	1	13-Feb-25	Barge Basin	59.6174	-151.4380	SAMN53858178	-
HOM-78	2	13-Feb-25	Barge Basin	59.6174	-151.4380	SAMN53858179	-
HOM-79	3	13-Feb-25	Barge Basin	59.6174	-151.4380	SAMN53858180	-
HOM-80	-	13-Feb-25	N/A: Method Blank	-	-	SAMN53858181	-

Samples HOM-1 through HOM-9 were collected during the first trip in April 2024 and are slightly offset from HOM samples collected after this first reconnaissance trip (thus the denotation with the term “Original” in Table P-7). Two metabarcoding primers were chosen, MiFish-U (Miya et al. 2015) and AKOncCytB (Menning et al. 2020). MiFish-U is a universal primer that targets the 12S-rRNA gene and possesses robust detection potential across numerous freshwater and marine fish taxa. However, MiFish-U exhibits low discriminatory power among salmonid species (Polanco et al. 2021; Miya et al. 2020). Thus, the AKOncCytB metabarcoding primer specifically designed for detection and discrimination of salmonid species in the cytochrome-B gene was used in conjunction with MiFish-U. The combined use of multiple primer pairs targeted for more than one gene marker (e.g., 12S-rRNA and cytochrome-B) is recommended to improve classification, reduce primer bias and false negatives, and better perform community-level analyses (Xiong et al. 2022). For eDNA samples in Table P-7, lack of Sequence Read Archive (SRA) accession numbers for MiFish-U and AKOncCytB primers indicates a lack of eDNA (species DNA is not present in the sample) or poor amplification of eDNA.

P.2.4.3 eDNA Results

The eDNA original sequence data has been deposited on the National Center for Biotechnology Information (NCBI) SRA, a public repository for sequence data. All reads are available under the standalone NCBI BioProject PRJNA1378815.

Table P-8, Table P-9, and Table P-10 summarize the presence of species detected by fish, mammal, and bird, respectively. While the primary analysis focused on fish species, mammal and bird DNA were also incidentally detected. It is important to note that while eDNA detections confirm a species' presence, they do not rule out the presence of other species, nor do they necessarily indicate abundance. However, when used in conjunction with data from other survey methods, such as beach seines and bottom trawls, this information is valuable for understanding the species in an area and the timing of their occurrence. Furthermore, eDNA allows for the detection of elusive species that are less likely to be captured by traditional survey techniques.

Table P-8. Fish detection results from eDNA analysis.

Scientific Name	Species	SITE 1: Deep Water Dock							SITE 2: Fishing Lagoon							SITE 3: Barge Basin							Note			
		9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	
<i>Ammodytes personatus</i>	Pacific Sand Lance			X	X								X	X						X						
<i>Anisarchus medius</i>	Stout Eelblenny	X						X																		
<i>Anoplopoma fimbria</i>	Sablefish							X									X									
<i>Artemius fenestralis</i>	Padded Sculpin				X	X				X																
<i>Asterias amurensis</i>	Northern Pacific Seastar							X									X									
Asteriidae Family	Seastar Family Species		X							X	X			X												
Bathymasteridae Family	Ronquil Family Species		X																							
<i>Beringraja binoculata</i>	Big Skate			X										X												
<i>Clinocottus acuticeps</i>	Sharpnose Sculpin										X															
<i>Clupea harengus</i>	Atlantic Herring																			X						This is likely a detection of <i>C. pallasii</i> .
<i>Clupea pallasii</i>	Pacific Herring	X	X	X	X	X			X	X	X	X	X	X			X	X	X	X	X	X				
<i>Clupea</i> sp.	Unidentified Herring Species	X	X	X	X	X				X	X	X						X	X	X	X	X				
Clupeidae Family	Herring Family Species																			X						
Cottidae Family	Sculpin Family Species																				X					
<i>Eleginus gracilis</i>	Saffron Cod					X								X									X			
Gadidae Family	Cod Family Species					X																				
<i>Gadus chalcogrammus</i>	Walleye Pollock (Alaska Pollock)					X			X								X		X							
<i>Gadus chalcogrammus</i> / <i>G. morhua</i>	Walleye Pollock (Alaska Pollock)/ Atlantic Cod	X																								<i>G. morhua</i> is the Atlantic species; the detection is likely <i>G. chalcogrammus</i> .
<i>Gadus macrocephalus</i> / <i>G. morhua</i>	Pacific Cod/ Atlantic Cod	X															X									<i>G. morhua</i> is the Atlantic species; the detection is likely <i>G. macrocephalus</i> .
<i>Gadus morhua</i>	Atlantic Cod																X									This is likely a detection of <i>G. chalcogrammus</i> .
<i>Gadus</i> sp.	Cod Species	X	X	X	X	X	X		X	X		X	X				X	X	X		X	X				
<i>Gasterosteus aculeatus</i>	Threespine Stickleback		X		X							X								X	X			X		
<i>Gymnocanthus intermedius</i> / <i>G. pistilliger</i>	Intermediate/ Threaded Sculpin																							X		
<i>Gymnocanthus pistilliger</i>	Threaded Sculpin				X									X							X		X			
<i>Hexagrammos lagocephalus</i>	Rock Greenling					X										X		X								
<i>Hexagrammos</i> sp.	Unidentified Greenling Species					X																				

Scientific Name	Species	SITE 1: Deep Water Dock							SITE 2: Fishing Lagoon							SITE 3: Barge Basin							Note		
<i>Pholis laeta</i>	Crescent Gunnel				X	X				X			X						X	X	X				
<i>Pholis ornata</i>	Saddleback Gunnel					X													X						
<i>Pholis</i> sp.	Unidentified Gunnel Species					X				X			X						X						
<i>Platichthys stellatus</i>	Starry Flounder																		X						
<i>Pleuronectes quadrituberculatus</i>	Alaska Plaice	X																							
Pleuronectidae Family	Righteye Flounder Family	X	X		X	X	X	X		X	X	X	X	X	X			X	X						
<i>Podothecus accipenserinus</i>	Sturgeon Poacher											X													
<i>Reinhardtius hippoglossoides</i>	Greenland Halibut																		X						
<i>Ronquilus jordani</i>	Northern Ronquil				X																				
<i>Salvelinus malma</i>	Dolly Varden																		X						
<i>Salvelinus</i> sp.	Unidentified Char Species																						X		
Stichaeidae Family	Prickleback Family Species	X																							
<i>Trichodon trichodon</i>	Pacific Sandfish					X	X												X						
		9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25

Table P-9. Incidental mammal detection results from eDNA analysis.

Scientific Name	Species	SITE 1: Deep Water Dock							SITE 2: Fishing Lagoon							SITE 3: Barge Basin							Note		
<i>Enhydra lutris</i>	Northern Sea Otter	X		X	X	X	X		X	X	X	X	X	X				X			X	X	X		
<i>Odocoileus hemionus</i>	Mule Deer																		X						Sitka black-tailed deer (<i>Odocoileus hemionus sitkensis</i>) is a subspecies of mule deer native to Alaska.
<i>Phoca vitulina</i>	Harbor Seal		X		X									X				X	X						
		9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25

Table P-10. Incidental bird detection results from eDNA analysis.

Scientific Name	Species	SITE 1: Deep Water Dock							SITE 2: Fishing Lagoon							SITE 3: Barge Basin							Note			
<i>Anas</i> sp.	Unidentified Dabbling Duck Species								X							X							X			
<i>Aythya marila</i>	Greater Scaup															X										
<i>Brachyramphus marmoratus</i>	Marbled Murrelet																						X			
<i>Bucephala albeola</i>	Bufflehead								X																	
<i>Clangula hyemalis</i>	Long-tailed Duck															X										
<i>Haliaeetus leucocephalus</i>	Bald Eagle								X																	
<i>Larus hyperboreus/ L. smithsonianus</i>	Glaucous Gull/ American Herring Gull													X				X						Likely a detection of <i>Larus</i> sp.		
<i>Larus smithsonianus</i>	American Herring Gull									X				X				X				X				
<i>Larus</i> sp.	Unidentified Gull Species				X	X								X				X								
<i>Melanitta deglandi</i>	White-winged Scoter			X																						
<i>Melanitta perspicillata</i>	Surf Scoter			X							X							X								
<i>Rissa tridactyla</i>	Black-legged Kittiwake								X																	
		9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	9-Apr-24	8-May-24	6-Jun-24	22-Jul-24	20-Aug-24	19-Sep-24	16-Jan-25	13-Feb-25	

The eDNA data provides a valuable snapshot of the marine biodiversity at these sites, revealing patterns of species distribution, seasonality, and highlighting the presence of key forage fish, salmon, and marine mammals. This eDNA analysis identified a diverse array of fish and several mammal species across three distinct sites designated for the study, Deep Water Dock, Fishing Lagoon, and Barge Basin, from April to September 2024 and January to February 2025.

Key summary findings from eDNA are as follows:

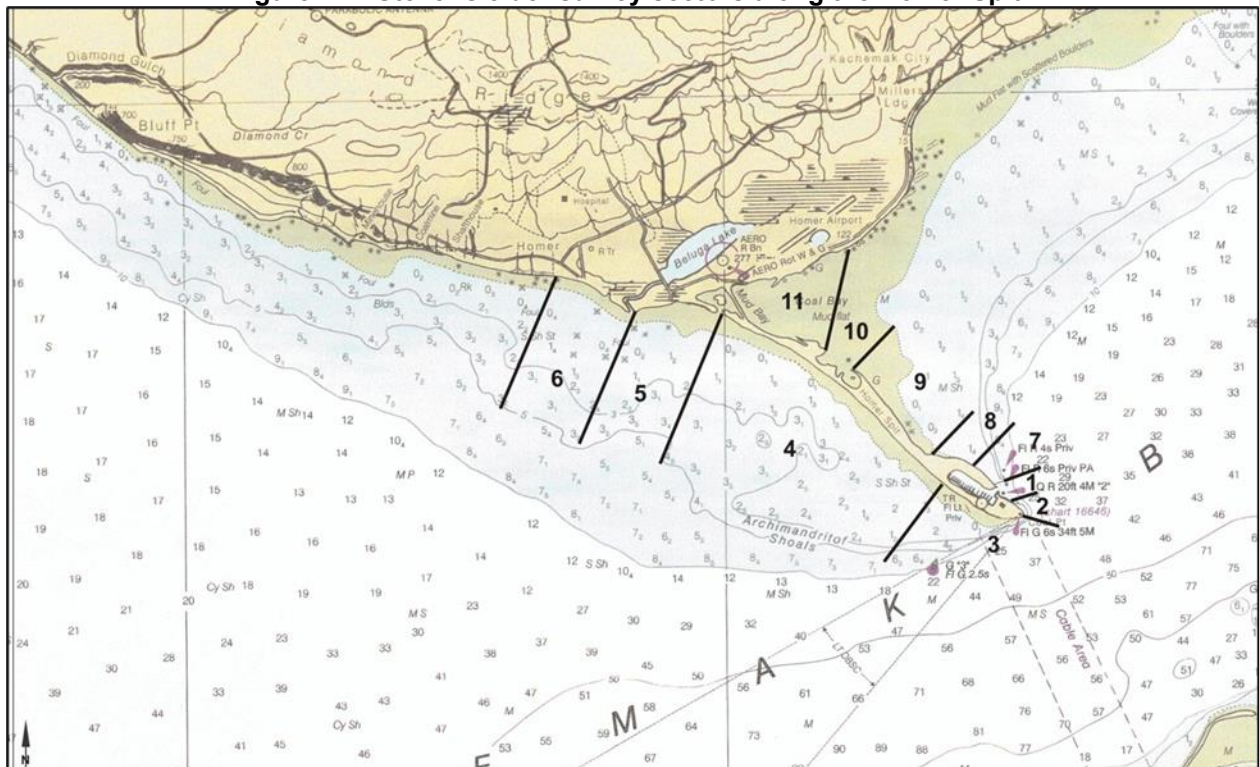
- **High Species Diversity:** A wide range of fin fishes were detected. Species from the herring (*Clupeidae*), cod (*Gadidae*), sculpin (*Cottidae*), and right-eye flounder (*Pleuronectidae*) families being particularly common. The Barge Basin (Site 3) appears to have the highest overall species richness.
- **Abundant Herring and Cod:** Pacific Herring (*Clupea pallasii*) and various cod species (*Gadus* spp.) were detected frequently across all three locations, suggesting they are common in the area.
- **Seasonal Salmon Presence:** Four species of Pacific Salmon, including Chinook (*Oncorhynchus tshawytscha*), Coho (*O. kisutch*), Pink (*O. gorbuscha*), and Sockeye (*O. nerka*), were detected, primarily during the summer and fall months (July–September), which is consistent with their seasonal migration and spawning patterns.
- **Marine Mammal Detections:** The incidental detection of Northern Sea Otters (*Enhydra lutris*) was consistent across most of the year, particularly at the Deep Water Dock and Fishing Lagoon. Harbor Seals (*Phoca vitulina*) were also detected, though less frequently.
- **Detection of Atlantic Species:** A notable pattern is the repeated detection of Atlantic species. These have been noted as likely detections of their closely related Pacific counterparts. This highlights a limitation in the specificity of the eDNA markers used for the analysis.
- **Anomalous Detection:** The detection of mule deer (*Odocoileus hemionus*) DNA at the Barge Basin has potential to be Sitka black-tailed deer (*Odocoileus hemionus sitkensis*), a subspecies of mule deer native to Alaska. Though Sitka black-tailed deer do not have an established population on the Kenai Peninsula, ADFG formally introduced this species to the area in October 2025. This detection could be due to human activities (i.e., processing a deer carcass near the harbor or while onboard a vessel). This practice is common in other areas of Alaska where this species is found (i.e., deboned deer carcasses are often disposed of in deeper waters near harbors in Southeast Alaska).

P.2.5 Steller's Eider Surveys

The Steller's eider (*Polysticta stelleri*) is an Arctic breeding sea duck that is the smallest and rarest of the four eider species. The Alaska-breeding population of Steller's eider was listed as threatened under the Endangered Species Act in 1997, due to concerns over apparent declines in numbers inferred from a reduction of nesting range in western Alaska (62 FR 31748).

The Steller's eider has historically wintered in a few ephemeral areas within Kachemak Bay and lower Cook Inlet. USACE conducted surveys in Kachemak Bay during the late winter and early spring of 2025 to determine the presence and relative location of Steller's eiders near the Homer Harbor and along the Homer Spit. The Homer Spit was broken up into 11 sectors for the surveys in 2025 (Figure P-7). These sectors match the sectors used in historical USACE Steller's eider surveys that occurred in the years 2003, 2004, and 2005. This created consistency between datasets, allowing for a general comparison in changes in Steller's eider abundance in Kachemak Bay across a span of 20 years. The boundaries of these survey sectors were originally selected given the presence of land-based features that were visible from offshore. This minimized potential overlap of species observed and mitigated for accidental recounting of individuals.

Figure P-7. Steller's eider survey sectors along the Homer Spit.



P.2.5.1 Survey Methodology

In 2025, Steller’s eider surveys were conducted both by land and by boat. Point count data was compiled for each survey date, with the number of species observed within each zone recorded along with the date and time. Survey dates include January 15–16, February 12–13, and March 25, in 2025. During each survey, species counts were recorded for each sector. Though the surveys focused on Steller’s eiders, the presence of other avian taxa (e.g., other sea ducks, grebes, and loons) were recorded. Surveys were conducted using a combination of 10x42 binoculars and a 20-60X spotting scope on a tripod. Species that could not be identified in the field were identified to the lowest taxon based on field markings presented in the *Birds of North America Field Guide* (National Geographic Society 1999).

P.2.5.2 Survey Results

Point counts for Steller’s eider for each sector from 2025, and data from historical USACE surveys in 2002–2004 for comparison, are reported in Table P-11. Given that each survey effort was conducted across one to three day(s), Steller’s eider point counts are combined here for simplicity. It is possible that some Steller’s eiders that were observed on the first survey day may have been the same individuals observed in the subsequent survey day(s) and therefore could have been a duplicate count. However, the focus of these surveys was to describe the general areas where this species is most commonly observed along the Homer Spit; not to quantify the patterns observed on the individual level.

Table P-11. Steller eider 2002/2003, 2003/2004, and 2024/2025 winter surveys.

Date	Map Sector										
	1	2	3	4	5	6	7	8	9	10	11
2002/2003 Winter Surveys											
2002, December 14-15	0	0	0	253	150	0	0	0	0	0	0
2003, January 15-16 ¹	0	0	0	596	90	1	0	0	0	0	0
2003, March 5-7	0	0	0	359	8	0	0	0	0	0	0
2004 Winter Surveys											
2004, January 15-17	0	0	0	16	0	0	0	0	0	0	0
2004, March 6-8	0	0	0	140	10	4	0	0	0	3	0
2025 Winter Surveys											
2025, January 15-16 ⁴	0	0	0	0	0	0	0	0	0	0	0
2025, February 12-13 ⁴	0	0	0	19	24	0	0	4	0	0	0
2025, March 25 ⁴	0	0	0	0	0	0	0	0	0	0	0
Sector Totals	0	0	0	1,383	282	5	0	4	0	3	0

Notes:

- ¹ Sector 5 and 6 not surveyed January 17, 2003, due to weather.
- ² Sectors 5 through 11 not surveyed January 16 and 17, 2004, due to ice.
- ³ Sectors 1 through 6 not surveyed March 7, 2004, due to rough seas.
- ⁴ Homer, Alaska, experienced a very mild winter in 2025 that has potential to have impacted the distribution and use of Kachemak Bay in 2025 than if conditions were harsher.

In general, Steller’s eiders were observed in the greatest numbers in Sector 4 and Sector 5. Sector 4 and Sector 5 are located in outer Kachemak Bay, which is outside of the project area near the Homer Harbor in Sector 7 and Sector 8. This trend is apparent

even given the 20-year gap in data. However, no Steller’s eiders were observed during the surveys in January 2025 or the single survey March of 2025. The winter of 2024–2025 was extremely mild across southcentral Alaska, with limited snow and ice cover. These environmental conditions may have altered the distribution of Steller’s eiders in Kachemak Bay and lower Cook Inlet.

P.2.6 Water Temperature and Salinity Data Collection

Conductivity, water temperature (°C) and salinity (Practical Salinity Scale [PSS]) were measured using a YSI Castaway CTD (a conductivity, temperature, and depth profiler) at the beach seine site near the Deep Water Dock during each monthly site visit from April through September 2024 after nearshore beach seining was complete. The beach seine site where CTD measurements were collected varied in depth, since sampling occurred immediately after the low tide once seine catch had been processed. Given the variation, the point measurement mode was used on the CTD. Point measurements are for surface waters and depth is not recorded in point measurement mode.

P.2.6.1 CTD Results

Measurements from the CTD profiler indicate that temperature varied between 7.1°C to 12.6°C during the summer months, while salinity varied between 20.5 PSS to 31.52 PSS (Table P-12). The measurement for the April 2024 site visit is not reported due to it being an invalid reading with no data collected. Two measurements were collected in September 2024 due to uncertainty with data collection on the CTD profiler.

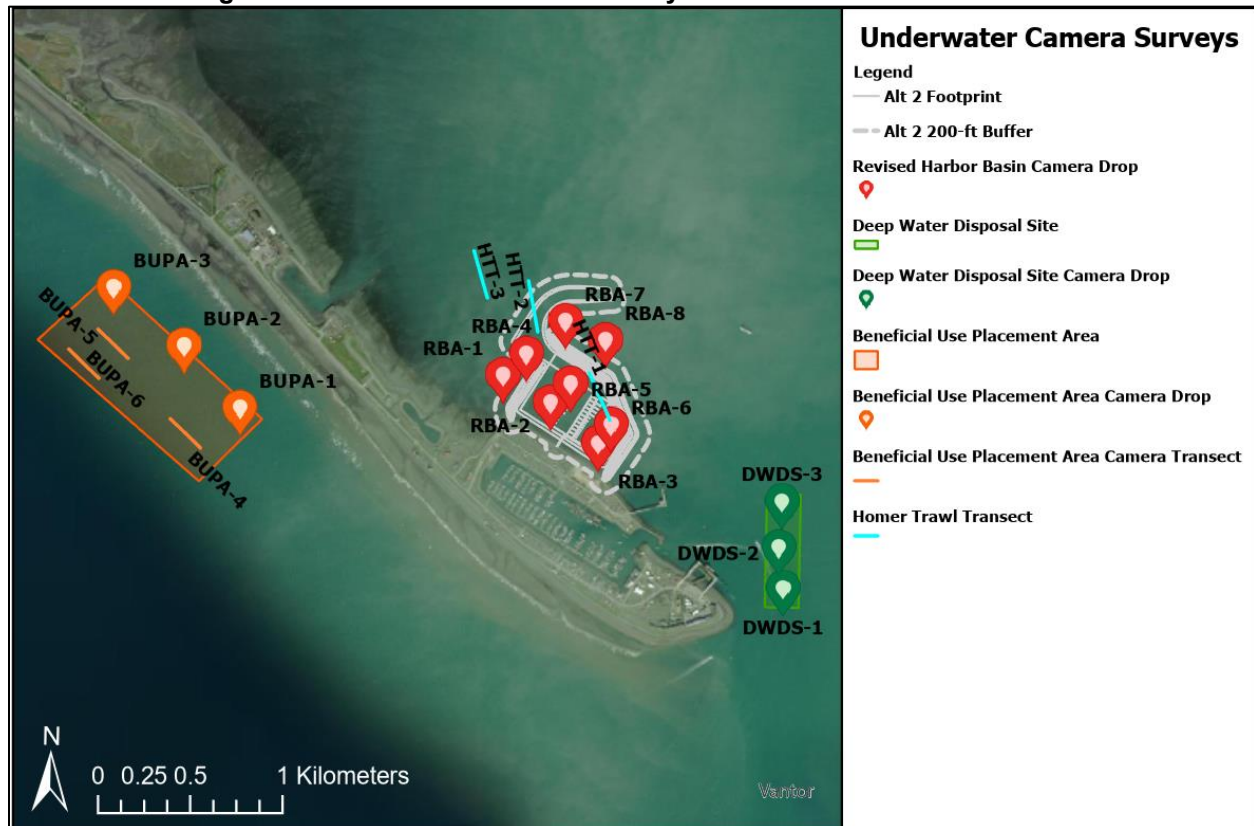
Table P-12. Monthly CTD point measurements from May through September 2024.

Date	Latitude (°N)	Longitude (°W)	Temperature (°C)	Conductivity (Microsiemens/Centimeter)	Salinity (PSS)
8-May-24	59.6113	-151.4305	7.1654	32254.1933	31.5283
6-Jun-24	59.6085	-151.4223	8.6921	31439.0097	29.3381
22-Jul-24	59.6089	-151.4227	12.6858	25226.3867	20.6430
20-Aug-24	59.6119	-151.4293	12.0080	24664.3320	20.5050
19-Sep-24	59.6113	-151.4283	10.8491	31482.0332	27.6757
	59.6112	-151.4285	10.8741	31192.0078	27.3754

P.2.7 Underwater Camera Surveys

To characterize the habitat in potential project areas, USACE conducted underwater surveys of the area in and around the project footprint and potential dredged material discharge sites newly under consideration for this feasibility study. Underwater video surveys were conducted concurrent bottom trawl efforts on June 6, 2024, and in the Proposed Project Footprint, Deep Water Disposal Site (DWDS), and Beneficial Use Placement Area (BUPA) on April 10, 2025.

Figure P-8. Underwater camera surveys conducted in 2024 and 2025.



P.2.7.1 Survey Methodology

Underwater camera surveys were completed using a variety of equipment and methods.

Compact Action Camera

The Homer Trawl Transect footage was recorded June 6, 2024, and provided preliminary data for characterizing the benthic habitat and environmental resources in the deeper water portions of the anticipated Proposed Project Footprint. Homer Trawl Transect underwater camera surveys were conducted by zip-tying a compact action camera encased in waterproof housing onto the center of headline of the bottom trawl netting. The captured footage entailed the full extent of each 5-minute bottom trawl transect.

Benthic Drop Camera

A *Group B Benthic GPH-1750* camera was deployed from the Homer Harbor’s tug on April 10, 2025, and lowered to the bottom of the water column to record benthic footage at designated survey points within the Proposed Project Footprint, DWDS, and BUPA. Videos were approximately 5-minutes in duration for each survey point. The array was deployed from the side of the vessel and held steady manually. The vessel was allowed to drift naturally.

USACE Alaska District developed a benthic camera array to capture a high-definition video of the benthic floor, to enhance the characterization of the habitat and environmental resources within project areas of interest. The array consists of an adjustable frame, flashlight, and camera. It is portable, flexible, and resilient. The flashlight and camera are enclosed in 3-inch aluminum housings and equipped with lithium-ion batteries. The camera records high-definition video on an internal 128 Gigabyte Secure Digital card. The camera and light are both rated to over 5,000-feet deep, can be synchronized to capture time-lapse video, and have sufficient battery life for long-term deployments. The flashlight contains two high output Cree light-emitting diodes providing 3,500 lumens. It includes an 18-mode programmable timer and allows the diodes to be disconnected if necessary to extend battery life.

The frame was constructed from galvanized and stainless steel and can be broken down for transport in about 5 minutes. The lengths of the upper and lower bridles are configured to maintain appropriate geometry for consistently capturing quality video. The camera and light are angled slightly towards each other and at a downward angle to illuminate the seafloor about 3 feet in front of the cannonball, which anchors the system on the bottom. A hardball buoy is tied into the mainline above the bridle.

Live Feed Shallow Water Drop Camera

This camera set-up was deployed from the Homer Harbor's tug on April 10, 2025. USACE intended to utilize a *SplashCam Delta Vision High-Definition* camera to record footage in the BUPA and Proposed Project Footprint but, there were battery issues that lead to only three transects in the BUPA being completed. Other planned transects were converted to drop camera survey points. The camera was deployed manually from the Homer Harbor's tug and lowered to the bottom of the water column. USACE adjusted the camera's depth based on the live-feed footage. Due to the sea state, it was difficult to maintain consistent and good quality visibility of the seafloor.

P.2.7.2 Survey Results

Species identification beyond the genus level was problematic due to inconsistent lighting, viewing angle, dimensional perspective, and other factors of camera footage acquired during underwater camera surveys. Timing is also a consideration that must be accounted for when using the footage to describe the habitat in the areas of interest. For example, April is not an ideal season for delineating the presence of eelgrass. The predominate species composition and habitat features (e.g., sediment) observed will be described by each area of interest in Table P-13.

Table P-13. Underwater camera survey observation results.

Area of Interest	Underwater Camera Observations			
	Water	Sediment	Vegetation	Animals
Homer Trawl Transect (HTT-1 to HTT-3) Figure P-9	The water was visually turbid.	Predominately consisted of material composed of sand and silt with shell hash and fragments throughout site.	Observed algae, seagrass (i.e., eelgrass), seaweed (e.g., sea lettuce), and kelp (e.g., sugar kelp). Vegetation density varied throughout site.	Anemones (e.g., plumose anemone), sea star species (e.g., purple sea star), sea pens, groundfish (e.g., sculpin and flatfish), and crabs (e.g., hermit and tanner).
Proposed Project Footprint (RBA-1 to RBA-8) Figure P-10	There was a green hue to the water likely due to microscopic algae and/or phytoplankton. Visual turbidity may have been masked by green hue.	Predominately consisted of sand and silt. Cobbles were observed within the sediment at RBA-2 and RBA-3. Shell hash and fragments observed.	Observed algae, seagrass (i.e., eelgrass), seaweed (e.g., sea lettuce), and kelp (e.g., sugar kelp). Vegetation density varied throughout site. A rock with coralline encrusted algae was observed at HTT-3.	Anemones (e.g., plumose anemone), sea stars (e.g., purple sea star), and sea pens.
Deep Water Disposal Site (DWDS-1 to DWDS-3) Figure P-11	The water was visually turbid.	Predominately consisted of sandy material throughout site. Shell hash was most evident in sediment at DWDS-1.	Established vegetation appeared absent, but seaweed debris/remnants were observed.	Anemones (e.g., plumose anemone) sea stars, crabs, and groundfish (e.g., starry flounder).
Beneficial Use Placement Area (BUPA-1 to BUPA-3) Figure P-12	There was a green hue to the water likely due to microscopic algae and/or phytoplankton. Visual turbidity may have been masked by green hue.	Predominately consisted of patches of and intermixed areas of sand and cobble with shell hash and fragments throughout site.	Observed algae and unidentified species of seaweed and kelp. The density of this vegetation varied throughout site.	Sea stars.

Note: The content included in this table is solely based on the footage captured during underwater camera surveys. Solely by itself, the observations should not be used as to assume (1) the potential density, abundance, or exclusion of a species at the subject site or (2) the general turbidity and hue of the water, which is subjected to other factors like tide and season.

Figure P-9. Homer Trawl Transect underwater camera screen captures from June 6, 2024.

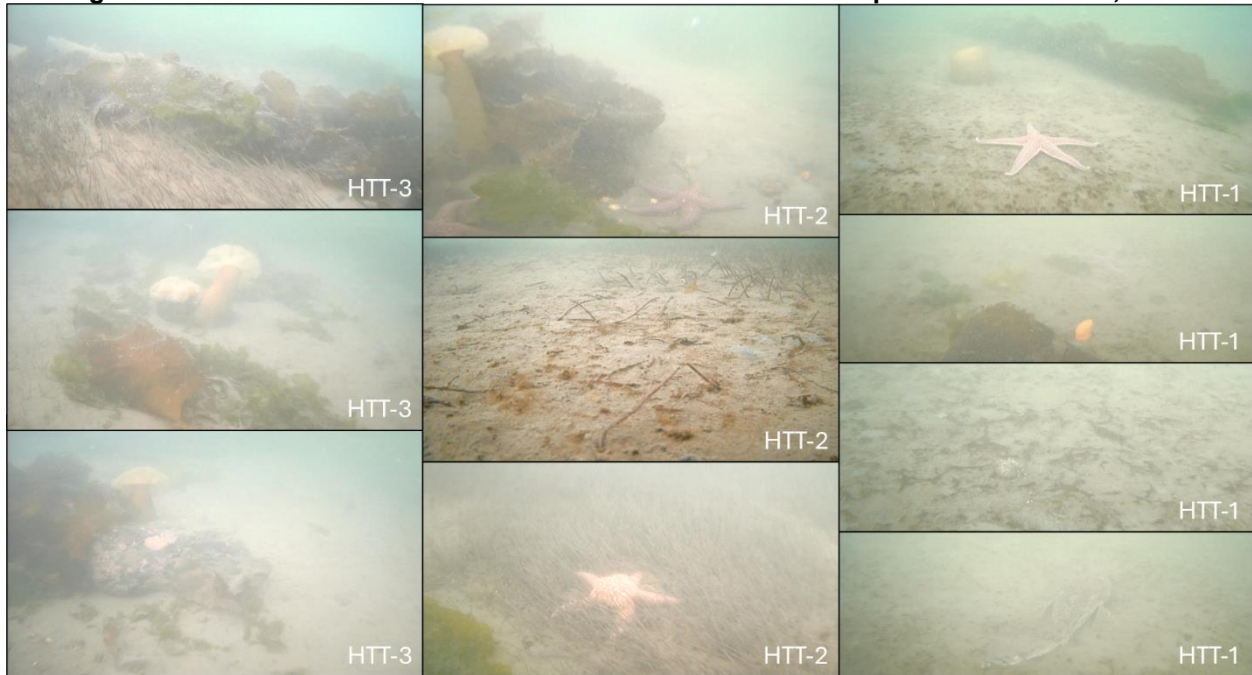


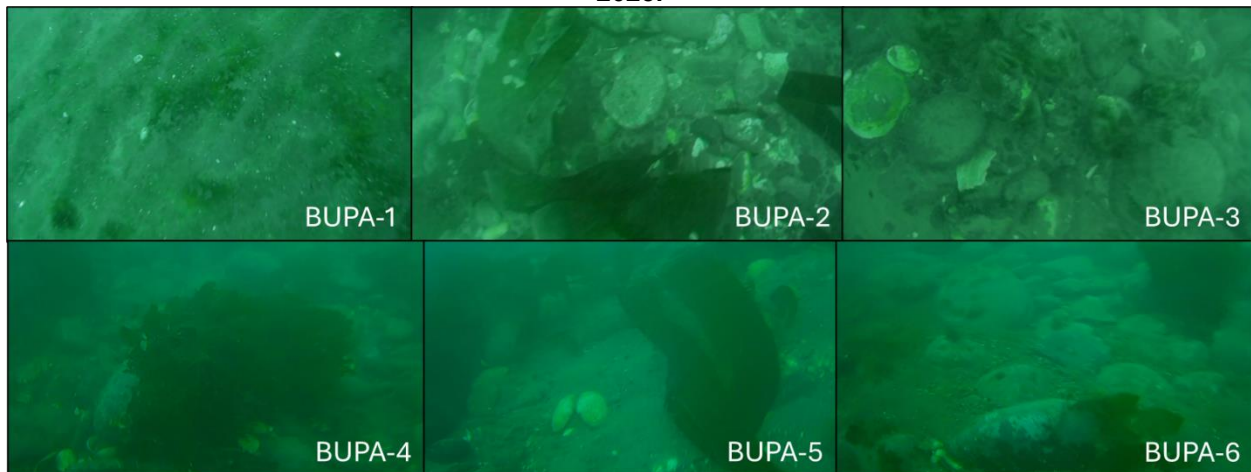
Figure P-10. Proposed Project Footprint underwater camera screen captures from April 10, 2025.



Figure P-11. Deep Water Disposal Site underwater camera screen captures from April 10, 2025.



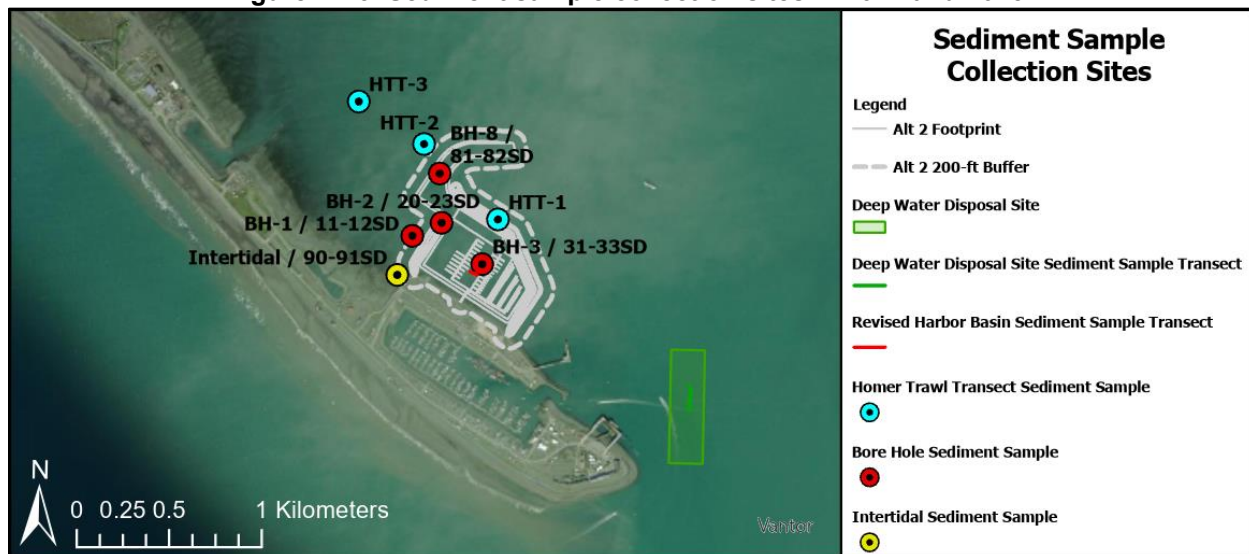
Figure P-12. Beneficial Use Placement Area underwater camera screen captures from April 10, 2025.



P.2.8 Sediment Sample Collection and Analysis

Sediment samples were collected to provide preliminary means to inform the type of surface material within the (1) Proposed Project Footprint on June 6, 2024, and April 10, 2025, and October 7-11, 2025, and (2) the DWDS on April 10, 2025 (Figure P-13). USACE Biologists predominately collected surface sediment samples. However, both surface and subsurface samples were collected during the geotechnical investigation for the study from October 7-11, 2025, to assess material characteristics to inform project design and proper management dredged material.

Figure P-13. Sediment sample collection sites in 2024 and 2025.



P.2.8.1 Sample Methodology

The Sampling and Analysis Plan and fieldnotes (Attachment 2) detail the collection and analysis methodology for samples from the geotechnical investigation for chemical analysis as summarized in Table P-14. Physical composition was not analyzed in the lab, as the data that would be available from the Geotechnical Data Report (S&W 2026) was deemed sufficient for preliminary characterization. For complete details on the geotechnical methodology and its results, refer to the Geotechnical Data Report. This section will now focus on the grab sample collection method and visual characterization of other samples collected during fieldwork.

Table P-14. Sediment samples collect during geotechnical investigations in October 2025.

Sampling Point	Mudline Elevation (Feet MLLW)	Sample ID 26HOM-CW__	Sample Interval (Feet below Mudline)	Sampled Interval (Feet MLLW)	Date (MM/DD/YY)	Time (24-hour)	Notes
BH-1	-7	11SD	0 – 2	-7 to -9	10/11/25	1028	
		12SD	15 - 17	-22 to -24	10/11/25	1050	
BH-2	-9	20SD	0 - 2	-9 to -11	10/8/25	2200	Field Duplicate of -21SD
		21SD	0 - 2	-9 to -11	10/8/25	2215	
		22SD	9 - 10	-18 to -19	10/9/25	0020	-
		23SD	15 - 17	-24 to -26	10/9/25	0040	-
BH-3	-10.5	31SD	0 - 2	-10.5 to -12.5	10/10/25	2013	
		32SD	5.5 - 6	-16 to -16.5	10/10/25	2106	
		33SD	9.5 - 10	-20 to -20.5	10/10/25	2115	-
BH_8	-10	81SD	0 - 2	-10 to -12	10/7/25	1515	
			17.8 - 19.8	-27.8 to -29.8	10/7/25	1615	
Intertidal (-151.431111, 59.609722)	~+0.3	90SD	0 – 0.5	-0.3 to -0.8	10/9/25	1130	Field Duplicate of -91SD
		91SD	0 – 0.5	-0.3 to -0.8	10/9/25	1140	

Sediment grab samples were collected with a stainless-steel barrel sampler. The barrel sampler was custom fabricated to perform surface sediment collection in very deep water. It is approximately 6 inches in diameter and 30 inches long with an inverted cone-shaped collar on the open end to help collect material as the sampler is dragged along the bottom. An adequately long section of crab pot line for the desired scope within the context of the depth was tied into the chain bridle. The barrel sampler was deployed from the Homer Harbor's tug and lowered to the seafloor at the sample locations. The barrel sampler was then towed in an arc until the operator determined that an adequate volume of material was collected, then the sampler was retrieved with tugboat winch. Once the sampler reached the surface, the excess water was decanted and the contents of the sampler was dumped into a plastic barrel for characterization.

The Proposed Project Footprint and DWDS sediment value, hue, and chroma was determined using the 2009 Year Revised version of the Munsell Soil Color Book (Munsell Color 2015):

- Hue: indicates the relation of the sediment's color to red, yellow, green, blue, and purple.
- Value: Indicates lightness of the sediment's color.
- Chroma: indicates the strength of the sediment's color.

All other characterization was judged by field observation.

P.2.8.2 Sample Results

The findings in the Geotechnical Data Report (S&W 2026) support the visual field characterizations previously conducted by USACE biologists (Table P-15; Figure P-14, and Figure P-15). The geotechnical analysis confirmed that the material observed on the surface is consistent with the sediment layer targeted for dredging. This layer, which encompasses the dredge prism, greatest depth design to -26 feet MLLW and -28 feet MLLW with a 2-foot overdepth. The layer consists primarily of fines (e.g., silt).

Table P-15. Sediment grab sample observed characteristics.

Area of Interest	Value, Chroma, and Hue	Texture	Other Notable Characteristics
Homer Trawl Transect (HTT-1 to HTT-3) Figure P-14	Did not compare sediment with Munsell Soil Color Book.	HTT-1: Silt HTT-2: Silty with sand HTT-3: Silty sand	HTT-1: Dark grey color. HTT-2: Dark grey color with trace of shell hash. HTT-3: Dark grey color with trace of shell hash.
Proposed Project Footprint Figure P-14	Hue: GLEY 1 Value: 2.5 Chroma: N	Silt with sand	Traces of shell hash and tube building worms throughout matrix.
Deep Water Disposal Site Figure P-15	Hue: 5Y Value: 2.5 Chroma: 2	Sandy silt	Trace organic material (cellulose) and black grain (potentially due to coal).

Source: (Munsell Color 2015)

Note: The content included in this table is solely based on visual observation during fieldwork to provide a preliminary understanding of the characteristics of the surface material within project areas of interest prior to the completion of geotechnical and chemical analyses.

Figure P-14. Proposed Project Footprint (April 10, 2025) and Homer Trawl Transect (June 6, 2024) sediment grab samples.

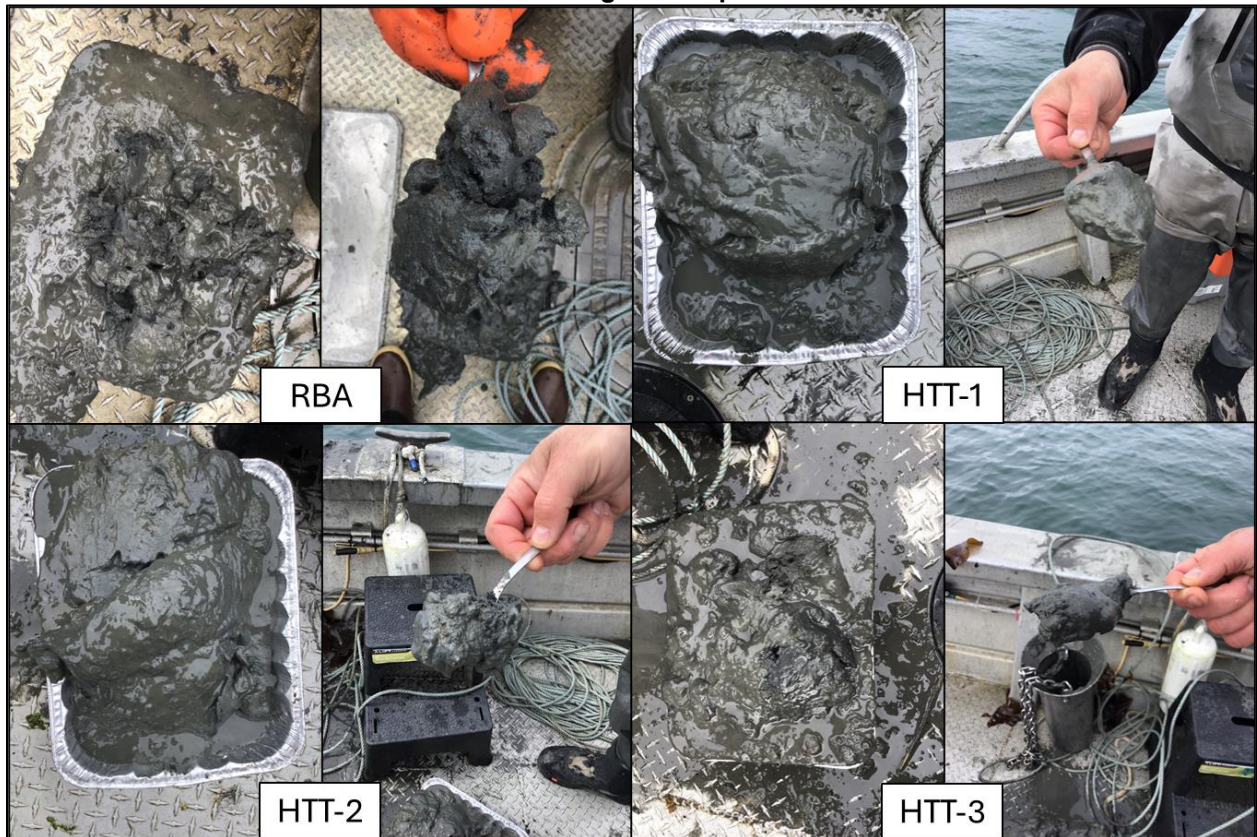
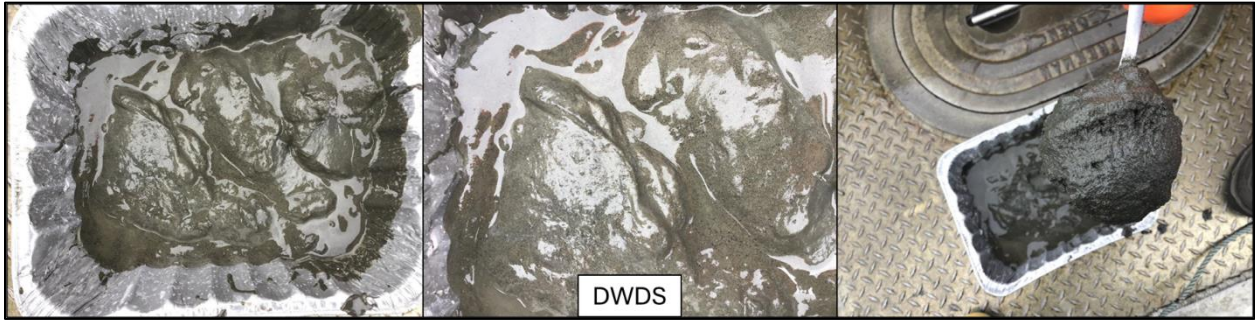


Figure P-15. Deep Water Disposal Site (April 10, 2025) sediment grab sample.



Based on the chemical data quality review (Attachment 2), the dredged material from Homer Harbor is suitable for unconfined aquatic placement. The analysis confirmed that no contaminants exceeded the Dredged Material Management Program marine screening levels. Although levels of naturally occurring arsenic and an isolated detection of naphthalene surpassed the more conservative Alaska Department of Environmental Conservation (ADEC) cleanup levels for informing potential upland placement of dredged material, the overall data quality was deemed acceptable.

P.2.9 Intertidal Narrative Transect

An intertidal narrative transect was conducted on May 9, 2024, to characterize the intertidal habitat based on visual observation. The transect was in the original potential project footprint but now lies northwest of the Proposed Project Footprint (Figure P-16).

Figure P-16. Transect for intertidal habitat narration recorded on May 9, 2024.



P.2.9.1 Survey Methodology

Surveyor measuring tape was laid from the high tide line out towards the waterline in 100-foot increments on May 9, 2024, starting at 9:02 AM and ending at 9:45 AM. Low tide was scheduled to occur at approximately 10:05 AM. A cellular device was used to capture visual and audio narration by a USACE biologist for each 100-foot increment,

while another USACE biologist recorded additional observations and notes and took georeferenced photos on a GPS tablet.

P.2.9.2 Survey Results

Table P-16 summarizes the narrated observations of USACE biologists, and Figure P-17 depicts the transect for the narration in relation to the Homer Spit. This information will support characterization of the intertidal zone in and around the Proposed Project Footprint.

Table P-16. Intertidal observation summary by 100-foot increment.

Transect Increment	Description of Habitat and Resources
High Tide Line– 100 Feet	Surface sediment consisted of large gravel and small cobble. There was some organic debris at the high tide line, and the sediment was dry to about 12 feet. Between the high tide line to about 26 feet there was no established vegetation observed. The steepest down slope occurs between high tide to 26 feet. After 26 feet, more sand exposed in surface sediment consisting of small and large and small cobble. After 40 feet the area flattened and stayed relatively flat with a minimal slope. Increment included scattered large rocks and sand patches. The first large rock encounter along the transect occurred at 62 feet. The first sand patch along the transect occurred at 73 feet.
100–200 Feet	Surface sediment consisted of large gravel and medium sized cobble on top of sand. Sand patches and scour pools were scattered throughout increment. Scour pools likely held invertebrates during low tide. This increment was a low energy beach due to minimal slope gradient. At about 125 feet, small periwinkles (<i>Littorina littorea</i>), sediment embedded small blue mussels (<i>Mytilus edulis</i>), and very small barnacles (<i>balanus glandula</i>) on rocks were observed.
200–300 Feet	Similar surface sediment and intertidal life observed. However, at 235 feet the density and size of blue mussels increased. A small scour pool encountered along the transect was observed with green algae and observed brown algae in a scour pool further away. Sand patches continue to be observed along transect as well. At 251 feet, seagrass (eelgrass [<i>Zostera marina</i>]) was observed in a scour pool with algae.
300–400 Feet	Surface sediment remained similar. Blue mussels continued to increase in density along transect. At 328 feet a slight relief area was encountered with brown algae and sea lettuce (<i>Ulva</i> sp.). Small, scattered patches of sea lettuce appeared to account for approximately 5% coverage in increment. At 380 feet an outflow area of slow flowing water was encountered. In the outflow area, there was a dense eelgrass bed with ribbon kelp (<i>Alaria marginata</i>) and brown algae. The higher areas were absent of seagrass and kelp but occupied by blue mussels.
400–500 Feet	Surface sediment remained similar. The eelgrass bed that begun at 380 feet ended at 468 feet. Blue mussels observed continue to increase in size and density further along the transect. This is presumed partly due to the area being less exposed during low tide. The observation of very small barnacles was noted again.
500–600 Feet	Surface sediment remained similar until approximately midway through the increment. At 533 feet, in addition to the sediment that had been occurring in prior increments, a silt component was observed for the first time along the transect. Silt was observed covering algae and blue mussels. Silt layer was more apparent starting at 585 feet. At 518 feet there was a small patch of green algae. Blue mussels were also observed less embedded and established more on top of sediment. A dense blue mussel patch extended to around 590 feet, beyond which the presence of blue muscles was absent.

Transect Increment	Description of Habitat and Resources
600–700 Feet	Surface sediment consists of large gravel and small cobble on a silty sediment layer. There were large rocks scattered throughout increment. Blue mussels continued to be absent. Sea lettuce patches and filamentous green algae (<i>Spirogyra</i> sp.) were observed. The first sea star observed along the transect was at 648 feet and continued to be observed in increment. Ribbon kelp and green, brown, and red algae were present. At 687 feet, larger ribbon kelp attached to rocks were observed. First encounter of rockweed (<i>Fucus distichus</i>) at 697 feet.
700–800 Feet	Continue to observe similar surface sediment and life as that observed in prior increment. At 742 feet, there were more areas of standing water (approximately 1–2 inches deep). The primary vegetation was ribbon kelp and sea stars were frequently sighted. At 783 feet, old snail egg casings were observed.
800–900 Feet	Surface sediment consisted of a silt layer with embedded large gravel and small to medium cobble with the occasional small rock. Water in increment was flowing slowly otop of sediment. Snail egg casing continued to be observed and. There were longer stretches of ribbon kelp and larger bladed kelp that began in this increment.
900–1,000 Feet ¹	Surface sediment was similar to prior increment but silt content was notably increasing as the distance from the high tide line increased. Scattered rocks and boulders were observed throughout area. The density of kelp was high and relatively uniform. Kelp consisted predominately of wider bladed kelp than prior increments. Some holes were sighted in increments that may have indicated otter digging and potential sea star predation. At 992 feet patchy eelgrass in low density and abundance was observed.
1,000–1,100 Feet ¹	Surface sediment remained similar to prior increment with increasing silt content and scattered rocks and boulders. Empty clamshells and snail egg casings were observed. At 1,029 feet, small polychaete patches began to occur.
1,100–1,200 Feet ¹	At 1,105 feet there was standing water. Surface sediment became very silty and life observed relatively remained consistent between 900 to 1,200 feet. At 1,185 feet, plumose anemones (<i>Metridium farcimen</i>) were observed.
Observing beyond 1,200 Feet ¹	Surface sediment was very silty. Vegetation began to visibility become less dense and water was ankle deep. Water clarity began to deteriorate due to the outgoing tide suspending the silty sediment. Anemone density was increasing, and a rock was observed with coralline encrusted algae.

¹ USACE biologists identified species of aquatic vegetation observed after the narration was completed. The species included: sugar kelp (*Saccharina latissimi*), stiff red ribbon kelp (*Nereocystis luetkeana*), filament green algae, ribbon kelp, eelgrass, and sea lettuce.

Figure P-17. Intertidal environmental captured on May 9, 2024.



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Attachment 1: Aquatic Resource Permit Documentation

Note: Only the consolidated data from the Aquatic Resource Permits is included.

This sheet provides raw data collected over the term of the permits.

ADF&G Permit Number: CF-24-005 and CF-25-029

Date	Location	Latitude	Longitude	Collection Method	Species Common Name	Species Scientific Name	Life Stage	Size (mm)	Number	Disposition	Comments
09-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Original: 59.6099, -151.4294)	59.6099	-151.4294	beach seine	sharpnose sculpin	<i>Clinocottus acuticeps</i>	adult	47	1	ID'd, measured, and released	Site will not be reset due to presence of boulders.
09-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Original: 59.6097, -151.4332)	59.6097	-151.4332	beach seine	capelin	<i>Mallotus villosus</i>	juvenile	130	1	ID'd, measured, and released	
09-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Original: 59.6097, -151.4332)	59.6097	-151.4332	beach seine	great sculpin	<i>Myoxocephalus polyacanthocephalus</i>	juvenile	90	1	ID'd, measured, and released	
09-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Original: 59.6097, -151.4332)	59.6097	-151.4332	beach seine	Pacific staghorn sculpin	<i>Leptocottus armatus</i>	adult	250	1	ID'd, measured, and released	
09-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 3 - Barge Basin (Original: 59.6203, -151.4541)	59.6203	-151.4541	beach seine	N/A	N/A	N/A	N/A	N/A	N/A	No catch. Site will be aborted due to depth, rapidly incoming tide, shallow gradient, and presence of boulders that hindered activity.
10-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	59.6110	-151.4211	bottom trawl	unidentified arthropod	N/A	N.D.	N.D.	1	released	
10-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	59.6152	-151.4276	bottom trawl	unidentified arthropod	N/A	N.D.	N.D.	4	released	
10-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	59.6152	-151.4276	bottom trawl	unidentified shrimp	N/A	N.D.	N.D.	2	released	
10-Apr-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	unidentified crab	N/A	N.D.	10	1	measured and released	Size represents the carapace width.
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	59.6090	-151.4229	beach seine	graceful decorator crab	<i>Oregonia gracilis</i>	N.D.	Various	6	ID'd, measured, and released	Some lost on beach. Sizes approximately between 10 - 20 mm carapace.
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	59.6090	-151.4229	beach seine	kelp greenling	<i>Hexagrammos decagrammus</i>	juvenile	193	1	ID'd, measured, and released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	59.6090	-151.4229	beach seine	rock greenling	<i>Hexagrammos lagocephalus</i>	YOY	55	1	ID'd, measured, and released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	crescent gunnel	<i>Pholis laeta</i>	juvenile	N.D.	1	ID'd and released	Cannot measure accurately when alive.
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	helmet crab	<i>Telmessus cheiragonus</i>	N.D.	65	1	ID'd, measured, and released	Size represents the carapace width.
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	rock sole	<i>Lepidopsetta bilineata</i>	juvenile	154	1	ID'd, measured, and released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	rock sole	<i>Lepidopsetta bilineata</i>	juvenile	102	1	ID'd, measured, and released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	whitespotted greenling	<i>Hexagrammos stelleri</i>	juvenile	200	1	ID'd, measured, and released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	whitespotted greenling	<i>Hexagrammos stelleri</i>	juvenile	169	1	ID'd, measured, and released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 3 - Barge Basin (Modified: 59.6173, -151.4379)	59.6173	-151.4379	beach seine	N/A	N/A	N/A	N/A	N/A	N/A	No catch. Site was aborted due to depth, rapidly incoming tide, shallow gradient, and presence of boulders that hindered activity.
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	59.6110	-151.4211	bottom trawl	yellowfin sole	<i>Limanda aspera</i>	adult	320	2	ID'd, measured, and released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	59.6152	-151.4276	bottom trawl	helmet crab	<i>Telmessus cheiragonus</i>	N.D.	50	1	ID'd, measured, and released	Size represents the carapace width.
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	59.6152	-151.4276	bottom trawl	unidentified crab	N/A	N.D.	N.D.	5	released	Sizes approximately around 10 mm carapace.
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	59.6152	-151.4276	bottom trawl	unidentified shrimp	N/A	N.D.	N.D.	3	released	
08-May-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	rock sole	<i>Lepidopsetta bilineata</i>	adult	365	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	59.6090	-151.4229	beach seine	kelp greenling	<i>Hexagrammos decagrammus</i>	juvenile	155	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	59.6090	-151.4229	beach seine	rock greenling	<i>Hexagrammos lagocephalus</i>	YOY	54	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	chinook salmon	<i>Oncorhynchus tshawytscha</i>	juvenile	155	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	crescent gunnel	<i>Pholis laeta</i>	juvenile	N.D.	1	ID'd and released	Cannot measure accurately when alive.
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	graceful decorator crab	<i>Oregonia gracilis</i>	N.D.	N.D.	1	released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	great sculpin	<i>Myoxocephalus polyacanthocephalus</i>	juvenile	110	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	helmet crab	<i>Telmessus cheiragonus</i>	N.D.	90	1	ID'd, measured, and released	Size represents the carapace width.
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	helmet crab	<i>Telmessus cheiragonus</i>	N.D.	60	1	ID'd, measured, and released	Size represents the carapace width.
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	helmet crab	<i>Telmessus cheiragonus</i>	N.D.	50	1	ID'd, measured, and released	Size represents the carapace width.
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	helmet crab	<i>Telmessus cheiragonus</i>	N.D.	30	2	ID'd, measured, and released	Size represents the carapace width.
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	kelp greenling	<i>Hexagrammos decagrammus</i>	juvenile	150	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	kelp greenling	<i>Hexagrammos decagrammus</i>	juvenile	145	2	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	kelp greenling	<i>Hexagrammos decagrammus</i>	juvenile	137	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	kelp greenling	<i>Hexagrammos decagrammus</i>	juvenile	60	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	rock greenling	<i>Hexagrammos lagocephalus</i>	YOY	67	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	rock greenling	<i>Hexagrammos lagocephalus</i>	YOY	65	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	rock greenling	<i>Hexagrammos lagocephalus</i>	YOY	62	1	ID'd, measured, and released	
06-Jun-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	59.6113	-151.4302	beach seine	rock greenling	<i>Hexagrammos lagocephalus</i>	YOY	61	1	ID'd, measured, and released	

20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	59.6167	-151.4327	bottom trawl	unidentified greenling	N/A	N.D.	91	1	measured and released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	59.6152	-151.4276	bottom trawl	unidentified snailfish	N/A	N.D.	50	1	measured and released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	lyre crab	<i>Hyas coarctatus</i>	N.D.	N.D.	1	released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	rock sole	<i>Lepidopsetta bilineata</i>	adult	342	1	ID'd, measured, and released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	saffron cod	<i>Eleginus gracilis</i>	juvenile	72	1	ID'd, measured, and released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	saffron cod	<i>Eleginus gracilis</i>	juvenile	69	1	ID'd, measured, and released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	silverspotted sculpin	<i>Blepsias cirrhosus</i>	juvenile	65	1	ID'd, measured, and released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	unidentified snailfish	N/A	N.D.	71	1	measured and released
20-Aug-24	US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	59.6167	-151.4327	bottom trawl	unidentified snailfish	N/A	N.D.	43	1	measured and released

This sheet provides the total numbers of individuals per species by location, collection method, and over the term of the permits.

Location				Collection Method	Species Common Name	Species Scientific Name	Number of Individuals by Location	Percentage of Total Individuals of Species at Location	Number of Total Individuals of Species by Collection Method	Percentage of Total Individuals of Species by Collection Method	Fieldwork Total Number of Individuals
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	Arctic shanny	<i>Stichaeus punctatus</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	buffalo sculpin	<i>Enophrys bison</i>	2	100%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Original; 59.6097, -151.4332)	Beach Seine	capelin	<i>Mallotus villosus</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	chinook salmon	<i>Oncorhynchus tshawytscha</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	crested gunnel	<i>Pholis laeta</i>	9	53%	17	100%	17	100%	17	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	crested gunnel	<i>Pholis laeta</i>	8	47%	17	100%	17	100%	17	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	graceful decorator crab	<i>Oregonia gracilis</i>	9	32%	22	36%	28	36%	28	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	graceful decorator crab	<i>Oregonia gracilis</i>	1	4%	10	36%	28	36%	28	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	graceful decorator crab	<i>Oregonia gracilis</i>	3	11%	18	64%	28	64%	28	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	graceful decorator crab	<i>Oregonia gracilis</i>	10	36%	18	64%	28	64%	28	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	graceful decorator crab	<i>Oregonia gracilis</i>	5	18%	18	64%	28	64%	28	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	great sculpin	<i>Myoxocephalus polyacanthocephalus</i>	12	55%	22	100%	22	100%	22	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	great sculpin	<i>Myoxocephalus polyacanthocephalus</i>	9	41%	22	100%	22	100%	22	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Original; 59.6097, -151.4332)	Beach Seine	great sculpin	<i>Myoxocephalus polyacanthocephalus</i>	1	5%	22	100%	22	100%	22	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	helmet crab	<i>Telmessus cheiragonus</i>	5	29%	14	82%	17	82%	17	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	helmet crab	<i>Telmessus cheiragonus</i>	9	53%	14	82%	17	82%	17	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	helmet crab	<i>Telmessus cheiragonus</i>	1	6%	3	18%	17	18%	17	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	helmet crab	<i>Telmessus cheiragonus</i>	1	6%	3	18%	17	18%	17	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	helmet crab	<i>Telmessus cheiragonus</i>	1	6%	3	18%	17	18%	17	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	hermit crab	<i>Paguroidea sp.</i>	2	100%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	kelp greenling	<i>Hexagrammos decagrammus</i>	10	37%	27	100%	27	100%	27	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	kelp greenling	<i>Hexagrammos decagrammus</i>	17	63%	27	100%	27	100%	27	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	lyre crab	<i>Hyas coarctatus</i>	1	33%	3	100%	3	100%	3	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	lyre crab	<i>Hyas coarctatus</i>	1	33%	3	100%	3	100%	3	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	lyre crab	<i>Hyas coarctatus</i>	1	33%	3	100%	3	100%	3	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	masked greenling	<i>Hexagrammos octogrammus</i>	28	49%	57	100%	57	100%	57	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	masked greenling	<i>Hexagrammos octogrammus</i>	29	51%	57	100%	57	100%	57	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 3 - Barge Basin (Modified; 59.6173, -151.4379)	Beach Seine	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 3 - Barge Basin (Original; 59.6203, -151.4541)	Beach Seine	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	Pacific cod	<i>Gadus macrocephalus</i>	3	100%	3	100%	3	100%	3	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	Pacific halibut	<i>Hippoglossus stenolepis</i>	1	20%	5	100%	5	100%	5	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	Pacific halibut	<i>Hippoglossus stenolepis</i>	4	80%	5	100%	5	100%	5	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	Pacific herring	<i>Clupea pallasii</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	Pacific staghorn sculpin	<i>Leptocottus armatus</i>	1	50%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Original; 59.6097, -151.4332)	Beach Seine	Pacific staghorn sculpin	<i>Leptocottus armatus</i>	1	50%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	padded sculpin	<i>Arctidius fenestralis</i>	2	100%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	purple sea star	<i>Pisaster ochraceus</i>	3	60%	5	100%	5	100%	5	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	purple sea star	<i>Pisaster ochraceus</i>	1	20%	5	100%	5	100%	5	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	purple sea star	<i>Pisaster ochraceus</i>	1	20%	5	100%	5	100%	5	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	ribbon snailfish	<i>Liparis cyclopus</i>	1	50%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	ribbon snailfish	<i>Liparis cyclopus</i>	1	50%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	rock greenling	<i>Hexagrammos lagocephalus</i>	8	44%	17	94%	18	94%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	rock greenling	<i>Hexagrammos lagocephalus</i>	9	50%	17	94%	18	94%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	rock greenling	<i>Hexagrammos lagocephalus</i>	1	6%	1	6%	18	6%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	rock sole	<i>Lepidopsetta bilineata</i>	2	8%	12	48%	25	48%	25	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	rock sole	<i>Lepidopsetta bilineata</i>	10	40%	12	48%	25	48%	25	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	rock sole	<i>Lepidopsetta bilineata</i>	4	16%	13	52%	25	52%	25	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	rock sole	<i>Lepidopsetta bilineata</i>	6	24%	13	52%	25	52%	25	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	rock sole	<i>Lepidopsetta bilineata</i>	3	12%	13	52%	25	52%	25	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	saffron cod	<i>Eleginus gracilis</i>	2	18%	5	45%	11	45%	11	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	saffron cod	<i>Eleginus gracilis</i>	3	27%	5	45%	11	45%	11	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	saffron cod	<i>Eleginus gracilis</i>	1	9%	6	55%	11	55%	11	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	saffron cod	<i>Eleginus gracilis</i>	2	18%	6	55%	11	55%	11	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	saffron cod	<i>Eleginus gracilis</i>	3	27%	6	55%	11	55%	11	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	sea pen	<i>Ptilosarcus gurneyi</i>	21	84%	25	100%	25	100%	25	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	sea pen	<i>Ptilosarcus gurneyi</i>	4	16%	25	100%	25	100%	25	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Original; 59.6099, -151.4294)	Beach Seine	sharpnose sculpin	<i>Clinocottus acuticeps</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)	Beach Seine	silverspotted sculpin	<i>Blepsias cirrhosus</i>	8	44%	16	89%	18	89%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	silverspotted sculpin	<i>Blepsias cirrhosus</i>	8	44%	16	89%	18	89%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	silverspotted sculpin	<i>Blepsias cirrhosus</i>	1	6%	2	11%	18	11%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	silverspotted sculpin	<i>Blepsias cirrhosus</i>	1	6%	2	11%	18	11%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	silverspotted sculpin	<i>Blepsias cirrhosus</i>	1	6%	2	11%	18	11%	18	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	snake prickleback	<i>Lumpenus sagitta</i>	33	87%	33	87%	38	87%	38	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	snake prickleback	<i>Lumpenus sagitta</i>	4	11%	5	13%	38	13%	38	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	snake prickleback	<i>Lumpenus sagitta</i>	1	3%	5	13%	38	3%	38	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	starry flounder	<i>Platichthys stellatus</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	sturgeon poacher	<i>Podothtes accipenserinus</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	surf smelt	<i>Hypomesus pretiosus</i>	4	100%	4	100%	4	100%	4	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	tanner crab	<i>Chionoecetes bairdi</i>	1	50%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	tanner crab	<i>Chionoecetes bairdi</i>	1	50%	2	100%	2	100%	2	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	three-spine stickleback	<i>Gasterosteus aculeatus</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)	Beach Seine	tubenose poacher	<i>Pallasina barbata</i>	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	unidentified aneome	N/A	1	100%	1	100%	1	100%	1	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	unidentified arthropod	N/A	1	17%	6	100%	6	100%	6	
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	unidentified arthropod	N/A	5	83%	6	100%	6	100%	6	

US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	unidentified crab	N/A	11	92%	12	100%	12
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	unidentified crab	N/A	1	8%	12	100%	12
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	Beach Seine	unidentified flatfish	N/A	12	75%	12	75%	16
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	unidentified flatfish	N/A	1	6%	4	25%	16
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	unidentified flatfish	N/A	1	6%	4	25%	16
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	unidentified flatfish	N/A	2	13%	4	25%	16
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	unidentified greenling	N/A	1	100%	1	100%	1
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	Beach Seine	unidentified sculpin	N/A	1	6%	14	88%	16
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	Beach Seine	unidentified sculpin	N/A	13	81%	14	88%	16
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	unidentified sculpin	N/A	2	13%	2	13%	16
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	unidentified sea star	N/A	3	100%	3	100%	3
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	Beach Seine	unidentified shrimp	N/A	31	25%	34	28%	122
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	Beach Seine	unidentified shrimp	N/A	3	2%	34	28%	122
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	unidentified shrimp	N/A	54	44%	88	72%	122
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	unidentified shrimp	N/A	34	28%	88	72%	122
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	Beach Seine	unidentified snailfish	N/A	1	17%	3	50%	6
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	Beach Seine	unidentified snailfish	N/A	2	33%	3	50%	6
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)	Bottom Trawl	unidentified snailfish	N/A	1	17%	3	50%	6
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)	Bottom Trawl	unidentified snailfish	N/A	2	33%	3	50%	6
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified: 59.6090, -151.4229)	Beach Seine	whitespotted greenling	<i>Hexagrammos stelleri</i>	1	25%	4	100%	4
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified: 59.6113, -151.4302)	Beach Seine	whitespotted greenling	<i>Hexagrammos stelleri</i>	3	75%	4	100%	4
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)	Bottom Trawl	yellowfin sole	<i>Limanda aspera</i>	2	100%	2	100%	2

This sheet provides comparison of number of individuals by species collected over the term of the permits through each the used collection methods.

Species Common Name	Species Scientific Name	Collection Method	Number of Total Individuals of Species by Collection Method	Percentage of Total Individuals of Species by Collection Method	Fieldwork Total Number of Individuals
Arctic shanny	<i>Stichaeus punctatus</i>	Beach Seine	1	100%	1
buffalo sculpin	<i>Enophrys bison</i>	Beach Seine	2	100%	2
capelin	<i>Mallotus villosus</i>	Beach Seine	1	100%	1
chinook salmon	<i>Oncorhynchus tshawytscha</i>	Beach Seine	1	100%	1
crescent gunnel	<i>Pholis laeta</i>	Beach Seine	17	100%	17
graceful decorator crab	<i>Oregonia gracilis</i>	Beach Seine	10	36%	28
graceful decorator crab	<i>Oregonia gracilis</i>	Bottom Trawl	18	64%	28
great sculpin	<i>Myoxocephalus polyacanthocephalus</i>	Beach Seine	22	100%	22
helmet crab	<i>Telmessus cheiragonus</i>	Beach Seine	14	82%	17
helmet crab	<i>Telmessus cheiragonus</i>	Bottom Trawl	3	18%	17
hermit crab	<i>Paguroidea</i> sp.	Bottom Trawl	2	100%	2
kelp greenling	<i>Hexagrammos decagrammus</i>	Beach Seine	27	100%	27
lyre crab	<i>Hyas coarctatus</i>	Bottom Trawl	3	100%	3
masked greenling	<i>Hexagrammos octogrammus</i>	Beach Seine	57	100%	57
Pacific cod	<i>Gadus macrocephalus</i>	Bottom Trawl	3	100%	3
Pacific halibut	<i>Hippoglossus stenolepis</i>	Beach Seine	5	100%	5
Pacific herring	<i>Clupea pallasii</i>	Bottom Trawl	1	100%	1
Pacific staghorn sculpin	<i>Leptocottus armatus</i>	Beach Seine	2	100%	2
padded sculpin	<i>Artedius fenestralis</i>	Beach Seine	2	100%	2
purple sea star	<i>Pisaster ochraceus</i>	Bottom Trawl	5	100%	5
ribbon snailfish	<i>Liparis cyclopus</i>	Beach Seine	2	100%	2
rock greenling	<i>Hexagrammos lagocephalus</i>	Beach Seine	17	94%	18
rock greenling	<i>Hexagrammos lagocephalus</i>	Bottom Trawl	1	6%	18
rock sole	<i>Lepidopsetta bilineata</i>	Beach Seine	12	48%	25
rock sole	<i>Lepidopsetta bilineata</i>	Bottom Trawl	13	52%	25
saffron cod	<i>Eleginus gracilis</i>	Beach Seine	5	45%	11
saffron cod	<i>Eleginus gracilis</i>	Bottom Trawl	6	55%	11
sea pen	<i>Ptilosarcus gurneyi</i>	Bottom Trawl	25	100%	25
sharpnose sculpin	<i>Clinocottus acuticeps</i>	Beach Seine	1	100%	1
silverspotted sculpin	<i>Blepsias cirrhosus</i>	Beach Seine	16	89%	18
silverspotted sculpin	<i>Blepsias cirrhosus</i>	Bottom Trawl	2	11%	18
snake prickleback	<i>Lumpenus sagitta</i>	Beach Seine	33	87%	38
snake prickleback	<i>Lumpenus sagitta</i>	Bottom Trawl	5	13%	38
starry flounder	<i>Platichthys stellatus</i>	Bottom Trawl	1	100%	1
sturgeon poacher	<i>Podothecus accipenserinus</i>	Beach Seine	1	100%	1
surf smelt	<i>Hypomesus pretiosus</i>	Beach Seine	4	100%	4
tanner crab	<i>Chionoecetes bairdi</i>	Bottom Trawl	2	100%	2
three-spine stickleback	<i>Gasterosteus aculeatus</i>	Beach Seine	1	100%	1
tubenose poacher	<i>Pallasina barbata</i>	Beach Seine	1	100%	1
unidentified aneome	N/A	Bottom Trawl	1	100%	1
unidentified arthropod	N/A	Bottom Trawl	6	100%	6
unidentified crab	N/A	Bottom Trawl	12	100%	12
unidentified flatfish	N/A	Beach Seine	12	75%	16
unidentified flatfish	N/A	Bottom Trawl	4	25%	16
unidentified greenling	N/A	Bottom Trawl	1	100%	1
unidentified sculpin	N/A	Beach Seine	14	88%	16
unidentified sculpin	N/A	Bottom Trawl	2	13%	16
unidentified sea star	N/A	Bottom Trawl	3	100%	3
unidentified shrimp	N/A	Beach Seine	34	28%	122
unidentified shrimp	N/A	Bottom Trawl	88	72%	122
unidentified snailfish	N/A	Beach Seine	3	50%	6
unidentified snailfish	N/A	Bottom Trawl	3	50%	6
whitespotted greenling	<i>Hexagrammos stelleri</i>	Beach Seine	4	100%	4
yellowfin sole	<i>Limanda aspera</i>	Bottom Trawl	2	100%	2

This sheet provides the number of individuals by species collected over the term of the permits.

Species Common Name	Species Scientific Name	Fieldwork Total Number of Individuals
Arctic shanny	<i>Stichaeus punctatus</i>	1
buffalo sculpin	<i>Enophrys bison</i>	2
capelin	<i>Mallotus villosus</i>	1
chinook salmon	<i>Oncorhynchus tshawytscha</i>	1
crescent gunnel	<i>Pholis laeta</i>	17
graceful decorator crab	<i>Oregonia gracilis</i>	28
great sculpin	<i>Myoxocephalus polyacanthocephalus</i>	22
helmet crab	<i>Telmessus cheiragonus</i>	17
hermit crab	<i>Paguroidea</i> sp.	2
kelp greenling	<i>Hexagrammos decagrammus</i>	27
lyre crab	<i>Hyas coarctatus</i>	3
masked greenling	<i>Hexagrammos octogrammus</i>	57
Pacific cod	<i>Gadus macrocephalus</i>	3
Pacific halibut	<i>Hippoglossus stenolepis</i>	5
Pacific herring	<i>Clupea pallasii</i>	1
Pacific staghorn sculpin	<i>Leptocottus armatus</i>	2
padded sculpin	<i>Arteidius fenestralis</i>	2
purple sea star	<i>Pisaster ochraceus</i>	5
ribbon snailfish	<i>Liparis cyclopus</i>	2
rock greenling	<i>Hexagrammos lagocephalus</i>	18
rock sole	<i>Lepidopsetta bilineata</i>	25
saffron cod	<i>Eleginus gracilis</i>	11
sea pen	<i>Ptilosarcus gurneyi</i>	25
sharpnose sculpin	<i>Clinocottus acuticeps</i>	1
silverspotted sculpin	<i>Blepsias cirrhosus</i>	18
snake prickleback	<i>Lumpenus sagitta</i>	38
starry flounder	<i>Platichthys stellatus</i>	1
sturgeon poacher	<i>Podothecus accipenserinus</i>	1
surf smelt	<i>Hypomesus pretiosus</i>	4
tanner crab	<i>Chionoecetes bairdi</i>	2
three-spine stickleback	<i>Gasterosteus aculeatus</i>	1
tubenose poacher	<i>Pallasina barbata</i>	1
unidentified anneome	N/A	1
unidentified arthropod	N/A	6
unidentified crab	N/A	12
unidentified flatfish	N/A	16
unidentified greenling	N/A	1
unidentified sculpin	N/A	16
unidentified sea star	N/A	3
unidentified shrimp	N/A	122
unidentified snailfish	N/A	6
whitespotted greenling	<i>Hexagrammos stelleri</i>	4
yellowfin sole	<i>Limanda aspera</i>	2
Total Number of Individuals Observed		533
Total Number of Species Observed	(excludes unidentified species)	34

Location

US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 1 (Start at 59.6121, -151.4221 to Stop at 59.6098, -151.4200)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 2 (Start at 59.6165, -151.4281 to Stop at 59.6140, -151.4271)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Bottom Trawl Transect 3 (Start at 59.6179, -151.4334 to Stop at 59.6156, -151.4320)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Original; 59.6099, -151.4294)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 1 - Deep Water Dock (Modified; 59.6090, -151.4229)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Original; 59.6097, -151.4332)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 2 - Fishing Lagoon (Modified; 59.6113, -151.4302)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 3 - Barge Basin (Original; 59.6203, -151.4541)
US: Alaska, Kachemak Bay, Eastern Shore of the Homer Spit, Beach Seine Site 3 - Barge Basin (Modified; 59.6173, -151.4379)

Attachment 2:
Sampling Analysis Plan and Results



**US Army Corps
of Engineers**

Alaska District

Sampling and Analysis Plan

Homer Navigation Improvements Feasibility Study Dredged Material Characterization, Homer, Alaska



September 2025

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Sampling and Analysis Plan

Homer Navigation Improvements Feasibility Study Dredged Material Characterization,
Homer, Alaska

1. Introduction

This sampling and analysis plan (SAP) has been prepared by the Environmental Resources Section, Civil Works Project Management Branch of the Alaska District US Army Corps of Engineers (CEPOA-PMC-E). This plan outlines the procedures to be followed during a chemical and physical evaluation of existing sediments within the proposed Homer Navigation Improvements dredging footprint.

2. Project Background

2.1. Location

Homer Harbor is part of The City of Homer and located on the north shore of Kachemak Bay on the southwestern edge of the Kenai Peninsula in Alaska (Figure 1, Figure 2). It is 227 miles by road south of Anchorage at the southernmost point of the Sterling Highway at approximately 59° 38' north latitude and 151° 33' west longitude. Homer is in the Homer Recording District and hosts a population of 5,522 as of 2020. It is accessible via air, road, and water year-round and is the economic center of the southern Kenai Peninsula. It is the southernmost town on Alaska's contiguous highway system and part of the Alaska Marine Highway, a ferry service that operates along the south-central coast of the state. The Port of Homer is located at the end of the Homer Spit (Figure 3), a narrow promontory of land demarcating Kachemak Bay from Cook Inlet.

2.2. Purpose

The Alaska District is investigating navigation improvements that would provide safe, reliable and efficient waterborne transportation systems and moorage to support Homer's current and future fleet. This study is utilizing the authority of Section 2006 of WRDA 2007, Remote and Subsistence Harbors, as amended. Section 2006 states:

The Secretary [of the Army] may recommend a project without the need to demonstrate that the project is justified solely by national economic development benefits if [...] the project would be located in the State of Hawaii or Alaska [...] and over 80 percent of the goods transported through the harbor would be consumed within the United States.

The implementation of any of the structural alternatives under consideration would include dredging and dredged material management to create adequate depths for the

design vessel. Dredging would excavate material down to -26' mean lower low water (MLLW) for the entrance channel, -24' MLLW for the fairway, and -20' MLLW or -15' MLLW for the mooring area. USACE must characterize the dredged material to be excavated and managed to complete the cost estimate and environmental analysis needed for the feasibility study. The feasibility study currently assumes the dredged material is suitable for unconfined aquatic placement as the least costly environmentally acceptable management option

2.3. Past Sampling

Two key historical studies/data include:

- NOAA, 2009, Sediment Quality Triad Assessment in Kachemak Bay: Characterization of Soft Bottom Benthic Habitats and Contaminant Bioeffects Assessment
- Historical Homer Harbor Maintenance Dredging Sampling (most recent sampling events were conducted by USACE in 2019).

Past sampling activities have identified concentrations of metals, polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), and butyl tin exceeding applicable screening levels.

Most recently (spring 2019 sampling efforts) harbor samples containing diesel range organics (DRO), selenium and arsenic were all detected at concentrations exceeding ADEC Soil Cleanup screening levels. DRO and selenium results appeared to be artifacts of non-sediment materials being present in the sediment and analyzed by the lab (coal and brass respectively). Arsenic (a naturally occurring metal) was present in all samples at concentrations exceeding ADEC criteria. However, all results were within natural background considering they are similar to the concentrations of other samples throughout Kachemak Bay. There is potential that selenium/arsenic levels are impacted by potential presence of rare earth metals (note: no data indicating such) if the method for analysis included use of inductively Coupled Plasma Mass Spectrometry (ICP-MS). The 2019 method of testing metal concentrations were the same methods used to test metal concentrations for Dillingham Small Boat Harbor in 2022 sampling efforts, which reflected heightened selenium concentrations due to rare earth metals. This should be considered when reviewing results of analysis for selenium.

In-water placement for beach nourishment was evaluated as well in 2019 and results were compared to screening levels in the Sediment Evaluation Framework for the Pacific Northwest (SEF). Zinc was the only analyte detected at a concentration above the screening level. The presence of this metal is attributed to the brass that was likely present in the sample and is not representative of metals contamination. It was recommended that if in water placement became the preferred alternative, additional

sampling for ammonia, total volatile solids and tributyltin should be performed during the fall sampling event to characterize the Z-layer.

In 2019 fall sampling efforts, samples showed no indication that analytes were detected above SEF screening criteria in Z-layer samples taken. There were exceedances of ADEC levels, and except for Arsenic levels, other exceedances were flagged and attributed to false positive/contamination of samples. The arsenic levels were consistent with those throughout Kachemak Bay and assessed naturally occurring. There is also a lack of arsenic-source(s). Ammonia, total volatile solids and TBT analysis were conducted in this sampling effort.

3. Project Organization

The parties responsible for performing each of the following sequence of steps required to successfully complete this investigation are summarized as follows:

- The Homer Navigation Improvements Project Delivery Team (PDT) determined that characterization of the dredged material is necessary to evaluate the parameters and impacts of dredged material management.
- The Environmental Resources Section (CEPOA-PMC-E) is responsible for the development of a Sampling and Analysis Plan (SAP) in accordance with the draft Alaska Dredged Material Evaluation Framework (ADMEDF). The Dredged Materials Evaluation and Disposal Procedures User Manual (USACE 2021), commonly referred to simply as the “User Manual”, was also considered during the preparation of this SAP.
- All laboratories will submit data directly to the Environmental Engineering (CEPOA-ECE) chemist who is responsible for assuring that all results are received within the normal turnaround time period and that all aspects of quality control are properly performed.
- The Environmental Resources section will review the concentrations of CoCs in the dredged material and assimilate the data into the alternative development prior to finalization of the Integrated Feasibility Report.

4. Sampling and Analysis Strategy

4.1. Purpose of the Sampling

The purpose of sampling is to determine if the sediments to be dredged are suitable for unconfined aquatic placement. The Alaska District applies the draft Alaska Dredged Material Evaluation Framework (ADMEDF), which is based on the Dredged Material Evaluation and Disposal Procedures (DMEDP) “User Manual”, to the evaluation of dredging projects with in-water placement options (DMMO, 2021).

The proposed Homer Port Expansion is adjacent to the existing Homer Small Boat Harbor and proximal to the Homer Marine Repair and Salvage Facility. There are many existing sources of potential anthropogenic contamination in the area and the proposed dredged material is believed to contain a significant volume of silt, which is likely to bind to contaminants. The Port Expansion would be constructed on the leeward side of the Homer Spit, so it would be protected from large waves originating in the Gulf of Alaska or Lower Cook Inlet. There is no basis for considering an exemption from testing. The existing Homer Harbor is ranked “Moderate” in accordance with the ADMEF and the Port Expansion sediment is also tentatively ranked Moderate based on the lack of sediment quality data, proximity to sources, grain size, and low level of energy exposure.

The sampling activities conducted during the feasibility study are constrained by the limitations of the geotechnical investigation and so are not intended to conform with the sampling intensity standards of the ADMEF. The intent of the feasibility-level investigation is to reduce risk and provide information for the next phase of the study, which is expected to include a more robust geotechnical investigation and allow for an ADMEF-compliant sampling program.

The testing of Homer Port Expansion dredged material will be limited to samples collected from a maximum of four borehole locations and one intertidal location. USACE will make every practical effort to adjust the borehole locations to appropriately characterize the dredged material in the turning basin and entrance channel by attempting to site two boreholes in each general navigation feature, but geotechnical data collection requirements and environmental conditions such as wave energy and shallow water may cause deviations.

4.2. Scope of Sample Collection

Samples should be collected from four borehole locations within the dredged features (entrance channel and turning basin), plus a surface sample from the intertidal area near the marine salvage and repair facility, for a combined total of up to five locations. Preliminary determinations regarding the relevant borehole locations to be sampled indicate boreholes 1, 2, 3, and 8 would include the dredged material associated with the current array of alternatives and would provide the most valuable data. The relevant borehole locations and preliminary sampling stations are shown in Figure 4 and Table 1. The final decision regarding which borehole locations will be chemically sampled will be made prior to mobilization based on the current understanding of where each borehole is located with respect to the general navigation features under consideration at the time.

The samples will be analyzed discretely (i.e., not composite sampled) and samples will only be collected from sediments within the project depth. Three samples may be

collected from each of the four boreholes; one representing the surface material and up to two representing differing types of buried material; i.e., if the sediment profile above the project depth consists of a layer of silt over layers of sand and clay, a sample should be collected from each of the three layers above project depth.

The field crew will determine the appropriate depth to sample based on the observed physical characteristics at the time of sample collection. In general, sediment samples for chemical analysis should be collected from each distinct sediment formation above the project depth greater than one foot thick. Sediment is sand, silt or clay settled or deposited under a water body. Sand is material between 62.5 and 2,000 microns, so material greater than 2,000 microns is not considered sediment for the purpose of this sampling and analysis plan. Gravel, cobble, boulder, breccia, bedrock, etc are not required to be sampled. An example scenario where the mudline is six feet below mean lower low water (-6' MLLW) follows and is visualized in Figure 5:

- a. Sand layer two feet thick. Collect sample for chemical analysis because the layer is sediment and greater than one foot thick.
- b. Boulder layer four feet thick. Do not sample because the matrix of this layer is mostly rock. Sand in the interstices is presumably associated with the overlying stratum.
- c. Sand layer less than one foot thick. Do not sample because the layer is less than one foot thick.
- d. Breccia layer five feet thick. Do not sample because the layer is consolidated and mostly rock.
- e. Sand layer three feet thick. Collect sample because the layer is sediment and greater than one foot thick.

The decision to collect a sample must be made in the field prior to the availability of laboratory grain size data. Best professional judgement will be used to determine the composition of the strata in the field; i.e., whether a stratum is composed of sediment or rock. If the field crew decides not to collect a sample from a given stratum, the rationale for that decision must be documented in the field notes. If the field crew is in doubt regarding whether a sample should be collected from a stratum, the crew should either:

- a. Contact the USACE before proceeding or,
- b. Collect the sample and notify the USACE of the uncertainty by email within 24-hours. USACE POCs will decide whether the sample should be analyzed or discarded.

4.3. Scope of Analytical Methods

Each sample will be analyzed for the following parameters, using the recommended analytical methods listed in table 8-3, and Section 8.3 of the ADMEF. If the

recommended analytical methods are not used, a statement explaining the rationale for deviation should be provided by the entity responsible for the deviation.

4.3.1. Recommended Analytical Methods for Standard COCs (ADMEF Table 8-3)

- Metals, total:
 - Antimony (EPA 6010/6020)
 - Arsenic (EPA 6010/6020)
 - Cadmium (EPA 6010/6020)
 - Chromium (EPA 6010/6020)
 - Copper (EPA 6010/6020)
 - Lead (EPA 6010/6020)
 - Silver (EPA 6010/6020)
 - Zinc (EPA 6010/6020)
 - Selenium (EPA 6020/7740/7742)
 - Mercury (EPA 7471/6020Mod)
- Polyaromatic Hydrocarbons (PAHs) (EPA 8270)
- Chlorinated Hydrocarbons:
 - 1,2-Dichlorobenzene (EPA 8560/8270)
 - 1,4-Dichlorobenzene (EPA 8560/8270)
 - 1,2,4-Trichlorobenzene (EPA 8560/8270)
 - Hexachlorobutadiene (EPA 8270/8081)
- Phthalates (EPA 8270)
- Phenols (EPA 8270)
- Miscellaneous Extractables:
 - Benzyl alcohol (EPA 8270)
 - Benzoic acid (EPA 8270)
 - Dibenzofuran ((EPA 8270)
 - N-Nitrosophenylamine (EPA 8270)
 - Hexachlorobutadiene (EPA 8270/8081)
- Pesticides (EPA 8081)
- PCB Aroclors (EPA 8082)
- Petroleum Hydrocarbons
 - Gasoline range organics (AK101)
 - Diesel range organics (AK102)
 - Residual range organics (AK103)

4.3.2. Recommended Analytical Methods for Special Contaminants of Concern
(ADMEF Section 8.3)

- Tributyltin (PSEP 1997)

5. Sediment Sampling

5.1. Sampling Equipment

The major equipment required to accomplish this sampling effort are listed below:

- nitrile gloves
- sample containers
- stainless steel spoons
- zip top plastic bags
- Liquinox soap and buckets
- paper towels
- brushes

5.2. Sampling Procedures

Sampling of the project sediments will be accomplished using the following procedure:

- Water depth and the real-time tidal stage will be measured at the time of sampling at each station. Water depth will be measured to the nearest 1/10th foot.
- Sediment will be collected using the geotechnical coring device.
- The split spoon will be opened and the sediment or soil core will be photographed with a measuring stick or tape for scale. The measuring device will be placed alongside the core sample, measuring down from the top of the core.
- The following protocol will be followed to collect analytical samples from the core:
 1. Determine whether the core sample contains material meeting the chemical sampling criteria:
 - a. If the core sampler does contain material meeting the criteria for chemical sample collection, note the rationale for electing not to collect a sample and proceed to the next core sample.
 - b. If the core sampler does contain material meeting the chemical sampling criteria, describe the rationale for electing to collect a sample and move to step 2
 2. Fill two lab provided sample jars from sediment core using stainless steel spoon, noting:

- a. Name/Initials of collector
 - b. Sample Number
 - c. Collected Date and Time
 - d. Analytical Method
 - e. Matrix
 - f. Preservative
 - g. Horizontal location
3. Place primary sample in cooler.
 4. Fill one septimated lab provided sample jar from sediment core for GRO analysis using stainless steel spoon, noting:
 - a. Name/Initials of collector
 - b. Sample Number
 - c. Collected Date and Time
 - d. Analytical Method
 - e. Matrix
 - f. Preservative
 - g. Horizontal location
 5. Preserve sample with methanol.
 6. Place sample in cooler.
 7. Proceed to next vertical core sample section above the project depth for the respective navigation feature (entrance channel or turning basin) and repeat steps 1-6.
 8. Proceed to the next borehole location within the dredging limits and repeat steps 1-7.
- Collect QA duplicates and matrix spike duplicates from sample locations with adequate volumes of sediment for all required analyses, including geotechnical analyses.
 - Immediately after filling, the sample containers will be placed into a cooler with sufficient ice to maintain the samples at 0-6°C.
 - Once the samples have been collected and properly stored, the sampler will be washed with water from the site until no more sediment adheres to the device.
 - A rinsate blank will not be collected unless sheen or other obvious sign of contamination is observed during sample collection or sampler washing.

5.3. Sampling Logs

As sediment is collected, sampling/field logs must be completed. The following should be included in the log:

- Date and time of collection of each sediment sample.
- Map of the harbor and sample locations.

- Names of field supervisors and person(s) collecting and logging in the sample.
- Weather conditions.
- The sample station number.
- Notation of any resistance during collection
- Depth of refusal
- The outcome of each sampling attempt – either ‘accepted’ or ‘discarded’.
- The measured water depth at each sampling station and the real-time tidal stage at the time of sampling at each station. The mudline elevation, referenced to MLLW, must then be calculated by subtracting the tidal stage from the measured water depth. The method/procedure used to determine the real-time tidal stage should be documented in the log. Real time mudline calculations must be in the sampling log, referencing the appropriate datum.
- Physical sediment description, including type, density, color, consistency, odor, vegetation, debris, biological activity, presence of an oil sheen, or any other distinguishing characteristics or features.
- Photo of each sample taken when in the split spoon. Each photo will include a notation that includes date, time, sample station number.
- Any deviation from the approved sampling plan.

5.4. Sample Preservation, Containers, and Holding Times

The sample containers, preservation methods, and holding times discussed in Sections 7.4.9 of the ADMEF will be observed. In general, the maximum sample hold times will be controlled by the 14-day hold time associated with semi-volatiles, pesticides, PCBs, and TPH.

5.5. Sample Identification and Labeling

Each sample must be sealed in a labeled container immediately after it is collected. Labels may be filled out, firmly affixed to the container and covered with clear tape prior to collection to minimize handling of the sample containers. The labels will include (at a minimum) the following information:

- Name/Initials of collector
- Sample Number
- Collected Date and Time
- Analytical Method
- Matrix
- Preservative

The Corps of Engineers sample numbering system will be used for all samples. The first two digits represent the fiscal year: 25. If the drilling schedule causes the sampling start after September 30th, the first two digits will be “26” for fiscal year 2026. The next three letters represent the specific site designation for the project: HOM, for “Homer”. This will be followed by a dash (-), then followed by “CW” for “Civil Works”; followed by the sequential borehole location number (i.e. 01 through 08); followed by “SD” for “sediment”; and finally a dash preceding vertical position identifier (i.e. 01, 02, or 03 for each layer encountered in a borehole). For example: 25HOM-CW01SD-01 representing the surface sample from borehole location #1. Duplicate samples will be submitted blind to the lab using a false vertical position identifier (e.g., 25HOM-CW01SD-04).

5.6. Sample Packing and Shipping

Samples will be shipped to the analytical laboratory with sufficient time allowed for the laboratory to extract the sample within the holding time requirements of the test method. Samples will be shipped to the laboratories using the fastest delivery possible and packaged according to the Department of Transportation (DOT) regulations. Samples will be packed and shipped according to the following specifications:

- Wrap each jar in bubble wrap or other cushioning material (e.g., foam blocks or sleeves) and then individually seal jars in plastic bags labeled with the sample number to keep jars separate and prevent breakage.
- Place the jars upright inside waterproof plastic coolers.
- Insert break-proof artificial ice packs around, among, and on top of the jars to maintain a sample temperature around 4 degrees Celsius.
- Place paperwork (chain of custody record, appropriate field notes and return address labels for the coolers) in a waterproof plastic bag and taped to the inside of the cooler.
- Secure the lid of the cooler and drain (if present) with tape. Wrap the cooler with strapping tape at a minimum of two locations, without obscuring any labels.
- Affix the completed shipping label to the top of the cooler.
- Adhere dated and signed custody seals to front right and back left of cooler to prevent accidental opening.
- Notify the analytical laboratory prior to shipment of any known or suspected highly contaminated samples; these samples will be stored separately from less contaminated samples to minimize the potential for cross-contamination.

5.7. Equipment Decontamination

Dedicated or single-use disposable sampling equipment (sampling spoons) will be used to the extent possible. Non-disposable sample handling equipment, such as hand trowel or shovels, will be wiped free of adhering soil with a new paper towel and rinsed with

seawater prior to moving to the next sampling point. Non-disposable sampling equipment will be decontaminated at the end of the sampling effort. All sampling equipment that potentially encountered contaminants, including split spoon samplers, will be decontaminated before each use to prevent cross contamination as well. The decontamination procedure consists of the following steps:

- Clean equipment using a non-ionic detergent and a brush to remove particulate matter or surface films
- Perform a double potable water rinse
- Allow equipment to air dry
- Package or otherwise prevent equipment from coming into contact with contaminated materials

Disposable sampling equipment will not be decontaminated since it arrives clean and factory-sealed and is used only for a single sample collection. The analytical laboratory will provide pre-cleaned sample containers.

5.8. Investigation Derived Waste

Single-use sampling equipment and trash (paper towels, etc.) will be disposed of appropriately as ordinary solid waste.

6. Data Quality Management

The following steps will be followed to assure adequate quality of data:

6.1. Laboratories

The primary laboratory must hold a current Department of Defense (DOD) Environmental Laboratory Accreditation Program (ELAP) approval and be approved by the State of Alaska to perform the relevant analyses. The laboratory will inspect each cooler of samples received to ensure that proper packaging; labeling and preservation practices were followed and will document and report any discrepancies.

6.2. Field Duplicate Samples

Two quality control field duplicate samples will be collected from the overall project sediment. This frequency meets the ADEC recommend rate of 10% duplicates for each analytical chemical method and each matrix sampled.

6.3. Matrix Spike/Matrix Duplicates

Matrix spikes and matrix spike duplicates will be collected at a frequency of at least one per twenty samples, with at least one collected per sediment type (e.g., sand, silt, clay, etc).

6.4. Analytical Data Quality Objectives

The analytical data quality objectives (DQOs) for sensitivity are based upon expected use of the data: comparison to the project screening levels presented in the ADMEF.

Laboratory reporting limits will be at or below project screening levels where technically feasible. Site-specific DQOs for accuracy or precision have not been developed for this project; therefore, the default limits in the QSM (DoD 2021) and Alaska Methodologies (18 AAC 75) will apply.

7. References

- Alaska Department of Environmental Consideration (ADEC). 2018. Guidance for Evaluating Metals at Contaminated Sites. August 2018.
- Dredged Material Management Office (DMMO). 2021. Dredged Material Evaluation and Disposal Procedures User Manual. Dredged Material Management Office, U.S. Army Corps of Engineers, Seattle District. July 2021.
- EPA, 1980. 40 CFR Part 230 Section 404(b)(1) Guidelines for Specification of Disposal Sites for Dredged or Fill Material, Environmental Protection Agency, December 1980.
- Gough, L.P., Severson, R.C., and Shacklette, H.T., 1988. Element Concentrations in Soils and Other Surficial Materials of Alaska. U.S. Geological Survey Professional Paper 1458. 53 p.
- EPA/USACE. 1998. Evaluation of dredged material proposed for discharge in waters of the U.S. – Testing manual. EPA-823-B-98-004, Washington, D.C
- Northwest Regional Sediment Evaluation Team (RSET). 2018. Sediment Evaluation Framework for the Pacific Northwest. Prepared by the RSET Agencies, May 2018, 183 pp plus appendices

Figures and Tables



Figure 1. Homer, Alaska location map

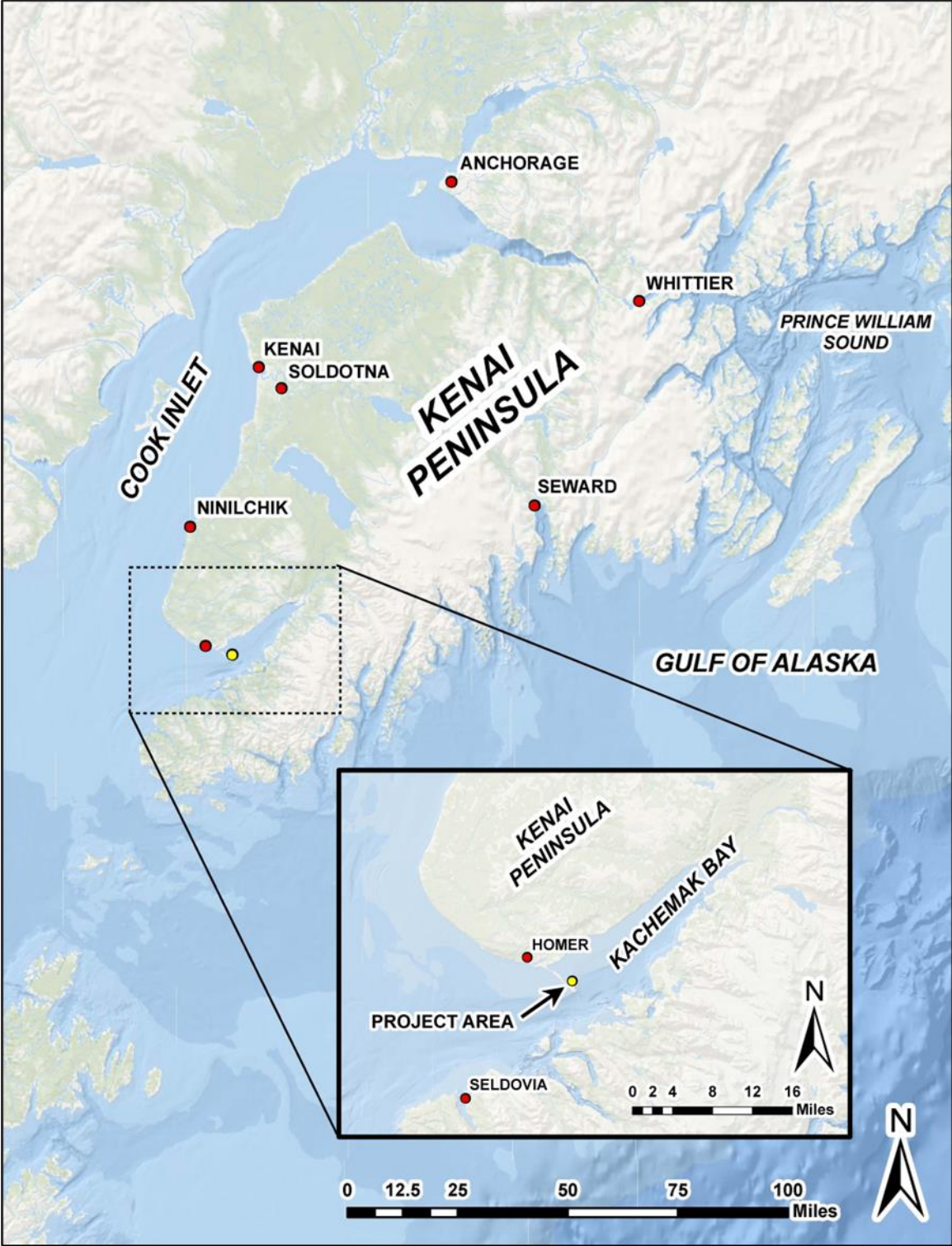


Figure 2. Homer and Kenai Peninsula map

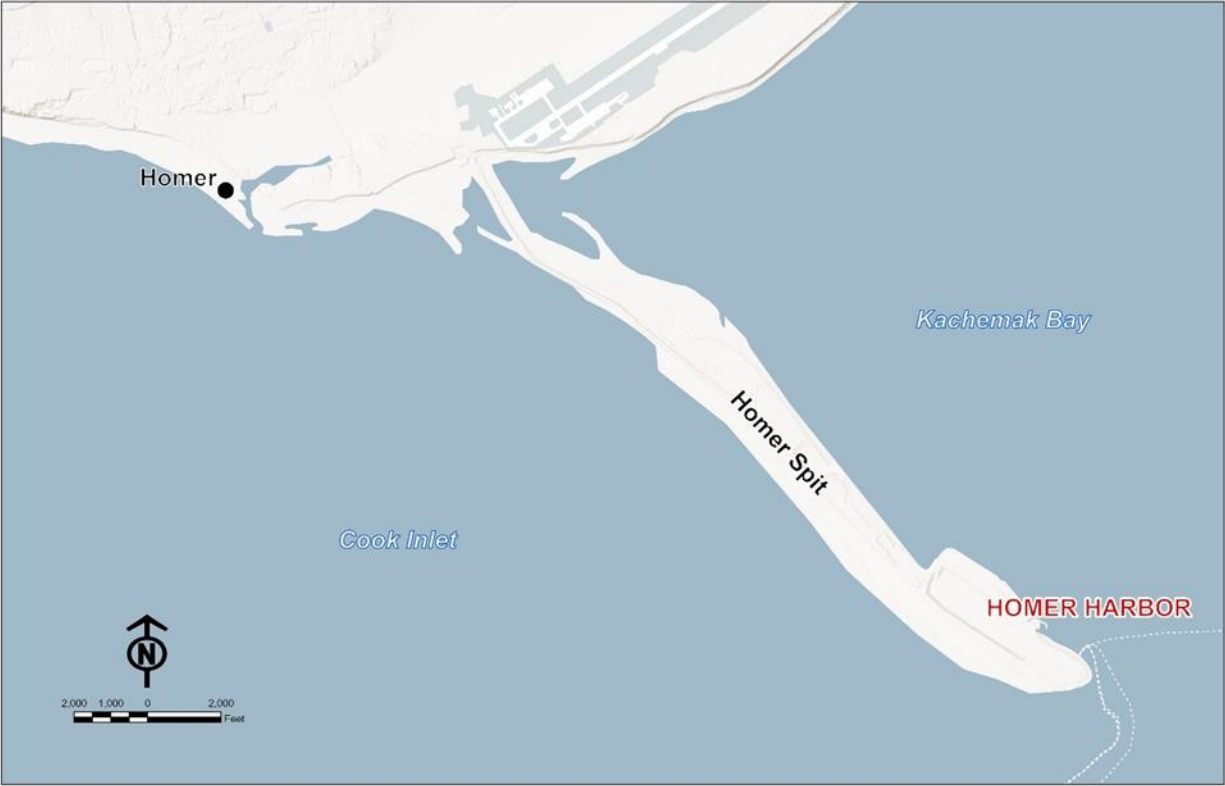


Figure 3. Homer Spit and existing small boat harbor.

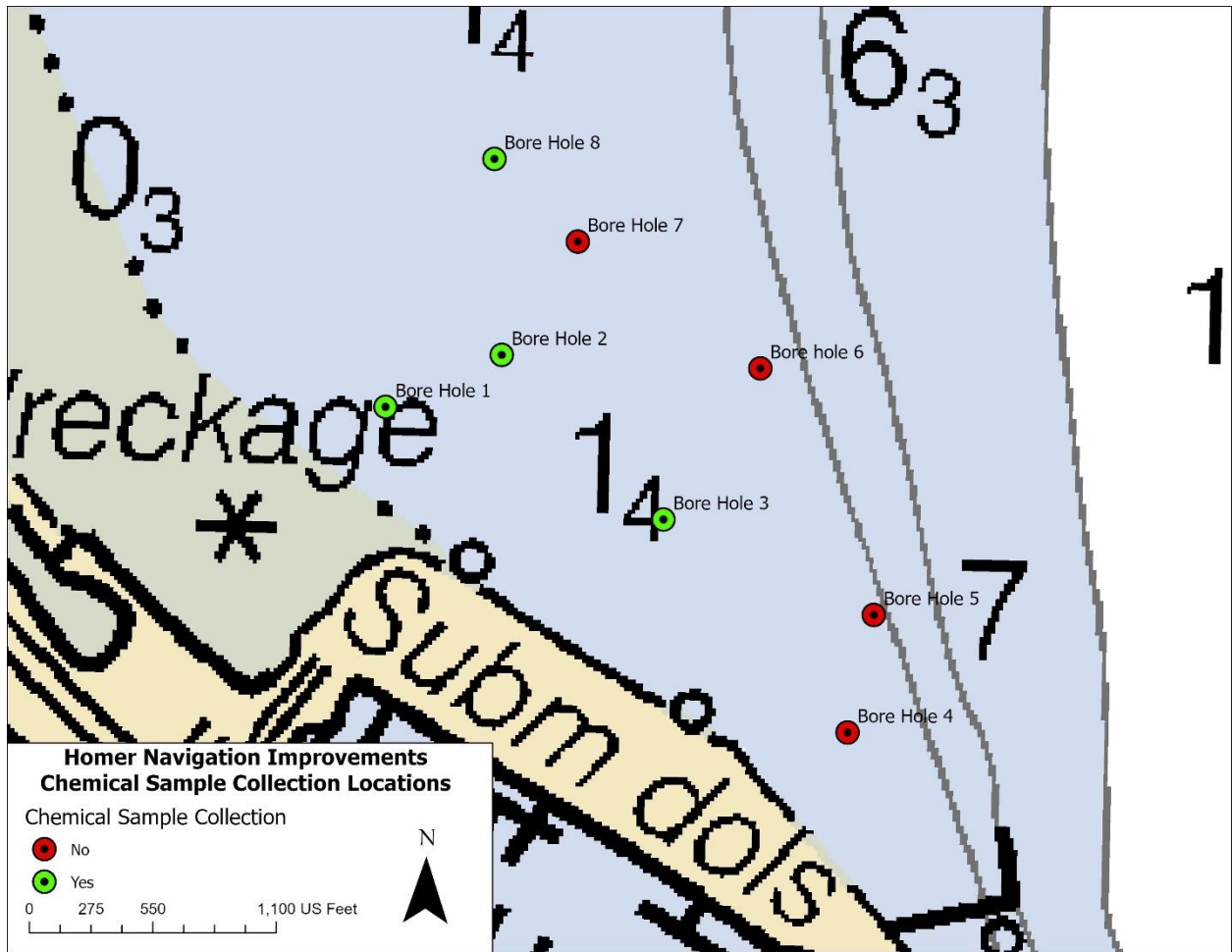


Figure 4. Geotechnical investigation borehole locations relevant to the characterization of dredged material

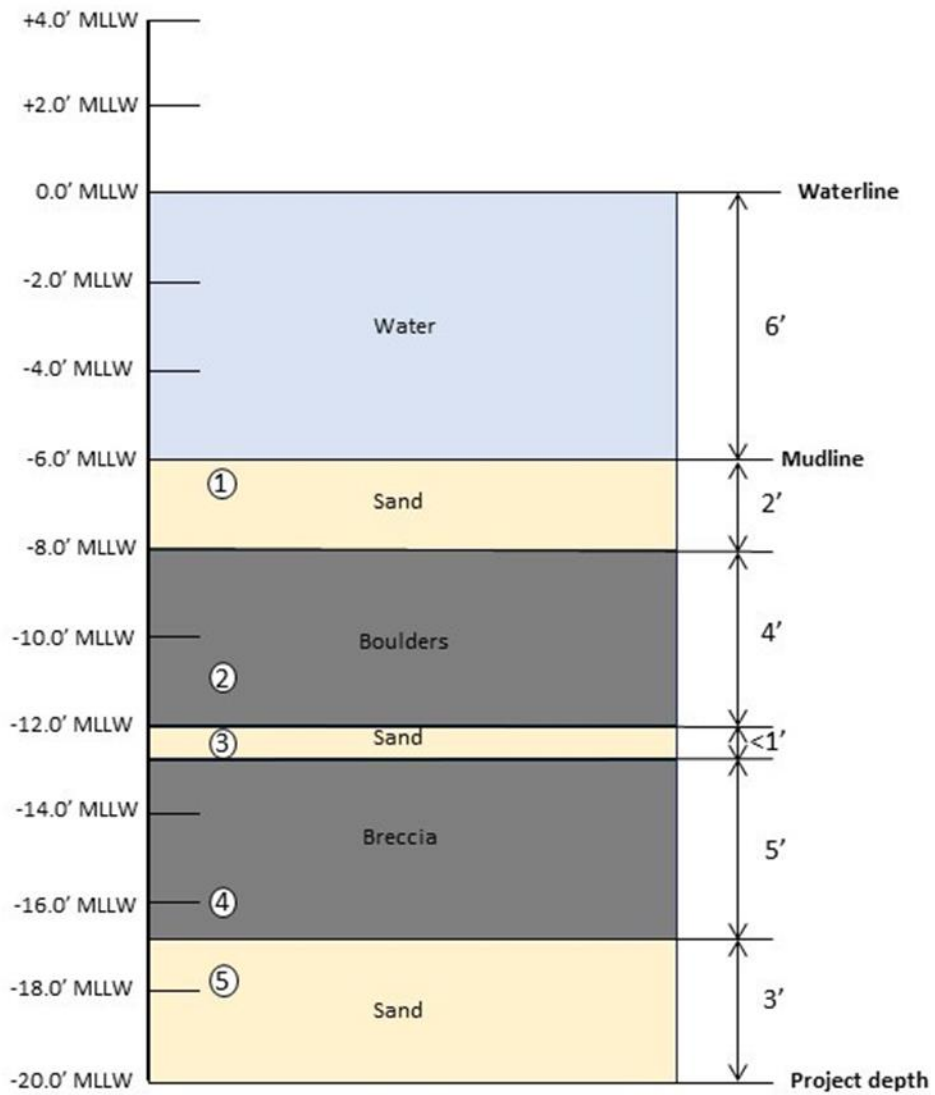


Figure 5. Example chemical sample collections section view diagram

Table 1. Borehole locations, depth, and preliminary determination regarding chemical relevance

Point	Easting (ft)	Northing (ft)	Elevation (ft)	Depth	Longitude	Latitude	Chemical Sample
Bore Hole 1	2052799.855	1375649.472	-7	80ft	-151.429752	59.611627	Yes
Bore Hole 2	2053031.104	1376167.519	-9	80ft	-151.426982	59.61229	Yes
Bore Hole 3	2052298.727	1376887.798	-10.5	20ft	-151.423009	59.610328	Yes
Bore Hole 4	2051351.092	1377708.303	-11	80ft	-151.418471	59.607784	No
Bore Hole 5	2051874.169	1377825.069	-16.5	80ft	-151.417901	59.609222	No
Bore hole 6	2052971.079	1377319.767	-15	80ft	-151.420755	59.612193	No
Bore Hole 7	2053535.138	1376506.431	-11	80ft	-151.425211	59.613688	No
Bore Hole 8	2053902.962	1376135.073	-10	20ft	-151.427258	59.614672	Yes

7 OCT 2025

HOMER NAV. IMPRO

~~FLYGLASS CAMERA SYSTEM~~
~~CF 1st sample per second~~

ARRIVE HOMER HBR 0630

MEET SETH FROM STW

0657 DRILLERS NOT ARRIVED

0730 Back to room

1130 Back to harbor

1141 board landing craft
with Seth

1155 Leave harbor, head for
barling 8

Conditions: Pthy Clouds,
light breeze, 49°F, sea calm

Scale: 1 square =

7 OCT 2025

3

1213 Arrive barling 8
Seth using hand-held GPS

1220 Release anchor

1232 Still maneuvering
swell kicking up

1251 2 anchors out, still maneuvering
1257 in position

1310 Drillers working on
another problem

1322 Under depth 30ft, tide rising

1335 waiting for tide to settle
before starting drilling

1508 start drilling BH 8

1511 1st sample gray mud
spoon pressed in to mud like

24" full spoon gravel towards
bottom

Scale: 1 square =

4 7 OCT 2025

1515 BHZ

Sample 81 3 jars

1542 second drive 7.2-9.2
problems w heave
more gray silt w sand
Shor & sample - give to geotech

1609 third drive ~15-17
more heave - spoon capped
and preserved by geotech

1615 fourth drive 17.8-19.8
Sample 82 3 jars
Stiff gray silt

mechanical problems

Head back to harbor for crew
change ~ 1830

Scale: 1 square =

Plot in the Rain

5 8 OCT 2025 HOMER NAV

1911 - head out to BHZ
conditions - overcast, light wind
seas calm

1920 - arrive at general location
of BHZ

1945 on station @ BHZ

Drillers going back to harbor
for sand reodor

2025 Drillers return
much pissing around
lights go out, back on again

2147 getting ready to take a
sample?

2155 Sample retrieved

Scale: 1 square =

8 OCT 2025

BH 2

2200

Sample 21, 20 (acc dip)

5 ft sleeve partly filled with dark gray silt, scattered gravel

Sample Collected from top 24"

2311 - Drillers broke something

9 OCT 2025

0020 - sample arrives

5 ft sleeve partly filled with stiff dark gray silt

Sample 22 9'-10'

Some coal fragments

Scale: 1 square =

Scale: 1 square =

Rate in inches

9 OCT 2025

0040 - Sample 23

dark gray silt w few gravel pieces

15'-17'

I leave barge ~0130

9 OCT 2025

Drilling on B-2 continues until ~1230

DR BARGE BLOWS PRENT

ANCHOR ROTOR - RETURNS TO HARBOR - PROJECT DOWN

1500 - PARTS ordered, due to arrive pm 10 OCT

10 OCT 2025

1151 - Barge still awaiting parts

8 9 OCT 2025

Intertidal Samples

Low tide 1102
59°36'35" N 151°25'52"

Flat pebbly silt with sparse
eelgrass - very wet

1130-1140

90+ 91 SD collected
0-0.5'

End of day



Scale: 1 square =

Rate in the rain

10 OCT 2025

Board barge @ 1605

1747 - in position @ BH-3

1918 - lands on barge after
5 h:14 change

2013 - 1st 5' core

dark gray silt w scattered
gravel - surface smooth

Sample surface 24"

-31 SD

2106 = 2nd 5ft

dark gray silt w scattered gravel

-32 SD top 6" (-15' MHW)

-33 SD bottom 6" (20' MHW)

Scale: 1 square =

10 OCT 2025

BH-3

2139 15'-17" below water level
gray silt
NOT SAMPLED

2155 20-22' below ML

NOT SAMPLED

- Wait for tide to move Barge
Drillers need some chairs

head in to station ~ 0100

end of day



Scale: 1 square =

Scale: 1 square =

11 OCT 2025

BH-1

Arrive on Barge ~ 1025
sample waiting

1028 - 1150

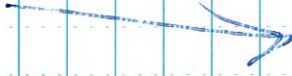
soft dark gray silt, no gravel

1040 - short sample 5-10'
give to geotech

1050 - 1230 15-17'

Soft dark gray silt

end of sampling



Scale: 1 square =

Plot on the Rain

Chemical Data Quality Review – Homer Navigation Improvements Feasibility Study Dredged Material Characterization, 2025

1. Introduction

1.1. The U.S. Army Corps of Engineers Alaska District (USACE-AK), Engineering Division, Environmental Engineering Branch (CEPOA-ENE) prepared this data review at the request of the USACE Civil Works branch, Environmental Resources Section (CEPOA-PMC-E). This report presents a review of the results from the 2025 sediment investigation conducted by USACE-AK personnel at Homer Harbor in support of Dredging Navigation Improvements efforts. This CDQR covers Sample Delivery Group (SDG) 580-154935-1/2.

2. Project Description

2.1. The Alaska District is investigating navigation improvements that would provide safe, reliable and efficient waterborne transportation systems and moorage to support Homer's current and future fleet. The implementation of any of the structural alternatives under consideration would include dredging and dredged material management to create adequate depths for the design vessel. Dredging would excavate material down to -26' mean lower low water (MLLW) for the entrance channel, -24' MLLW for the fairway, and -20' MLLW or -15' MLLW for the mooring area. USACE must characterize the dredged material to be excavated and managed to complete the cost estimate and environmental analysis needed for the feasibility study. The feasibility study currently assumes the dredged material is suitable for unconfined aquatic placement as the least costly environmentally acceptable management option.

2.2. Project results are compared to the marine limits presented in Section 8 of the Dredged Material Evaluation and Disposal Procedures User Manual (July 2021) developed by the Dredged Material Management Program (DMMP) and Alaska Department of Environmental Conservation (ADEC) 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels. Project data comparisons to these project action limits (PALs) are presented in Tables 2 and 3 at the end of this narrative.

2.3. A total of 14 samples (including 11 primary, two duplicates, and one trip blank) were delivered to Eurofins Anchorage Receiving Station (ARS) in Anchorage, AK with proper custody procedures. ARS submitted samples to EURO Lancaster who is approved for all analyses provided (6020B, 7471B, 8081B, 8082A, 8260D, 8270E, 8270ESIM, and AK101). This lab is approved by ADEC through the Underground Storage Tank (UST) Program and is certified by the Department of Defense (DOD) Environmental Laboratory Accreditation Program (ELAP) for the requested analytical methods. However, ARS also submitted samples for Tributyl Tin via GC/MS SIM to EURO CAL Science who is *not* approved by ADEC or ELAP for that method/analyte. Neither ADEC nor ELAP validate a method for Tributyl Tin. This deviation was

communicated to the Project Team prior to sample submittal, is considered acceptable, and does not impact data usability.

- 2.4.** The project data was reviewed for deviations to the requirements presented in the Sampling and Analysis Plan, the DOD-QSM (Version 6.0), and ADEC Technical Memorandum 22-001, Guidelines for Data Reporting (dated August 2022) in the following areas – precision, accuracy, representativeness, comparability, completeness, and sensitivity (PARCCS). Elements reviewed include sample handling, holding times, method and trip blanks, laboratory control sample/laboratory control sample duplicate (LCS/LCSD) recoveries and relative percent differences (RPDs), matrix spikes and matrix spike duplicates (MS/MSD) recoveries and RPDs, surrogate recovery, and field duplicate comparability. Calibration curves and continuing calibration standard recoveries were not specifically reviewed; however, laboratories are required to document such failures in the appropriate case narratives. These narratives were reviewed for each SDG.
- 2.5.** The laboratory staged electronic data deliverable (SEDD) for this project was used to generate this report. When discrepancies between the hardcopy data and the SEDD are found, the SEDD has been modified to reflect values from the hardcopy, unless the hardcopy is found to be in error. Results used to generate this report are deemed to be accurate.
- 2.6.** The following qualifiers, listed below in order of increasing severity, are used in the data tables to indicate quality control deficiencies. The most severe flag will be utilized when quality issues indicate the use of more than one qualifier.

Qualifier	Definition
J	Analyte result is considered an estimated value because the level is below the laboratory LOQ but above the DL
B	Analyte result is considered a high estimated value due to contamination present in the method, trip, or equipment blank.
J+ J- JN	The result is considered an estimated value biased (high, low, indeterminant) due to a quality control deficiency. Result is considered acceptable.
R	Analyte result is rejected - result is not usable for project.
Note:	Flags may be combined when more than one quality deficiency exists.

Details of the data review are presented by SDG below:

3. SDG 580-154935-1/2

- 3.1.** Collection and Preservation: Fourteen project samples were included in this SDG, including 11 primary samples, two duplicates, and one trip blank. All samples included in this SDG were shipped to the laboratory in a single cooler. Samples were received at ASR in one cooler at 1.3° C. ASR submitted samples to two different network labs: EURO Lancaster (SDG 580-154935-1, rec'd at 1.5° C for all methods

but Tributyl Tin); and EURO Cal Science (580-154935-2, rec'd at 3.5° C for Tributyl Tin) and all containers were deemed to be in acceptable condition.

- 3.2.** Holding times: All reported sample analyses were completed within applicable holding times.
- 3.3.** The laboratory case narrative did identify quality deficiencies. Deficiencies that impact results are discussed below.
- 3.4.** Method, equipment, and trip blanks were analyzed at the required frequency. The following analytes were detected in an associated method blank: Fluoranthene (8270 and 8270SIM), Pyrene (8270 and 8270SIM), Anthracene, Benzo(a)anthracene, Benzo(a)pyrene, Benzo(b)fluoranthene, Chrysene, and Phenanthrene (8270SIM only). All associated detected sample results less than 10 times the associated blank value are flagged "B" in the data tables. However, all detections are significantly below (i.e. less than 20%) of the applicable DMMP screening criterion.
- 3.5.** LCS/LCSDs were analyzed at the required frequency and recoveries were within the QSM acceptance limits for all analytes in project samples.
- 3.6.** Per the Sampling and Analysis Plan (SAP), at least one MS/MSD was to be submitted and analyzed for each method. However, a MS/MSD was not identified on the project chain of custody (CoC). The lab, however, ran a MS/MSD for method 8270E and for one of three extraction batches for 8270ESIM (but none on the other methods). Several analytes for both methods failed low in either the MS, the MSD, or both. Analytes which failed low but recovered above 10% are flagged "J-" and analytes which recovered below 10% are flagged "R" (unusable). Analytes with unusable results include 2,4-Dinitrophenol, Benzoic Acid, and Hexachlorocyclopentadiene. Only the associated sample results (26-HOM-CW 32 SD) are flagged.
- 3.7.** LCS/MS Precision: Either a LCSD, MSD, or an internal laboratory duplicate is required for evaluation of laboratory precision. Thus, precision cannot be evaluated for methods 6020B, 7471B, D2216, and 8082A (no duplicate, LCSD, or MSD). Likewise, precision cannot be evaluated for 8081B batch 410-717133, 8270ESIM batch 410-719765, and 8270ESIM batch 410-721118. However, all impacted results are below the applicable DMMP PAL, and data usability is not significantly impacted. For 8270E and 8270ESIM, several analytes also exceeded the RPD criterion for the reported MS/MSD. If the analyte failed low for MS and/or MSD recoveries (but above 10%), the "J-" flag was changed to "JN" to indicate potential bias of unknown direction. "R" flags due to lack of MS/MSD recoveries were not changed. Again, this pertains only to the impacted sample 26-HOM-CW 32 SD.
- 3.8.** Surrogate recoveries for all samples were within method and/or QSM acceptance limits except that one of four surrogates failed low for method 8260D in samples 26-HOM-CW 12 SD and TB 1; however, recoveries were greater than 10%. Consequently, all analytes for these samples are flagged "J-". One of one surrogate

failed low in multiple samples for AK101 where results indicate the samples were diluted significantly (i.e. at least 25X). Results are not further flagged.

3.9. There are two duplicates of site samples reported in this SDG which meets the ADEC 10% frequency requirement. Sample 26-HOM-CW 20 SD is a duplicate of 26-HOM-CW 21 SD and 26-HOM-CW 90 SD is a duplicate of 26-HOM-CW 91 SD. For comparison purposes, the limit of detection (LOD) is used for a nondetect result. For primary/duplicate pair -21 SD/-20 SD, the following methods/analytes did not meet ADEC requirement of less than 50% RPD: 8270E: Chrysene and Fluoranthene; 8270ESIM - Acenaphthene, Acenaphthylene, Anthracene, Benzo(a)anthracene, Benzo(a)pyrene, Benzo(b)fluoranthene, Benzo(g,h,i)perylene, Chrysene, Dibenzo(a,h)anthracene, and Fluoranthene. For primary duplicate pair -91 SD/-90 SD, the following methods/analytes did not meet ADEC requirements: 8081, nearly all analytes; 8270ESIM - Phenanthrene, Fluoranthene, Acenaphthene, Anthracene, Acenaphthylene. Impacted analytes are all flagged "JN"; results are either nondetect or well below the DMMP PAL; thus, usability is not impacted by this deviation.

3.10. Reporting/detection limits are defined by the QSM as follows: the Limit of Quantification (LOQ) is the lowest concentration that produces a quantitative result within specified limits of precision and bias. For DOD projects, the LOQ shall be set at or above the concentration of the lowest initial calibration standard corrected for sample preparation, dilution and moisture (if applicable). Laboratories can often detect analytes at levels less than the LOQ, albeit less quantitatively; therefore, the Limit of Detection (LOD) is defined as the smallest amount or concentration of a substance that must be present in a sample in order to be detected at a high level of confidence (99%). At the LOD, the false positive rate is 1%. Consequently, any nondetect result with an LOD greater than the associated cleanup limit cannot be used to prove the absence of that analyte at that limit.

A total of 21 nondetect analytes exceeded the DMMP PALs in one or more samples. These results are highlighted yellow in the DMMP data table (Table 2). A comparison of reported LODs to the most conservative ADEC Cleanup Limits specified in 18 AAC 75 was also completed; a total of 40 nondetect analytes exceeded the ADEC levels in at least one sample. These are also highlighted yellow in the ADEC data table (Table 3). Tables 4 and 5 present a summary of the numbers of samples and exceedances due to this deficiency.

3.11. Review of the laboratory case narrative indicates additional deficiencies not identified above. For 8270E, the narrative states that Acetone and Chloroethane failed low (76%/76%) in the opening Continuing Calibration Verification (CCV) standard associated with batch 410-716956, impacting all samples. Impacted results are flagged "J-". All impacted results are nondetect and are well below ADEC PALs (DMMP PAL is not established for these compounds). Results are considered usable.

The lab narrative also indicates that 8270E closing CCV for Hexachlorocyclopentadiene recovered low, that CCV and samples were rerun and the

closing CCV again recovered low and therefore the data is acceptable and the deviations are due to matrix interference. However, there is no indication in the data package that the samples were rerun. The original sample that was spiked was run first (26-HOM-CW 32 SD), followed by the MS and MSD, both of which did not recover (0%). The rest of the samples were analyzed next followed by the closing CCV, which recovered at 21%, which is below the acceptable DoD Limit of 50%. For these reasons, this compound is considered rejected in all samples.

4. Overall Assessment

The project data exhibits ADEC PAL exceedances for Arsenic in all samples. However, Arsenic is a naturally occurring metal in Alaska. It is often found in concentrations above the regulatory cleanup level. In addition, Naphthalene also exceeded the ADEC PAL in sample 26-HOM-CW 11 SD. This deviation may indicate that low level fuel contamination may be present in that location. There were no exceedances of DMMP PALs in this data set.

The data set does contain some rejected results. All results for Hexachlorocyclopentadiene and results for 2,4-Dinitrophenol and Benzoic acid in sample 26-HOM-CW 32 SD are considered rejected. All other results for this project are usable as reported and flagged. The overall completeness goal of 95% was met.

5. References

- 5.1. ADEC, 18 AAC 75, Oil and Other Hazardous Substances Pollution Control. October 2023.
- 5.2. ADEC, Technical Memorandum 22-001, Guidelines for Data Reporting. August 2022.
- 5.3. Department of Defense, Quality Systems Manual for Environmental Laboratories, Final Version 6.0. January 2023.
- 5.4. Dredged Material Management Office, Dredged Material Evaluation and Disposal Procedures User Manual. Dredged Material Management Office, U.S. Army Corps of Engineers, Seattle District. July 2021.
- 5.5. Eurofins, Laboratory Analytical Report; 580-154935-1/-2, 25-056 Homer Harbor Sediment Characterization. December 2025.
- 5.6. USACE, Sampling and Analysis Plan, Homer Navigation Improvements Feasibility Study Dredged Material Characterization, Homer, Alaska. September 2025.

Tables:

- Table 1 - 2025 Homer Feasibility Study Sample Summary
- Table 2 – 2025 Homer Feasibility Study, DMMP Comparison
- Table 3 – 2025 Homer Feasibility Study, ADEC Comparison
- Table 4 – 2025 Homer Feasibility Study, DMMP PAL Exceedances
- Table 5 – 2025 Homer Feasibility Study, ADEC PAL Exceedances

Attachment 1: ADEC Data Review Checklist, SDG 580-154935-1/2

**Table 1: 2025 Homer Feasibility Study
Sample Summary**

Sample Delivery Group	CoolerID	ClientSampleID	LabSampleID	CollectedDate	MatrixID	LabName	QCType	6020B	7471B	8081B	8082A	8260D	8270E	8270ESIM	AK101	D2216	TBT-SIM
580-154935-1	LB	26-HOM-CW 11 SD	580-154935-12	10/11/2025 10:28:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 11 SD	580-154935-12	10/11/2025 10:28:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 12 SD	580-154935-13	10/11/2025 10:50:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 12 SD	580-154935-13	10/11/2025 10:50:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 20 SD	580-154935-3	10/08/2025 22:00:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 20 SD	580-154935-3	10/08/2025 22:00:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 21 SD	580-154935-4	10/08/2025 22:15:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 21 SD	580-154935-4	10/08/2025 22:15:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 22 SD	580-154935-5	10/09/2025 00:20:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 22 SD	580-154935-5	10/09/2025 00:20:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 23 SD	580-154935-6	10/09/2025 00:40:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 23 SD	580-154935-6	10/09/2025 00:40:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 31 SD	580-154935-9	10/10/2025 20:13:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 31 SD	580-154935-9	10/10/2025 20:13:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 32 SD	580-154935-10	10/10/2025 21:06:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 32 SD	580-154935-10	10/10/2025 21:06:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 33 SD	580-154935-11	10/10/2025 21:15:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 33 SD	580-154935-11	10/10/2025 21:15:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 81 SD	580-154935-1	10/07/2025 15:15:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 81 SD	580-154935-1	10/07/2025 15:15:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 82 SD	580-154935-2	10/07/2025 16:15:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 82 SD	580-154935-2	10/07/2025 16:15:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 90 SD	580-154935-7	10/09/2025 11:30:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 90 SD	580-154935-7	10/09/2025 11:30:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	26-HOM-CW 91 SD	580-154935-8	10/09/2025 11:40:00	Soil	Eurofins Lancaster	Field_Sample	X	X	X	X	X	X	X	X	X	
580-154935-2	LB-1	26-HOM-CW 91 SD	580-154935-8	10/09/2025 11:40:00	Soil	Eurofins Cal Science	Field_Sample										X
580-154935-1	LB	TB 1	580-154935-14	10/06/2025 00:00:00	Soil	Eurofins Lancaster	Trip Blank					X			X		

**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	DMMP	Dup of 21 SD				
6020B	mg/kg	Antimony	150	0.37 [0.24]	0.39 [0.15]	0.44 [0.19]	0.34 [0.20]	0.37 [0.20]
6020B	mg/kg	Arsenic	57	14 [0.48]	13 [0.29]	10 [0.38]	11 [0.40]	9.2 [0.39]
6020B	mg/kg	Cadmium	5.1	0.079 [0.12] J	0.097 [0.073]	0.095 [0.095] J	0.095 [0.099] J	0.11 [0.098] J
6020B	mg/kg	Chromium	260	55 [0.57]	42 [0.35]	53 [0.45]	47 [0.47]	40 [0.47]
6020B	mg/kg	Copper	390	49 [0.54]	30 [0.33]	47 [0.43]	42 [0.45]	31 [0.44]
6020B	mg/kg	Lead	450	12 [0.24]	7.2 [0.15]	11 [0.19]	9.9 [0.20]	7.4 [0.20]
6020B	mg/kg	Nickel	NE	41 [0.57]	31 [0.35]	41 [0.45]	38 [0.47]	32 [0.47]
6020B	mg/kg	Selenium	3	0.30 [0.30] J	0.18 [0.18] J	0.29 [0.24] J	0.26 [0.25] J	0.20 [0.25] J
6020B	mg/kg	Silver	6.1	0.13 [0.12] J	0.085 [0.073] J	0.13 [0.095]	0.13 [0.099]	0.088 [0.098] J
6020B	mg/kg	Zinc	410	110 [12]	79 [7.3]	110 [9.5]	98 [9.9]	83 [9.8]
7471B	mg/kg	Mercury	0.41	0.14 [0.063]	0.084 [0.052]	0.13 [0.062]	0.12 [0.057]	0.098 [0.054]
8081B	mg/kg	4,4'-DDD	0.016	ND [0.013]	ND [0.011]	ND [0.013]	ND [0.012]	ND [0.011]
8081B	mg/kg	4,4'-DDE	0.009	ND [0.012]	ND [0.0094]	ND [0.011]	ND [0.011]	ND [0.0096]
8081B	mg/kg	4,4'-DDT	0.012	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Aldrin	0.0095	ND [0.0059]	ND [0.0048]	ND [0.0057]	ND [0.0054]	ND [0.0049]
8081B	mg/kg	alpha-BHC	NE	ND [0.0063]	ND [0.0051]	ND [0.0060]	ND [0.0057]	ND [0.0052]
8081B	mg/kg	beta-BHC	NE	ND [0.0074]	ND [0.0060]	ND [0.0071]	ND [0.0068]	ND [0.0062]
8081B	mg/kg	cis-Chlordane	0.0028	ND [0.0076]	ND [0.0062]	ND [0.0072]	ND [0.0069]	ND [0.0063]
8081B	mg/kg	delta-BHC	NE	ND [0.0074]	ND [0.0060]	ND [0.0071]	ND [0.0068]	ND [0.0062]
8081B	mg/kg	Dieldrin	0.0019	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Endosulfan I	NE	ND [0.0049]	ND [0.0040]	ND [0.0047]	ND [0.0045]	ND [0.0041]
8081B	mg/kg	Endosulfan II	NE	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8081B	mg/kg	Endosulfan sulfate	NE	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Endrin	NE	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Endrin aldehyde	NE	ND [0.014]	ND [0.011]	ND [0.013]	ND [0.013]	ND [0.012]
8081B	mg/kg	Endrin ketone	NE	ND [0.015]	ND [0.012]	ND [0.014]	ND [0.014]	ND [0.012]
8081B	mg/kg	gamma-BHC (Lindane)	NE	ND [0.0049]	ND [0.0040]	ND [0.0047]	ND [0.0045]	ND [0.0041]
8081B	mg/kg	Heptachlor	0.0015	ND [0.0051]	ND [0.0042]	ND [0.0049]	ND [0.0047]	ND [0.0043]
8081B	mg/kg	Heptachlor Epoxide	NE	ND [0.0058]	ND [0.0047]	ND [0.0055]	ND [0.0053]	ND [0.0048]
8081B	mg/kg	Methoxychlor	NE	ND [0.049]	ND [0.040]	ND [0.046]	ND [0.044]	ND [0.040]
8081B	mg/kg	Toxaphene	NE	ND [0.23]	ND [0.19]	ND [0.22]	ND [0.21]	ND [0.19]
8081B	mg/kg	trans-Chlordane	0.0028	ND [0.0049]	ND [0.0040]	ND [0.0047]	ND [0.0045]	ND [0.0041]
8082A	mg/kg	PCB-1016 (Aroclor 1016)	0.13	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]

¹MS/MSD was run only on 8270E and 8270ESIM methods

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NE - Not Established

**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	DMMP	Dup of 21 SD				
8082A	mg/kg	PCB-1221 (Aroclor 1221)	0.13	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1232 (Aroclor 1232)	0.13	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1242 (Aroclor 1242)	0.13	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1248 (Aroclor 1248)	0.13	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1254 (Aroclor 1254)	0.13	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1260 (Aroclor 1260)	0.13	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8260D	mg/kg	1,1,1-Trichloroethane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,1-Dichloroethane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,1-Dichloroethene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,1-Dichloropropene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2,3-Trichlorobenzene	NE	ND [5.5]	ND [0.33] J-	ND [0.50]	ND [0.36]	ND [0.35]
8260D	mg/kg	1,2,3-Trichloropropane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2,4-Trichlorobenzene	0.031	ND [5.5]	ND [0.33] J-	ND [0.50]	ND [0.36]	ND [0.35]
8260D	mg/kg	1,2,4-Trimethylbenzene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2-Dibromo-3-chloropropane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2-Dichlorobenzene	0.035	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2-Dichloropropane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,3,5-Trimethylbenzene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,3-Dichlorobenzene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,3-Dichloropropane	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	2,2-Dichloropropane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	2-Butanone	NE	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	2-Chlorotoluene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	4-Chlorotoluene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	4-Isopropyltoluene	NE	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	4-Methyl-2-pentanone	NE	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Acetone	NE	ND [11] J-	ND [0.67] J-	ND [1.0] J-	ND [0.72] J-	ND [0.69] J-
8260D	mg/kg	Bromobenzene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	Bromochloromethane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Carbon disulfide	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Carbon tetrachloride	NE	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Chlorobenzene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Chloroethane	NE	ND [2.7] J-	ND [0.17] J-	ND [0.25] J-	ND [0.18] J-	ND [0.17] J-
8260D	mg/kg	Chloromethane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	cis-1,2-Dichloroethene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]

¹MS/MSD was run only on 8270E and 8270ESIM methods

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	DMMP	Dup of 21 SD				
8260D	mg/kg	Dichlorodifluoromethane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Ethylbenzene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	Isopropylbenzene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	Methylene chloride	NE	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Methyl-tert-butyl ether (MTBE)	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	n-Butylbenzene	NE	ND [3.4]	ND [0.21] J-	ND [0.31]	ND [0.23]	ND [0.22]
8260D	mg/kg	n-Propylbenzene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	o-Xylene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	sec-Butylbenzene	NE	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Styrene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	tert-Butylbenzene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Tetrachloroethene (PCE)	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Toluene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	trans-1,2-Dichloroethene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Trichloroethene (TCE)	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Trichlorofluoromethane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Vinyl chloride	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Xylene, Isomers m & p	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8270E	mg/kg	1,2,4-Trichlorobenzene	0.031	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1,2-Dichlorobenzene	0.035	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1,3-Dichlorobenzene	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1,4-Dichlorobenzene	0.11	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1-Methylnaphthalene	NE	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	2,2-oxybis(1-Chloropropane)	NE	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	2,4,5-Trichlorophenol	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2,4,6-Trichlorophenol	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2,4-Dichlorophenol	NE	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	2,4-Dimethylphenol	0.029	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2,4-Dinitrophenol	NE	ND [1.4]	ND [1.1]	ND [1.3]	ND [1.3]	ND [1.2]
8270E	mg/kg	2,4-Dinitrotoluene	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	2,6-Dinitrotoluene	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2-Chloronaphthalene	NE	ND [0.044]	ND [0.036]	ND [0.041]	ND [0.040]	ND [0.037]
8270E	mg/kg	2-Chlorophenol	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2-Methyl-4,6-dinitrophenol	NE	ND [0.55]	ND [0.45]	ND [0.52]	ND [0.50]	ND [0.46]
8270E	mg/kg	2-Methylnaphthalene	0.67	ND [0.017]	ND [0.013]	ND [0.016]	ND [0.015]	ND [0.014]

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Method	Units	Analyte	DMMP	Dup of 21 SD				
8270E	mg/kg	2-Methylphenol (o-Cresol)	0.063	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	2-Nitroaniline	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2-Nitrophenol	NE	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	3,3'-Dichlorobenzidine	NE	ND [0.44]	ND [0.36]	ND [0.41]	ND [0.40]	ND [0.37]
8270E	mg/kg	3-Methylphenol/4-Methylphenol Coelution	0.67	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	3-Nitroaniline	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	4-Bromophenyl phenyl ether	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	4-Chloro-3-methylphenol	NE	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	4-Chloroaniline	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	4-Chlorophenyl phenyl ether	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	4-Nitroaniline	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	4-Nitrophenol	NE	ND [0.55]	ND [0.45]	ND [0.52]	ND [0.50]	ND [0.46]
8270E	mg/kg	Acenaphthene	0.5	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Acenaphthylene	0.56	ND [0.013]	ND [0.011]	ND [0.012]	ND [0.012]	ND [0.011]
8270E	mg/kg	Anthracene	0.96	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(a)anthracene	1.3	ND [0.022]	ND [0.018]	ND [0.021]	ND [0.020]	ND [0.018]
8270E	mg/kg	Benzo(a)pyrene	1.6	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(b)fluoranthene	3.2	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(g,h,i)perylene	0.67	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(k)fluoranthene	3.2	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzoic acid	0.65	ND [0.66]	ND [0.54]	ND [0.62]	ND [0.60]	ND [0.55]
8270E	mg/kg	Benzyl alcohol	0.057	ND [0.55]	ND [0.45]	ND [0.52]	ND [0.50]	ND [0.46]
8270E	mg/kg	Benzyl butyl phthalate	0.063	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	bis-(2-Chloroethoxy)methane	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	bis-(2-Chloroethyl)ether	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	bis-(2-Ethylhexyl)phthalate	1.3	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Carbazole	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Chrysene	1.4	ND [0.011]	ND [0.0089]	0.0056 [0.010] J JN	ND [0.010] JN	ND [0.0092]
8270E	mg/kg	Dibenzo(a,h)anthracene	0.23	ND [0.022]	ND [0.018]	ND [0.021]	ND [0.020]	ND [0.018]
8270E	mg/kg	Dibenzofuran	0.54	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Diethyl phthalate	0.2	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Dimethyl phthalate	0.071	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Di-n-butyl phthalate	1.4	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Di-n-octyl phthalate	6.2	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Fluoranthene	1.7	ND [0.011]	ND [0.0089]	0.0098 [0.010] J JN	0.0053 [0.010] J JN	ND [0.0092]
8270E	mg/kg	Fluorene	0.54	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]

¹MS/MSD was run only on 8270E and 8270ESIM methods

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NE - Not Established

**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	DMMP	Dup of 21 SD				
8270E	mg/kg	Hexachlorobenzene	0.022	ND [0.055]	ND [0.044]	ND [0.051]	ND [0.050]	ND [0.045]
8270E	mg/kg	Hexachlorobutadiene	0.011	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	Hexachlorocyclopentadiene	NE	R	R	R	R	R
8270E	mg/kg	Hexachloroethane	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	Indeno(1,2,3-cd)pyrene	0.6	ND [0.013]	ND [0.011]	ND [0.012]	ND [0.012]	ND [0.011]
8270E	mg/kg	Isophorone	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Naphthalene	2.1	0.069 [0.022]	ND [0.018]	ND [0.021]	ND [0.020]	ND [0.018]
8270E	mg/kg	Nitrobenzene	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	n-Nitrosodimethylamine	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	n-Nitrosodi-n-propylamine	NE	ND [0.083]	ND [0.067]	ND [0.078]	ND [0.075]	ND [0.069]
8270E	mg/kg	n-Nitrosodiphenylamine	0.028	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Pentachlorophenol	0.4	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Phenanthrene	1.5	ND [0.013]	ND [0.011]	0.0067 [0.012] J	0.0067 [0.012] J	ND [0.011]
8270E	mg/kg	Phenol	0.42	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Pyrene	2.6	ND [0.011]	0.0075 [0.0089] J B	0.0096 [0.010] J B	ND [0.010]	ND [0.0092]
8270ESIM	mg/kg	1-Methylnaphthalene	NE	ND [0.0022]	0.00094 [0.0018] J	ND [0.0021]	ND [0.0020]	ND [0.0018]
8270ESIM	mg/kg	2-Methylnaphthalene	0.67	0.0026 [0.0044] J	ND [0.0036]	ND [0.0041]	ND [0.0040]	ND [0.0037]
8270ESIM	mg/kg	Acenaphthene	0.5	ND [0.0022]	0.0012 [0.0018] J	0.0012 [0.0021] J JN	ND [0.0020] JN	ND [0.0018]
8270ESIM	mg/kg	Acenaphthylene	0.56	0.00062 [0.0022] J	0.0010 [0.0018] J	0.00091 [0.0021] J JN	ND [0.0020] JN	ND [0.0018]
8270ESIM	mg/kg	Anthracene	0.96	0.0011 [0.0022] J B	0.0011 [0.0018] J B	0.0026 [0.0021] JN	0.0012 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(a)anthracene	1.3	ND [0.0022]	ND [0.0018]	0.0034 [0.0021] JN	0.0014 [0.0020] J	ND [0.0018]
8270ESIM	mg/kg	Benzo(a)pyrene	1.6	ND [0.0022]	0.00091 [0.0018] J B	0.0024 [0.0021] J JN	0.0012 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(b)fluoranthene	3.2	ND [0.0022]	0.0011 [0.0018] J B	0.0030 [0.0021] JN	0.0013 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(g,h,i)perylene	0.67	ND [0.0022]	0.0016 [0.0018] J	0.0024 [0.0021] J JN	0.0014 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(k)fluoranthene	3.2	ND [0.0022]	ND [0.0018]	0.0016 [0.0021] J	ND [0.0020]	ND [0.0018]
8270ESIM	mg/kg	Chrysene	1.4	0.00082 [0.0022] J B	ND [0.0018]	0.0030 [0.0021] JN	0.0017 [0.0020] J JN	0.00079 [0.0018] J B
8270ESIM	mg/kg	Dibenzo(a,h)anthracene	0.23	ND [0.0022]	ND [0.0018]	0.0011 [0.0021] J JN	ND [0.0020] JN	ND [0.0018]
8270ESIM	mg/kg	Fluoranthene	1.7	0.0024 [0.0022] J B	0.0020 [0.0018] J B	0.0073 [0.0021] JN	0.0043 [0.0020] JN	0.0016 [0.0018] J B
8270ESIM	mg/kg	Fluorene	0.54	0.0020 [0.0022] J	0.0012 [0.0018] J	0.0018 [0.0021] J	0.0012 [0.0020] J	ND [0.0018]
8270ESIM	mg/kg	Indeno(1,2,3-cd)pyrene	0.6	ND [0.0022]	ND [0.0018]	0.0021 [0.0021] J	ND [0.0020]	ND [0.0018]
8270ESIM	mg/kg	Naphthalene	2.1	0.062 [0.0044]	0.0038 [0.0036] J	ND [0.0041]	ND [0.0040]	ND [0.0037]
8270ESIM	mg/kg	Phenanthrene	1.5	0.0040 [0.0033] B	0.0018 [0.0027] J B	0.0058 [0.0031] B	0.0039 [0.0030] B	0.0015 [0.0028] J
8270ESIM	mg/kg	Pyrene	2.6	0.0023 [0.0022] J B	0.0038 [0.0018] B	0.0075 [0.0021] B	0.0059 [0.0020] B	0.0025 [0.0018] B
AK101	mg/kg	Gasoline Range Organics (C6-C10)	NE	ND [4.7] J-	ND [0.72]	ND [1.1] J-	ND [0.78] J-	ND [0.75]

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

				Sample ID Location ID	26-HOM-CW 11 SD 25-056	26-HOM-CW 12 SD 25-056	26-HOM-CW 21 SD 25-056	26-HOM-CW 20 SD 25-056	26-HOM-CW 22 SD 25-056
				Collection Date	10/11/2025 10:28	10/11/2025 10:50	10/08/2025 22:15	10/08/2025 22:00	10/09/2025 00:20
				Lab Sample ID Matrix	580-154935-12 Sediment	580-154935-13 Sediment	580-154935-4 Sediment	580-154935-3 Sediment	580-154935-5 Sediment
Method	Units	Analyte	DMMP	Dup of 21 SD					
D2216	PERCENT	Percent Moisture	NE	40.0 [1.0]	26.0 [1.0]	36.5 [1.0]	34.4 [1.0]	27.8 [1.0]	

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**Table 2: 2025 Homer Feasibility Study
Data Table
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		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	DMMP	MS/MSD ¹				
6020B	mg/kg	Antimony	150	0.42 [0.20]	0.43 [0.20]	0.53 [0.19]	0.34 [0.15]	0.30 [0.20]
6020B	mg/kg	Arsenic	57	10 [0.41]	9.9 [0.41]	15 [0.39]	10 [0.29]	11 [0.40]
6020B	mg/kg	Cadmium	5.1	0.098 [0.10] J	0.13 [0.10]	0.13 [0.097]	0.11 [0.073]	0.11 [0.10] J
6020B	mg/kg	Chromium	260	39 [0.49]	52 [0.49]	44 [0.47]	41 [0.35]	42 [0.48]
6020B	mg/kg	Copper	390	32 [0.46]	44 [0.46]	40 [0.44]	34 [0.33]	37 [0.45]
6020B	mg/kg	Lead	450	7.3 [0.20]	10 [0.20]	9.1 [0.19]	7.8 [0.15]	8.7 [0.20]
6020B	mg/kg	Nickel	NE	30 [0.48]	39 [0.49]	36 [0.46]	33 [0.35]	33 [0.48]
6020B	mg/kg	Selenium	3	0.20 [0.26] J	0.33 [0.26] J	0.28 [0.24] J	0.19 [0.18] J	0.22 [0.25] J
6020B	mg/kg	Silver	6.1	0.085 [0.10] J	0.12 [0.10] J	0.10 [0.097] J	0.085 [0.073] J	0.10 [0.10] J
6020B	mg/kg	Zinc	410	78 [10]	100 [10]	90 [9.7]	82 [7.3]	86 [10]
7471B	mg/kg	Mercury	0.41	0.095 [0.053]	0.12 [0.060]	0.11 [0.052]	0.098 [0.051]	0.11 [0.056]
8081B	mg/kg	4,4'-DDD	0.016	ND [0.011]	ND [0.012]	ND [0.011]	ND [0.011]	ND [0.012]
8081B	mg/kg	4,4'-DDE	0.009	ND [0.0092]	ND [0.011]	ND [0.0095]	ND [0.0092]	ND [0.010]
8081B	mg/kg	4,4'-DDT	0.012	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Aldrin	0.0095	ND [0.0047]	ND [0.0054]	ND [0.0049]	ND [0.0048]	ND [0.0052]
8081B	mg/kg	alpha-BHC	NE	ND [0.0050]	ND [0.0057]	ND [0.0052]	ND [0.0050]	ND [0.0055]
8081B	mg/kg	beta-BHC	NE	ND [0.0059]	ND [0.0068]	ND [0.0061]	ND [0.0059]	ND [0.0065]
8081B	mg/kg	cis-Chlordane	0.0028	ND [0.0061]	ND [0.0069]	ND [0.0063]	ND [0.0061]	ND [0.0066]
8081B	mg/kg	delta-BHC	NE	ND [0.0059]	ND [0.0068]	ND [0.0061]	ND [0.0059]	ND [0.0065]
8081B	mg/kg	Dieldrin	0.0019	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Endosulfan I	NE	ND [0.0040]	ND [0.0045]	ND [0.0041]	ND [0.0040]	ND [0.0043]
8081B	mg/kg	Endosulfan II	NE	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8081B	mg/kg	Endosulfan sulfate	NE	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Endrin	NE	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Endrin aldehyde	NE	ND [0.011]	ND [0.013]	ND [0.012]	ND [0.011]	ND [0.012]
8081B	mg/kg	Endrin ketone	NE	ND [0.012]	ND [0.014]	ND [0.012]	ND [0.012]	ND [0.013]
8081B	mg/kg	gamma-BHC (Lindane)	NE	ND [0.0040]	ND [0.0045]	ND [0.0041]	ND [0.0040]	ND [0.0043]
8081B	mg/kg	Heptachlor	0.0015	ND [0.0041]	ND [0.0047]	ND [0.0042]	ND [0.0041]	ND [0.0045]
8081B	mg/kg	Heptachlor Epoxide	NE	ND [0.0046]	ND [0.0053]	ND [0.0048]	ND [0.0046]	ND [0.0050]
8081B	mg/kg	Methoxychlor	NE	ND [0.039]	ND [0.044]	ND [0.040]	ND [0.039]	ND [0.043]
8081B	mg/kg	Toxaphene	NE	ND [0.18]	ND [0.21]	ND [0.19]	ND [0.18]	ND [0.20]
8081B	mg/kg	trans-Chlordane	0.0028	ND [0.0040]	ND [0.0045]	ND [0.0041]	ND [0.0040]	ND [0.0043]
8082A	mg/kg	PCB-1016 (Aroclor 1016)	0.13	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]

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		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	DMMP	MS/MSD ¹				
8082A	mg/kg	PCB-1221 (Aroclor 1221)	0.13	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1232 (Aroclor 1232)	0.13	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1242 (Aroclor 1242)	0.13	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1248 (Aroclor 1248)	0.13	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1254 (Aroclor 1254)	0.13	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1260 (Aroclor 1260)	0.13	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8260D	mg/kg	1,1,1-Trichloroethane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,1-Dichloroethane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,1-Dichloroethene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,1-Dichloropropene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2,3-Trichlorobenzene	NE	ND [0.35]	ND [0.35]	ND [0.43]	ND [0.28]	ND [0.44]
8260D	mg/kg	1,2,3-Trichloropropane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2,4-Trichlorobenzene	0.031	ND [0.35]	ND [0.35]	ND [0.43]	ND [0.28]	ND [0.44]
8260D	mg/kg	1,2,4-Trimethylbenzene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2-Dibromo-3-chloropropane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2-Dichlorobenzene	0.035	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2-Dichloropropane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,3,5-Trimethylbenzene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,3-Dichlorobenzene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,3-Dichloropropane	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	2,2-Dichloropropane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	2-Butanone	NE	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	2-Chlorotoluene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	4-Chlorotoluene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	4-Isopropyltoluene	NE	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	4-Methyl-2-pentanone	NE	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Acetone	NE	ND [0.70] J-	ND [0.70] J-	ND [0.85] J-	ND [0.57] J-	ND [0.88] J-
8260D	mg/kg	Bromobenzene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	Bromochloromethane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Carbon disulfide	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Carbon tetrachloride	NE	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Chlorobenzene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Chloroethane	NE	ND [0.17] J-	ND [0.17] J-	ND [0.21] J-	ND [0.14] J-	ND [0.22] J-
8260D	mg/kg	Chloromethane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	cis-1,2-Dichloroethene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]

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Method	Units	Analyte	DMMP	MS/MSD ¹				
8260D	mg/kg	Dichlorodifluoromethane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Ethylbenzene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	Isopropylbenzene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	Methylene chloride	NE	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Methyl-tert-butyl ether (MTBE)	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	n-Butylbenzene	NE	ND [0.22]	ND [0.22]	ND [0.27]	ND [0.18]	ND [0.27]
8260D	mg/kg	n-Propylbenzene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	o-Xylene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	sec-Butylbenzene	NE	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Styrene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	tert-Butylbenzene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Tetrachloroethene (PCE)	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Toluene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	trans-1,2-Dichloroethene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Trichloroethene (TCE)	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Trichlorofluoromethane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Vinyl chloride	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Xylene, isomers m & p	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8270E	mg/kg	1,2,4-Trichlorobenzene	0.031	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1,2-Dichlorobenzene	0.035	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1,3-Dichlorobenzene	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1,4-Dichlorobenzene	0.11	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1-Methylnaphthalene	NE	ND [0.0088]	ND [0.0099]	ND [0.0091] JN	ND [0.0089]	ND [0.0097]
8270E	mg/kg	2,2-oxybis(1-Chloropropane)	NE	ND [0.053]	ND [0.060]	ND [0.055]	ND [0.053]	ND [0.058]
8270E	mg/kg	2,4,5-Trichlorophenol	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	2,4,6-Trichlorophenol	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	2,4-Dichlorophenol	NE	ND [0.053]	ND [0.060]	ND [0.055] JN	ND [0.053]	ND [0.058]
8270E	mg/kg	2,4-Dimethylphenol	0.029	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	2,4-Dinitrophenol	NE	ND [1.1]	ND [1.3]	R	ND [1.1]	ND [1.2]
8270E	mg/kg	2,4-Dinitrotoluene	NE	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	2,6-Dinitrotoluene	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	2-Chloronaphthalene	NE	ND [0.035]	ND [0.040]	ND [0.037]	ND [0.035]	ND [0.039]
8270E	mg/kg	2-Chlorophenol	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	2-Methyl-4,6-dinitrophenol	NE	ND [0.44]	ND [0.50]	ND [0.46] J-	ND [0.44]	ND [0.48]
8270E	mg/kg	2-Methylnaphthalene	0.67	ND [0.013]	ND [0.015]	ND [0.014] JN	ND [0.013]	ND [0.014]

¹MS/MSD was run only on 8270E and 8270ESIM methods

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	DMMP	MS/MSD ¹				
8270E	mg/kg	2-Methylphenol (o-Cresol)	0.063	ND [0.053]	ND [0.060]	ND [0.055]	ND [0.053]	ND [0.058]
8270E	mg/kg	2-Nitroaniline	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	2-Nitrophenol	NE	ND [0.053]	ND [0.060]	ND [0.055] JN	ND [0.053]	ND [0.058]
8270E	mg/kg	3,3'-Dichlorobenzidine	NE	ND [0.35]	ND [0.40]	ND [0.37] JN	ND [0.35]	ND [0.39]
8270E	mg/kg	3-Methylphenol/4-Methylphenol Coelution	0.67	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	3-Nitroaniline	NE	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	4-Bromophenyl phenyl ether	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	4-Chloro-3-methylphenol	NE	ND [0.053]	ND [0.060]	ND [0.055]	ND [0.053]	ND [0.058]
8270E	mg/kg	4-Chloroaniline	NE	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	4-Chlorophenyl phenyl ether	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	4-Nitroaniline	NE	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	4-Nitrophenol	NE	ND [0.44]	ND [0.50]	ND [0.46]	ND [0.44]	ND [0.48]
8270E	mg/kg	Acenaphthene	0.5	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Acenaphthylene	0.56	ND [0.011]	ND [0.012]	ND [0.011]	ND [0.011]	ND [0.012]
8270E	mg/kg	Anthracene	0.96	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(a)anthracene	1.3	0.013 [0.018] J	ND [0.020]	ND [0.018] J-	ND [0.018]	ND [0.019]
8270E	mg/kg	Benzo(a)pyrene	1.6	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(b)fluoranthene	3.2	0.0096 [0.0088] J	ND [0.0099]	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(g,h,i)perylene	0.67	0.0069 [0.0088] J	ND [0.0099]	ND [0.0091] JN	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(k)fluoranthene	3.2	0.0061 [0.0088] J	ND [0.0099]	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzoic acid	0.65	ND [0.53]	ND [0.60]	R	ND [0.53]	ND [0.58]
8270E	mg/kg	Benzyl alcohol	0.057	ND [0.44]	ND [0.50]	ND [0.46]	ND [0.44]	ND [0.48]
8270E	mg/kg	Benzyl butyl phthalate	0.063	ND [0.18]	ND [0.20]	ND [0.18]	ND [0.18]	ND [0.19]
8270E	mg/kg	bis-(2-Chloroethoxy)methane	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	bis-(2-Chloroethyl)ether	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	bis-(2-Ethylhexyl)phthalate	1.3	ND [0.18]	ND [0.20]	ND [0.18]	ND [0.18]	ND [0.19]
8270E	mg/kg	Carbazole	NE	ND [0.044]	ND [0.050]	ND [0.046] J-	ND [0.044]	ND [0.048]
8270E	mg/kg	Chrysene	1.4	0.011 [0.0088] J	0.0066 [0.0099] J	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Dibenzo(a,h)anthracene	0.23	ND [0.018]	ND [0.020]	ND [0.018]	ND [0.018]	ND [0.019]
8270E	mg/kg	Dibenzofuran	0.54	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Diethyl phthalate	0.2	ND [0.18]	ND [0.20]	ND [0.18] J-	ND [0.18]	ND [0.19]
8270E	mg/kg	Dimethyl phthalate	0.071	ND [0.18]	ND [0.20]	ND [0.18] J-	ND [0.18]	ND [0.19]
8270E	mg/kg	Di-n-butyl phthalate	1.4	ND [0.18]	ND [0.20]	ND [0.18] JN	ND [0.18]	ND [0.19]
8270E	mg/kg	Di-n-octyl phthalate	6.2	ND [0.18]	ND [0.20]	ND [0.18] J-	ND [0.18]	ND [0.19]
8270E	mg/kg	Fluoranthene	1.7	0.019 [0.0088] J B	0.0088 [0.0099] J B	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Fluorene	0.54	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]

¹MS/MSD was run only on 8270E and 8270ESIM methods

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	DMMP	MS/MSD ¹				
8270E	mg/kg	Hexachlorobenzene	0.022	ND [0.044]	ND [0.049]	ND [0.045]	ND [0.044]	ND [0.048]
8270E	mg/kg	Hexachlorobutadiene	0.011	ND [0.053]	ND [0.060]	ND [0.055] JN	ND [0.053]	ND [0.058]
8270E	mg/kg	Hexachlorocyclopentadiene	NE	R	R	R	R	R
8270E	mg/kg	Hexachloroethane	NE	ND [0.088]	ND [0.099]	ND [0.091] J-	ND [0.089]	ND [0.097]
8270E	mg/kg	Indeno(1,2,3-cd)pyrene	0.6	0.0086 [0.011] J	ND [0.012]	ND [0.011]	0.0060 [0.011] J	ND [0.012]
8270E	mg/kg	Isophorone	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Naphthalene	2.1	ND [0.018]	ND [0.020]	ND [0.018] J-	ND [0.018]	ND [0.019]
8270E	mg/kg	Nitrobenzene	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	n-Nitrosodimethylamine	NE	ND [0.088]	ND [0.099]	ND [0.091] JN	ND [0.089]	ND [0.097]
8270E	mg/kg	n-Nitrosodi-n-propylamine	NE	ND [0.066]	ND [0.074]	ND [0.069] JN	ND [0.066]	ND [0.072]
8270E	mg/kg	n-Nitrosodiphenylamine	0.028	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Pentachlorophenol	0.4	ND [0.18]	ND [0.20]	ND [0.18]	ND [0.18]	ND [0.19]
8270E	mg/kg	Phenanthrene	1.5	0.010 [0.011] J	ND [0.012]	ND [0.011] J-	ND [0.011]	ND [0.012]
8270E	mg/kg	Phenol	0.42	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Pyrene	2.6	0.015 [0.0088] J B	0.011 [0.0099] J B	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270ESIM	mg/kg	1-Methylnaphthalene	NE	ND [0.018]	ND [0.0020]	ND [0.018] JN	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	2-Methylnaphthalene	0.67	ND [0.035]	ND [0.0040]	ND [0.037]	ND [0.0035]	ND [0.0039]
8270ESIM	mg/kg	Acenaphthene	0.5	ND [0.018]	ND [0.0020]	ND [0.018] JN	0.00096 [0.0018] J	ND [0.0019]
8270ESIM	mg/kg	Acenaphthylene	0.56	ND [0.018]	ND [0.0020]	ND [0.018] JN	0.00052 [0.0018] J	ND [0.0019]
8270ESIM	mg/kg	Anthracene	0.96	ND [0.018]	0.0010 [0.0020] J B	ND [0.018]	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(a)anthracene	1.3	ND [0.018]	0.0036 [0.0020] B	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(a)pyrene	1.6	ND [0.018]	0.0021 [0.0020] J B	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(b)fluoranthene	3.2	0.012 [0.018] J	0.0029 [0.0020] B	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(g,h,i)perylene	0.67	ND [0.018]	0.0017 [0.0020] J	ND [0.018] JN	0.0010 [0.0018] J	0.00098 [0.0019] J
8270ESIM	mg/kg	Benzo(k)fluoranthene	3.2	ND [0.018]	0.0015 [0.0020] J	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Chrysene	1.4	0.0082 [0.018] J B	0.0030 [0.0020] B	ND [0.018] J-	ND [0.0018]	0.00094 [0.0019] J B
8270ESIM	mg/kg	Dibenzo(a,h)anthracene	0.23	ND [0.018]	ND [0.0020]	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Fluoranthene	1.7	0.017 [0.018] J	0.0068 [0.0020] B	ND [0.018] JN	0.0020 [0.0018] J B	0.0023 [0.0019] J B
8270ESIM	mg/kg	Fluorene	0.54	ND [0.018]	ND [0.0020]	ND [0.018]	0.00091 [0.0018] J	ND [0.0019]
8270ESIM	mg/kg	Indeno(1,2,3-cd)pyrene	0.6	ND [0.018]	0.0014 [0.0020] J	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Naphthalene	2.1	ND [0.035]	ND [0.0040]	ND [0.037]	ND [0.0035]	ND [0.0039]
8270ESIM	mg/kg	Phenanthrene	1.5	ND [0.026]	0.0023 [0.0030] J B	ND [0.027]	ND [0.0027]	0.0019 [0.0029] J B
8270ESIM	mg/kg	Pyrene	2.6	0.017 [0.018] J	0.0072 [0.0020] B	ND [0.018]	0.0027 [0.0018] B	0.0032 [0.0019] B
AK101	mg/kg	Gasoline Range Organics (C6-C10)	NE	ND [0.75]	ND [0.75] J-	ND [0.92]	ND [0.61] J-	ND [0.95]

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Data Table
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				Sample ID	26-HOM-CW 23 SD	26-HOM-CW 31 SD	26-HOM-CW 32 SD	26-HOM-CW 33 SD	26-HOM-CW 81 SD
				Location ID	25-056	25-056	25-056	25-056	25-056
				Collection Date	10/09/2025 00:40	10/10/2025 20:13	10/10/2025 21:06	10/10/2025 21:15	10/07/2025 15:15
				Lab Sample ID	580-154935-6	580-154935-9	580-154935-10	580-154935-11	580-154935-1
				Matrix	Sediment	Sediment	Sediment	Sediment	Sediment
Method	Units	Analyte	DMMP	MS/MSD ¹					
D2216	PERCENT	Percent Moisture	NE	24.6 [1.0]	33.6 [1.0]	27.3 [1.0]	25.0 [1.0]	31.3 [1.0]	

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**Table 2: 2025 Homer Feasibility Study
Data Table
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				Sample ID	26-HOM-CW 82 SD	26-HOM-CW 91 SD	26-HOM-CW 90 SD	TB 1
				Location ID	25-056	25-056	25-056	25-056
				Collection Date	10/07/2025 16:15	10/09/2025 11:40	10/09/2025 11:30	10/06/2025 00:00
				Lab Sample ID	580-154935-2	580-154935-8	580-154935-7	580-154935-14
				Matrix	Sediment	Sediment	Sediment	Soil
Method	Units	Analyte	DMMP	Dup of 91 SD				
6020B	mg/kg	Antimony	150	0.21 [0.16]	0.26 [0.16]	0.17 [0.15]	J	
6020B	mg/kg	Arsenic	57	7.8 [0.31]	13 [0.33]	12 [0.30]		
6020B	mg/kg	Cadmium	5.1	0.10 [0.079]	0.057 [0.082]	ND [0.075]		
6020B	mg/kg	Chromium	260	47 [0.38]	22 [0.39]	22 [0.36]		
6020B	mg/kg	Copper	390	40 [0.35]	15 [0.37]	15 [0.34]		
6020B	mg/kg	Lead	450	9.0 [0.16]	4.4 [0.16]	4.6 [0.15]		
6020B	mg/kg	Nickel	NE	36 [0.37]	16 [0.39]	18 [0.36]		
6020B	mg/kg	Selenium	3	0.21 [0.20]	0.14 [0.20]	0.15 [0.19]	J	
6020B	mg/kg	Silver	6.1	0.092 [0.079]	0.046 [0.082]	0.038 [0.075]	J	
6020B	mg/kg	Zinc	410	94 [7.9]	42 [8.2]	46 [7.5]		
7471B	mg/kg	Mercury	0.41	0.11 [0.055]	0.049 [0.049]	0.040 [0.046]	J	
8081B	mg/kg	4,4'-DDD	0.016	ND [0.011]	ND [0.021]	ND [0.0095]		
8081B	mg/kg	4,4'-DDE	0.009	ND [0.0096]	ND [0.018]	ND [0.0083]		
8081B	mg/kg	4,4'-DDT	0.012	ND [0.0082]	ND [0.016]	ND [0.0071]		
8081B	mg/kg	Aldrin	0.0095	ND [0.0049]	ND [0.0093]	ND [0.0043]		
8081B	mg/kg	alpha-BHC	NE	ND [0.0052]	ND [0.0098]	ND [0.0045]		
8081B	mg/kg	beta-BHC	NE	ND [0.0062]	ND [0.012]	ND [0.0053]		
8081B	mg/kg	cis-Chlordane	0.0028	ND [0.0063]	ND [0.012]	ND [0.0055]		
8081B	mg/kg	delta-BHC	NE	ND [0.0062]	ND [0.012]	ND [0.0053]		
8081B	mg/kg	Dieldrin	0.0019	ND [0.0082]	ND [0.016]	ND [0.0071]		
8081B	mg/kg	Endosulfan I	NE	ND [0.0041]	ND [0.0078]	ND [0.0036]		
8081B	mg/kg	Endosulfan II	NE	ND [0.015]	ND [0.028]	ND [0.013]		
8081B	mg/kg	Endosulfan sulfate	NE	ND [0.0082]	ND [0.016]	ND [0.0071]		
8081B	mg/kg	Endrin	NE	ND [0.0082]	ND [0.016]	ND [0.0071]		
8081B	mg/kg	Endrin aldehyde	NE	ND [0.012]	ND [0.022]	ND [0.010]		
8081B	mg/kg	Endrin ketone	NE	ND [0.012]	ND [0.023]	ND [0.011]		
8081B	mg/kg	gamma-BHC (Lindane)	NE	ND [0.0041]	ND [0.0078]	ND [0.0036]		
8081B	mg/kg	Heptachlor	0.0015	ND [0.0043]	ND [0.0080]	ND [0.0037]		
8081B	mg/kg	Heptachlor Epoxide	NE	ND [0.0048]	ND [0.0090]	ND [0.0042]		
8081B	mg/kg	Methoxychlor	NE	ND [0.041]	ND [0.076]	ND [0.035]		
8081B	mg/kg	Toxaphene	NE	ND [0.19]	ND [0.36]	ND [0.17]		
8081B	mg/kg	trans-Chlordane	0.0028	ND [0.0041]	ND [0.0078]	ND [0.0036]		
8082A	mg/kg	PCB-1016 (Aroclor 1016)	0.13	ND [0.015]	ND [0.014]	ND [0.013]		

¹MS/MSD was run only on 8270E and 8270ESIM methods

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Solid shade indicates screening value exceedance

NE - Not Established

**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 82 SD 25-056 10/07/2025 16:15 580-154935-2 Sediment	26-HOM-CW 91 SD 25-056 10/09/2025 11:40 580-154935-8 Sediment	26-HOM-CW 90 SD 25-056 10/09/2025 11:30 580-154935-7 Sediment	TB 1 25-056 10/06/2025 00:00 580-154935-14 Soil	
Method	Units	Analyte	DMMP	Dup of 91 SD			
8082A	mg/kg	PCB-1221 (Aroclor 1221)	0.13	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1232 (Aroclor 1232)	0.13	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1242 (Aroclor 1242)	0.13	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1248 (Aroclor 1248)	0.13	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1254 (Aroclor 1254)	0.13	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1260 (Aroclor 1260)	0.13	ND [0.015]	ND [0.014]	ND [0.013]	
8260D	mg/kg	1,1,1-Trichloroethane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,1-Dichloroethane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,1-Dichloroethene	NE	ND [0.082]	ND [0.070]	ND [0.065]	
8260D	mg/kg	1,1-Dichloropropene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2,3-Trichlorobenzene	NE	ND [0.33]	ND [0.28]	ND [0.26]	ND [0.40] J-
8260D	mg/kg	1,2,3-Trichloropropane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2,4-Trichlorobenzene	0.031	ND [0.33]	ND [0.28]	ND [0.26]	ND [0.40] J-
8260D	mg/kg	1,2,4-Trimethylbenzene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2-Dibromo-3-chloropropane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2-Dichlorobenzene	0.035	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2-Dichloropropane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,3,5-Trimethylbenzene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,3-Dichlorobenzene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,3-Dichloropropane	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	2,2-Dichloropropane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	2-Butanone	NE	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	2-Chlorotoluene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	4-Chlorotoluene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	4-Isopropyltoluene	NE	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	4-Methyl-2-pentanone	NE	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	Acetone	NE	ND [0.65] J-	ND [0.56] J-	ND [0.52] J-	ND [0.80] J-
8260D	mg/kg	Bromobenzene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	Bromochloromethane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Carbon disulfide	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Carbon tetrachloride	NE	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	Chlorobenzene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Chloroethane	NE	ND [0.16] J-	ND [0.14] J-	ND [0.13] J-	ND [0.20] J-
8260D	mg/kg	Chloromethane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	cis-1,2-Dichloroethene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

				Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 82 SD 25-056 10/07/2025 16:15 580-154935-2 Sediment	26-HOM-CW 91 SD 25-056 10/09/2025 11:40 580-154935-8 Sediment	26-HOM-CW 90 SD 25-056 10/09/2025 11:30 580-154935-7 Sediment	TB 1 25-056 10/06/2025 00:00 580-154935-14 Soil
Method	Units	Analyte	DMMP	Dup of 91 SD				
8260D	mg/kg	Dichlorodifluoromethane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10]	J-
8260D	mg/kg	Ethylbenzene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050]	J-
8260D	mg/kg	Isopropylbenzene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050]	J-
8260D	mg/kg	Methylene chloride	NE	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20]	J-
8260D	mg/kg	Methyl-tert-butyl ether (MTBE)	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10]	J-
8260D	mg/kg	n-Butylbenzene	NE	ND [0.20]	ND [0.18]	ND [0.16]	ND [0.25]	J-
8260D	mg/kg	n-Propylbenzene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050]	J-
8260D	mg/kg	o-Xylene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050]	J-
8260D	mg/kg	sec-Butylbenzene	NE	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20]	J-
8260D	mg/kg	Styrene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050]	J-
8260D	mg/kg	tert-Butylbenzene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10]	J-
8260D	mg/kg	Tetrachloroethene (PCE)	NE	ND [0.082]	ND [0.070]	ND [0.065]		
8260D	mg/kg	Toluene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10]	J-
8260D	mg/kg	trans-1,2-Dichloroethene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10]	J-
8260D	mg/kg	Trichloroethene (TCE)	NE	ND [0.082]	ND [0.070]	ND [0.065]		
8260D	mg/kg	Trichlorofluoromethane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10]	J-
8260D	mg/kg	Vinyl chloride	NE	ND [0.082]	ND [0.070]	ND [0.065]		
8260D	mg/kg	Xylene, isomers m & p	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10]	J-
8270E	mg/kg	1,2,4-Trichlorobenzene	0.031	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	1,2-Dichlorobenzene	0.035	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	1,3-Dichlorobenzene	NE	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	1,4-Dichlorobenzene	0.11	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	1-Methylnaphthalene	NE	ND [0.0092]	ND [0.0086]	ND [0.0079]		
8270E	mg/kg	2,2-oxybis(1-Chloropropane)	NE	ND [0.055]	ND [0.052]	ND [0.047]		
8270E	mg/kg	2,4,5-Trichlorophenol	NE	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	2,4,6-Trichlorophenol	NE	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	2,4-Dichlorophenol	NE	ND [0.055]	ND [0.052]	ND [0.047]		
8270E	mg/kg	2,4-Dimethylphenol	0.029	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	2,4-Dinitrophenol	NE	ND [1.2]	ND [1.1]	ND [0.99]		
8270E	mg/kg	2,4-Dinitrotoluene	NE	ND [0.092]	ND [0.086]	ND [0.079]		
8270E	mg/kg	2,6-Dinitrotoluene	NE	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	2-Chloronaphthalene	NE	ND [0.037]	ND [0.034]	ND [0.032]		
8270E	mg/kg	2-Chlorophenol	NE	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	2-Methyl-4,6-dinitrophenol	NE	ND [0.46]	ND [0.43]	ND [0.40]		
8270E	mg/kg	2-Methylnaphthalene	0.67	ND [0.014]	ND [0.013]	ND [0.012]		

¹MS/MSD was run only on 8270E and 8270ESIM methods

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 82 SD 25-056 10/07/2025 16:15 580-154935-2 Sediment	26-HOM-CW 91 SD 25-056 10/09/2025 11:40 580-154935-8 Sediment	26-HOM-CW 90 SD 25-056 10/09/2025 11:30 580-154935-7 Sediment	TB 1 25-056 10/06/2025 00:00 580-154935-14 Soil
Method	Units	Analyte	DMMP	Dup of 91 SD		
8270E	mg/kg	2-Methylphenol (o-Cresol)	0.063	ND [0.055]	ND [0.052]	ND [0.047]
8270E	mg/kg	2-Nitroaniline	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	2-Nitrophenol	NE	ND [0.055]	ND [0.052]	ND [0.047]
8270E	mg/kg	3,3'-Dichlorobenzidine	NE	ND [0.37]	ND [0.34]	ND [0.32]
8270E	mg/kg	3-Methylphenol/4-Methylphenol Coelution	0.67	ND [0.046]	0.21 [0.043]	ND [0.040]
8270E	mg/kg	3-Nitroaniline	NE	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	4-Bromophenyl phenyl ether	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	4-Chloro-3-methylphenol	NE	ND [0.055]	ND [0.052]	ND [0.047]
8270E	mg/kg	4-Chloroaniline	NE	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	4-Chlorophenyl phenyl ether	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	4-Nitroaniline	NE	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	4-Nitrophenol	NE	ND [0.46]	ND [0.43]	ND [0.40]
8270E	mg/kg	Acenaphthene	0.5	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Acenaphthylene	0.56	ND [0.011]	ND [0.010]	ND [0.0095]
8270E	mg/kg	Anthracene	0.96	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(a)anthracene	1.3	ND [0.018]	ND [0.017]	ND [0.016]
8270E	mg/kg	Benzo(a)pyrene	1.6	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(b)fluoranthene	3.2	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(g,h,i)perylene	0.67	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(k)fluoranthene	3.2	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzoic acid	0.65	ND [0.55]	ND [0.52]	ND [0.47]
8270E	mg/kg	Benzyl alcohol	0.057	ND [0.46]	ND [0.43]	ND [0.40]
8270E	mg/kg	Benzyl butyl phthalate	0.063	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	bis-(2-Chloroethoxy)methane	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	bis-(2-Chloroethyl)ether	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	bis-(2-Ethylhexyl)phthalate	1.3	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Carbazole	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	Chrysene	1.4	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Dibenzo(a,h)anthracene	0.23	ND [0.018]	ND [0.017]	ND [0.016]
8270E	mg/kg	Dibenzofuran	0.54	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	Diethyl phthalate	0.2	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Dimethyl phthalate	0.071	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Di-n-butyl phthalate	1.4	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Di-n-octyl phthalate	6.2	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Fluoranthene	1.7	ND [0.0092]	0.0053 [0.0086] J B	ND [0.0079]
8270E	mg/kg	Fluorene	0.54	ND [0.0092]	ND [0.0086]	ND [0.0079]

¹MS/MSD was run only on 8270E and 8270ESIM methods

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

				Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 82 SD 25-056 10/07/2025 16:15 580-154935-2 Sediment	26-HOM-CW 91 SD 25-056 10/09/2025 11:40 580-154935-8 Sediment	26-HOM-CW 90 SD 25-056 10/09/2025 11:30 580-154935-7 Sediment	TB 1 25-056 10/06/2025 00:00 580-154935-14 Soil
Method	Units	Analyte	DMMP	Dup of 91 SD				
8270E	mg/kg	Hexachlorobenzene	0.022	ND [0.045]	ND [0.043]	ND [0.039]		
8270E	mg/kg	Hexachlorobutadiene	0.011	ND [0.055]	ND [0.052]	ND [0.047]		
8270E	mg/kg	Hexachlorocyclopentadiene	NE	R	R	R		
8270E	mg/kg	Hexachloroethane	NE	ND [0.092]	ND [0.086]	ND [0.079]		
8270E	mg/kg	Indeno(1,2,3-cd)pyrene	0.6	ND [0.011]	ND [0.010]	ND [0.0095]		
8270E	mg/kg	Isophorone	NE	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	Naphthalene	2.1	ND [0.018]	ND [0.017]	ND [0.016]		
8270E	mg/kg	Nitrobenzene	NE	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	n-Nitrosodimethylamine	NE	ND [0.092]	ND [0.086]	ND [0.079]		
8270E	mg/kg	n-Nitrosodi-n-propylamine	NE	ND [0.069]	ND [0.064]	ND [0.059]		
8270E	mg/kg	n-Nitrosodiphenylamine	0.028	ND [0.046]	ND [0.043]	ND [0.040]		
8270E	mg/kg	Pentachlorophenol	0.4	ND [0.18]	ND [0.17]	ND [0.16]		
8270E	mg/kg	Phenanthrene	1.5	ND [0.011]	ND [0.010]	ND [0.0095]		
8270E	mg/kg	Phenol	0.42	ND [0.046]	0.062 [0.043]	ND [0.040]		
8270E	mg/kg	Pyrene	2.6	ND [0.0092]	0.0067 [0.0086] J B	0.0055 [0.0079] J B		
8270ESIM	mg/kg	1-Methylnaphthalene	NE	ND [0.0018]	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	2-Methylnaphthalene	0.67	ND [0.0037]	ND [0.0034]	ND [0.0032]		
8270ESIM	mg/kg	Acenaphthene	0.5	ND [0.0018]	0.00087 [0.0017] J JN	ND [0.0016] JN		
8270ESIM	mg/kg	Acenaphthylene	0.56	ND [0.0018]	0.00092 [0.0017] J JN	ND [0.0016] JN		
8270ESIM	mg/kg	Anthracene	0.96	ND [0.0018]	0.0016 [0.0017] J JN	0.00088 [0.0016] J JN		
8270ESIM	mg/kg	Benzo(a)anthracene	1.3	ND [0.0018]	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	Benzo(a)pyrene	1.6	ND [0.0018]	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	Benzo(b)fluoranthene	3.2	ND [0.0018]	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	Benzo(g,h,i)perylene	0.67	0.0013 [0.0018] J	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	Benzo(k)fluoranthene	3.2	ND [0.0018]	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	Chrysene	1.4	0.00078 [0.0018] J B	0.0013 [0.0017] J B	0.00055 [0.0016] J B		
8270ESIM	mg/kg	Dibenzo(a,h)anthracene	0.23	ND [0.0018]	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	Fluoranthene	1.7	0.0015 [0.0018] J B	0.0034 [0.0017] JN	0.0018 [0.0016] J JN		
8270ESIM	mg/kg	Fluorene	0.54	ND [0.0018]	0.0012 [0.0017] J	ND [0.0016]		
8270ESIM	mg/kg	Indeno(1,2,3-cd)pyrene	0.6	ND [0.0018]	ND [0.0017]	ND [0.0016]		
8270ESIM	mg/kg	Naphthalene	2.1	ND [0.0037]	0.0024 [0.0034] J	ND [0.0032]		
8270ESIM	mg/kg	Phenanthrene	1.5	0.0014 [0.0028] J B	0.0029 [0.0026] J JN	0.0015 [0.0024] J JN		
8270ESIM	mg/kg	Pyrene	2.6	0.0023 [0.0018] B	0.0043 [0.0017] B	0.0017 [0.0016] J B		
AK101	mg/kg	Gasoline Range Organics (C6-C10)	NE	ND [0.70] J-	ND [0.61]	ND [0.56]		ND [0.86]

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**Table 2: 2025 Homer Feasibility Study
Data Table
Comparison to DMMP Criteria**

				Sample ID	26-HOM-CW 82 SD	26-HOM-CW 91 SD	26-HOM-CW 90 SD	TB 1
				Location ID	25-056	25-056	25-056	25-056
				Collection Date	10/07/2025 16:15	10/09/2025 11:40	10/09/2025 11:30	10/06/2025 00:00
				Lab Sample ID	580-154935-2	580-154935-8	580-154935-7	580-154935-14
				Matrix	Sediment	Sediment	Sediment	Soil
Method	Units	Analyte	DMMP	Dup of 91 SD				
D2216	PERCENT	Percent Moisture	NE	27.8 [1.0]	22.8 [1.0]	16.4 [1.0]		

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**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	ADEC	Dup of 21 SD				
6020B	mg/kg	Antimony	4.6	0.37 [0.24]	0.39 [0.15]	0.44 [0.19]	0.34 [0.20]	0.37 [0.20]
6020B	mg/kg	Arsenic	0.2	14 [0.48]	13 [0.29]	10 [0.38]	11 [0.40]	9.2 [0.39]
6020B	mg/kg	Cadmium	9.1	0.079 [0.12] J	0.097 [0.073]	0.095 [0.095] J	0.095 [0.099] J	0.11 [0.098] J
6020B	mg/kg	Chromium	100000	55 [0.57]	42 [0.35]	53 [0.45]	47 [0.47]	40 [0.47]
6020B	mg/kg	Copper	370	49 [0.54]	30 [0.33]	47 [0.43]	42 [0.45]	31 [0.44]
6020B	mg/kg	Lead	400	12 [0.24]	7.2 [0.15]	11 [0.19]	9.9 [0.20]	7.4 [0.20]
6020B	mg/kg	Nickel	340	41 [0.57]	31 [0.35]	41 [0.45]	38 [0.47]	32 [0.47]
6020B	mg/kg	Selenium	6.9	0.30 [0.30] J	0.18 [0.18] J	0.29 [0.24] J	0.26 [0.25] J	0.20 [0.25] J
6020B	mg/kg	Silver	11	0.13 [0.12] J	0.085 [0.073] J	0.13 [0.095]	0.13 [0.099]	0.088 [0.098] J
6020B	mg/kg	Zinc	4900	110 [12]	79 [7.3]	110 [9.5]	98 [9.9]	83 [9.8]
7471B	mg/kg	Mercury	0.36	0.14 [0.063]	0.084 [0.052]	0.13 [0.062]	0.12 [0.057]	0.098 [0.054]
8081B	mg/kg	4,4'-DDD	0.098	ND [0.013]	ND [0.011]	ND [0.013]	ND [0.012]	ND [0.011]
8081B	mg/kg	4,4'-DDE	0.72	ND [0.012]	ND [0.0094]	ND [0.011]	ND [0.011]	ND [0.0096]
8081B	mg/kg	4,4'-DDT	5.1	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Aldrin	0.0099	ND [0.0059]	ND [0.0048]	ND [0.0057]	ND [0.0054]	ND [0.0049]
8081B	mg/kg	alpha-BHC	0.0029	ND [0.0063]	ND [0.0051]	ND [0.0060]	ND [0.0057]	ND [0.0052]
8081B	mg/kg	beta-BHC	0.01	ND [0.0074]	ND [0.0060]	ND [0.0071]	ND [0.0068]	ND [0.0062]
8081B	mg/kg	cis-Chlordane	0.18	ND [0.0076]	ND [0.0062]	ND [0.0072]	ND [0.0069]	ND [0.0063]
8081B	mg/kg	delta-BHC	NE	ND [0.0074]	ND [0.0060]	ND [0.0071]	ND [0.0068]	ND [0.0062]
8081B	mg/kg	Dieldrin	0.0047	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Endosulfan I	9.3	ND [0.0049]	ND [0.0040]	ND [0.0047]	ND [0.0045]	ND [0.0041]
8081B	mg/kg	Endosulfan II	9.3	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8081B	mg/kg	Endosulfan sulfate	9.3	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Endrin	0.61	ND [0.0099]	ND [0.0080]	ND [0.0094]	ND [0.0090]	ND [0.0082]
8081B	mg/kg	Endrin aldehyde	NE	ND [0.014]	ND [0.011]	ND [0.013]	ND [0.013]	ND [0.012]
8081B	mg/kg	Endrin ketone	NE	ND [0.015]	ND [0.012]	ND [0.014]	ND [0.014]	ND [0.012]
8081B	mg/kg	gamma-BHC (Lindane)	0.016	ND [0.0049]	ND [0.0040]	ND [0.0047]	ND [0.0045]	ND [0.0041]
8081B	mg/kg	Heptachlor	0.0076	ND [0.0051]	ND [0.0042]	ND [0.0049]	ND [0.0047]	ND [0.0043]
8081B	mg/kg	Heptachlor Epoxide	0.0019	ND [0.0058]	ND [0.0047]	ND [0.0055]	ND [0.0053]	ND [0.0048]
8081B	mg/kg	Methoxychlor	13	ND [0.049]	ND [0.040]	ND [0.046]	ND [0.044]	ND [0.040]
8081B	mg/kg	Toxaphene	0.72	ND [0.23]	ND [0.19]	ND [0.22]	ND [0.21]	ND [0.19]
8081B	mg/kg	trans-Chlordane	0.18	ND [0.0049]	ND [0.0040]	ND [0.0047]	ND [0.0045]	ND [0.0041]
8082A	mg/kg	PCB-1016 (Aroclor 1016)	1	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

[] - Laboratory LOD

Solid shade indicates screening value exceedance

NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	ADEC	Dup of 21 SD				
8082A	mg/kg	PCB-1221 (Aroclor 1221)	1	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1232 (Aroclor 1232)	1	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1242 (Aroclor 1242)	1	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1248 (Aroclor 1248)	1	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1254 (Aroclor 1254)	1	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8082A	mg/kg	PCB-1260 (Aroclor 1260)	1	ND [0.018]	ND [0.015]	ND [0.017]	ND [0.017]	ND [0.015]
8260D	mg/kg	1,1,1-Trichloroethane	32	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,1-Dichloroethane	0.092	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,1-Dichloroethene	1.2	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,1-Dichloropropene	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2,3-Trichlorobenzene	0.15	ND [5.5]	ND [0.33] J-	ND [0.50]	ND [0.36]	ND [0.35]
8260D	mg/kg	1,2,3-Trichloropropane	0.000031	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2,4-Trichlorobenzene	0.082	ND [5.5]	ND [0.33] J-	ND [0.50]	ND [0.36]	ND [0.35]
8260D	mg/kg	1,2,4-Trimethylbenzene	0.61	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2-Dibromo-3-chloropropane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2-Dichlorobenzene	2.4	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,2-Dichloropropane	0.03	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,3,5-Trimethylbenzene	0.66	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,3-Dichlorobenzene	2.3	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	1,3-Dichloropropane	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	2,2-Dichloropropane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	2-Butanone	15	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	2-Chlorotoluene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	4-Chlorotoluene	NE	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	4-Isopropyltoluene	NE	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	4-Methyl-2-pentanone	18	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Acetone	38	ND [11] J-	ND [0.67] J-	ND [1.0] J-	ND [0.72] J-	ND [0.69] J-
8260D	mg/kg	Bromobenzene	0.36	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	Bromochloromethane	NE	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Carbon disulfide	2.9	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Carbon tetrachloride	0.021	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Chlorobenzene	0.46	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Chloroethane	72	ND [2.7] J-	ND [0.17] J-	ND [0.25] J-	ND [0.18] J-	ND [0.17] J-
8260D	mg/kg	Chloromethane	0.61	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	cis-1,2-Dichloroethene	0.12	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]

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ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

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**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	ADEC	Dup of 21 SD				
8260D	mg/kg	Dichlorodifluoromethane	3.9	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Ethylbenzene	0.13	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	Isopropylbenzene	5.6	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	Methylene chloride	0.33	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Methyl-tert-butyl ether (MTBE)	0.4	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	n-Butylbenzene	20	ND [3.4]	ND [0.21] J-	ND [0.31]	ND [0.23]	ND [0.22]
8260D	mg/kg	n-Propylbenzene	9.1	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	o-Xylene	1.5	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	sec-Butylbenzene	28	ND [2.7]	ND [0.17] J-	ND [0.25]	ND [0.18]	ND [0.17]
8260D	mg/kg	Styrene	10	ND [0.68]	ND [0.042] J-	ND [0.063]	ND [0.045]	ND [0.043]
8260D	mg/kg	tert-Butylbenzene	11	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Tetrachloroethene (PCE)	0.19	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Toluene	6.7	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	trans-1,2-Dichloroethene	1.3	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Trichloroethene (TCE)	0.011	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Trichlorofluoromethane	41	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Vinyl chloride	0.0008	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8260D	mg/kg	Xylene, isomers m & p	1.5	ND [1.4]	ND [0.083] J-	ND [0.13]	ND [0.090]	ND [0.087]
8270E	mg/kg	1,2,4-Trichlorobenzene	0.082	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1,2-Dichlorobenzene	2.4	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1,3-Dichlorobenzene	2.3	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1,4-Dichlorobenzene	0.037	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	1-Methylnaphthalene	0.41	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	2,2-oxybis(1-Chloropropane)	NE	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	2,4,5-Trichlorophenol	28	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2,4,6-Trichlorophenol	0.092	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2,4-Dichlorophenol	0.21	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	2,4-Dimethylphenol	3.2	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2,4-Dinitrophenol	0.34	ND [1.4]	ND [1.1]	ND [1.3]	ND [1.3]	ND [1.2]
8270E	mg/kg	2,4-Dinitrotoluene	0.024	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	2,6-Dinitrotoluene	0.005	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2-Chloronaphthalene	26	ND [0.044]	ND [0.036]	ND [0.041]	ND [0.040]	ND [0.037]
8270E	mg/kg	2-Chlorophenol	0.71	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2-Methyl-4,6-dinitrophenol	NE	ND [0.55]	ND [0.45]	ND [0.52]	ND [0.50]	ND [0.46]
8270E	mg/kg	2-Methylnaphthalene	1.3	ND [0.017]	ND [0.013]	ND [0.016]	ND [0.015]	ND [0.014]

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ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

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Solid shade indicates screening value exceedance

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Method	Units	Analyte	ADEC	Dup of 21 SD				
8270E	mg/kg	2-Methylphenol (o-Cresol)	6.2	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	2-Nitroaniline	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	2-Nitrophenol	NE	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	3,3'-Dichlorobenzidine	0.056	ND [0.44]	ND [0.36]	ND [0.41]	ND [0.40]	ND [0.37]
8270E	mg/kg	3-Methylphenol/4-Methylphenol Coelution	6.1	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	3-Nitroaniline	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	4-Bromophenyl phenyl ether	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	4-Chloro-3-methylphenol	NE	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	4-Chloroaniline	0.015	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	4-Chlorophenyl phenyl ether	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	4-Nitroaniline	NE	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	4-Nitrophenol	NE	ND [0.55]	ND [0.45]	ND [0.52]	ND [0.50]	ND [0.46]
8270E	mg/kg	Acenaphthene	37	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Acenaphthylene	18	ND [0.013]	ND [0.011]	ND [0.012]	ND [0.012]	ND [0.011]
8270E	mg/kg	Anthracene	390	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(a)anthracene	0.7	ND [0.022]	ND [0.018]	ND [0.021]	ND [0.020]	ND [0.018]
8270E	mg/kg	Benzo(a)pyrene	1.5	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(b)fluoranthene	15	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(g,h,i)perylene	2300	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzo(k)fluoranthene	150	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]
8270E	mg/kg	Benzoic acid	200	ND [0.66]	ND [0.54]	ND [0.62]	ND [0.60]	ND [0.55]
8270E	mg/kg	Benzyl alcohol	5.7	ND [0.55]	ND [0.45]	ND [0.52]	ND [0.50]	ND [0.46]
8270E	mg/kg	Benzyl butyl phthalate	16	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	bis-(2-Chloroethoxy)methane	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	bis-(2-Chloroethyl)ether	0.00042	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	bis-(2-Ethylhexyl)phthalate	88	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Carbazole	NE	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Chrysene	600	ND [0.011]	ND [0.0089]	0.0056 [0.010] J JN	ND [0.010] JN	ND [0.0092]
8270E	mg/kg	Dibenzo(a,h)anthracene	1.5	ND [0.022]	ND [0.018]	ND [0.021]	ND [0.020]	ND [0.018]
8270E	mg/kg	Dibenzofuran	0.97	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Diethyl phthalate	60	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Dimethyl phthalate	48	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Di-n-butyl phthalate	16	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Di-n-octyl phthalate	370	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Fluoranthene	590	ND [0.011]	ND [0.0089]	0.0098 [0.010] J JN	0.0053 [0.010] J JN	ND [0.0092]
8270E	mg/kg	Fluorene	36	ND [0.011]	ND [0.0089]	ND [0.010]	ND [0.010]	ND [0.0092]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

[] - Laboratory LOD

Solid shade indicates screening value exceedance

NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 11 SD 25-056 10/11/2025 10:28 580-154935-12 Sediment	26-HOM-CW 12 SD 25-056 10/11/2025 10:50 580-154935-13 Sediment	26-HOM-CW 21 SD 25-056 10/08/2025 22:15 580-154935-4 Sediment	26-HOM-CW 20 SD 25-056 10/08/2025 22:00 580-154935-3 Sediment	26-HOM-CW 22 SD 25-056 10/09/2025 00:20 580-154935-5 Sediment	
Method	Units	Analyte	ADEC	Dup of 21 SD				
8270E	mg/kg	Hexachlorobenzene	0.0082	ND [0.055]	ND [0.044]	ND [0.051]	ND [0.050]	ND [0.045]
8270E	mg/kg	Hexachlorobutadiene	0.02	ND [0.066]	ND [0.054]	ND [0.062]	ND [0.060]	ND [0.055]
8270E	mg/kg	Hexachlorocyclopentadiene	0.0093	R	R	R	R	R
8270E	mg/kg	Hexachloroethane	0.018	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	Indeno(1,2,3-cd)pyrene	15	ND [0.013]	ND [0.011]	ND [0.012]	ND [0.012]	ND [0.011]
8270E	mg/kg	Isophorone	2.7	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Naphthalene	0.038	0.069 [0.022]	ND [0.018]	ND [0.021]	ND [0.020]	ND [0.018]
8270E	mg/kg	Nitrobenzene	0.0079	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	n-Nitrosodimethylamine	0.0000033	ND [0.11]	ND [0.089]	ND [0.10]	ND [0.10]	ND [0.092]
8270E	mg/kg	n-Nitrosodi-n-propylamine	0.00068	ND [0.083]	ND [0.067]	ND [0.078]	ND [0.075]	ND [0.069]
8270E	mg/kg	n-Nitrosodiphenylamine	4.6	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Pentachlorophenol	0.0043	ND [0.22]	ND [0.18]	ND [0.21]	ND [0.20]	ND [0.18]
8270E	mg/kg	Phenanthrene	39	ND [0.013]	ND [0.011]	0.0067 [0.012] J	0.0067 [0.012] J	ND [0.011]
8270E	mg/kg	Phenol	29	ND [0.055]	ND [0.045]	ND [0.052]	ND [0.050]	ND [0.046]
8270E	mg/kg	Pyrene	87	ND [0.011]	0.0075 [0.0089] J B	0.0096 [0.010] J B	ND [0.010]	ND [0.0092]
8270ESIM	mg/kg	1-Methylnaphthalene	0.41	ND [0.0022]	0.00094 [0.0018] J	ND [0.0021]	ND [0.0020]	ND [0.0018]
8270ESIM	mg/kg	2-Methylnaphthalene	1.3	0.0026 [0.0044] J	ND [0.0036]	ND [0.0041]	ND [0.0040]	ND [0.0037]
8270ESIM	mg/kg	Acenaphthene	37	ND [0.0022]	0.0012 [0.0018] J	0.0012 [0.0021] J JN	ND [0.0020] JN	ND [0.0018]
8270ESIM	mg/kg	Acenaphthylene	18	0.00062 [0.0022] J	0.0010 [0.0018] J	0.00091 [0.0021] J JN	ND [0.0020] JN	ND [0.0018]
8270ESIM	mg/kg	Anthracene	390	0.0011 [0.0022] J B	0.0011 [0.0018] J B	0.0026 [0.0021] JN	0.0012 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(a)anthracene	0.7	ND [0.0022]	ND [0.0018]	0.0034 [0.0021] JN	0.0014 [0.0020] J	ND [0.0018]
8270ESIM	mg/kg	Benzo(a)pyrene	1.5	ND [0.0022]	0.00091 [0.0018] J B	0.0024 [0.0021] J JN	0.0012 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(b)fluoranthene	15	ND [0.0022]	0.0011 [0.0018] J B	0.0030 [0.0021] JN	0.0013 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(g,h,i)perylene	2300	ND [0.0022]	0.0016 [0.0018] J	0.0024 [0.0021] J JN	0.0014 [0.0020] J JN	ND [0.0018]
8270ESIM	mg/kg	Benzo(k)fluoranthene	150	ND [0.0022]	ND [0.0018]	0.0016 [0.0021] J	ND [0.0020]	ND [0.0018]
8270ESIM	mg/kg	Chrysene	600	0.00082 [0.0022] J B	ND [0.0018]	0.0030 [0.0021] JN	0.0017 [0.0020] J JN	0.00079 [0.0018] J B
8270ESIM	mg/kg	Dibenzo(a,h)anthracene	1.5	ND [0.0022]	ND [0.0018]	0.0011 [0.0021] J JN	ND [0.0020] JN	ND [0.0018]
8270ESIM	mg/kg	Fluoranthene	590	0.0024 [0.0022] J B	0.0020 [0.0018] J B	0.0073 [0.0021] JN	0.0043 [0.0020] JN	0.0016 [0.0018] J B
8270ESIM	mg/kg	Fluorene	36	0.0020 [0.0022] J	0.0012 [0.0018] J	0.0018 [0.0021] J	0.0012 [0.0020] J	ND [0.0018]
8270ESIM	mg/kg	Indeno(1,2,3-cd)pyrene	15	ND [0.0022]	ND [0.0018]	0.0021 [0.0021] J	ND [0.0020]	ND [0.0018]
8270ESIM	mg/kg	Naphthalene	0.038	0.062 [0.0044]	0.0038 [0.0036] J	ND [0.0041]	ND [0.0040]	ND [0.0037]
8270ESIM	mg/kg	Phenanthrene	39	0.0040 [0.0033] B	0.0018 [0.0027] J B	0.0058 [0.0031] B	0.0039 [0.0030] B	0.0015 [0.0028] J
8270ESIM	mg/kg	Pyrene	87	0.0023 [0.0022] J B	0.0038 [0.0018] B	0.0075 [0.0021] B	0.0059 [0.0020] B	0.0025 [0.0018] B
AK101	mg/kg	Gasoline Range Organics (C6-C10)	300	ND [4.7] J-	ND [0.72]	ND [1.1] J-	ND [0.78] J-	ND [0.75]

1 - MS/MSD was run only on 8270E and 8270ESIM methods
ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil
[] - Laboratory LOD
Solid shade indicates screening value exceedance
NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

				Sample ID Location ID	26-HOM-CW 11 SD 25-056	26-HOM-CW 12 SD 25-056	26-HOM-CW 21 SD 25-056	26-HOM-CW 20 SD 25-056	26-HOM-CW 22 SD 25-056
				Collection Date	10/11/2025 10:28	10/11/2025 10:50	10/08/2025 22:15	10/08/2025 22:00	10/09/2025 00:20
				Lab Sample ID Matrix	580-154935-12 Sediment	580-154935-13 Sediment	580-154935-4 Sediment	580-154935-3 Sediment	580-154935-5 Sediment
Method	Units	Analyte	ADEC	Dup of 21 SD					
D2216	PERCENT	Percent Moisture	NE	40.0 [1.0]	26.0 [1.0]	36.5 [1.0]	34.4 [1.0]	27.8 [1.0]	

1 - MS/MSD was run only on 8270E and 8270ESIM methods
ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil
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**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	ADEC	MS/MSD ¹				
6020B	mg/kg	Antimony	4.6	0.42 [0.20]	0.43 [0.20]	0.53 [0.19]	0.34 [0.15]	0.30 [0.20]
6020B	mg/kg	Arsenic	0.2	10 [0.41]	9.9 [0.41]	15 [0.39]	10 [0.29]	11 [0.40]
6020B	mg/kg	Cadmium	9.1	0.098 [0.10] J	0.13 [0.10]	0.13 [0.097]	0.11 [0.073]	0.11 [0.10] J
6020B	mg/kg	Chromium	100000	39 [0.49]	52 [0.49]	44 [0.47]	41 [0.35]	42 [0.48]
6020B	mg/kg	Copper	370	32 [0.46]	44 [0.46]	40 [0.44]	34 [0.33]	37 [0.45]
6020B	mg/kg	Lead	400	7.3 [0.20]	10 [0.20]	9.1 [0.19]	7.8 [0.15]	8.7 [0.20]
6020B	mg/kg	Nickel	340	30 [0.48]	39 [0.49]	36 [0.46]	33 [0.35]	33 [0.48]
6020B	mg/kg	Selenium	6.9	0.20 [0.26] J	0.33 [0.26] J	0.28 [0.24] J	0.19 [0.18] J	0.22 [0.25] J
6020B	mg/kg	Silver	11	0.085 [0.10] J	0.12 [0.10] J	0.10 [0.097] J	0.085 [0.073] J	0.10 [0.10] J
6020B	mg/kg	Zinc	4900	78 [10]	100 [10]	90 [9.7]	82 [7.3]	86 [10]
7471B	mg/kg	Mercury	0.36	0.095 [0.053]	0.12 [0.060]	0.11 [0.052]	0.098 [0.051]	0.11 [0.056]
8081B	mg/kg	4,4'-DDD	0.098	ND [0.011]	ND [0.012]	ND [0.011]	ND [0.011]	ND [0.012]
8081B	mg/kg	4,4'-DDE	0.72	ND [0.0092]	ND [0.011]	ND [0.0095]	ND [0.0092]	ND [0.010]
8081B	mg/kg	4,4'-DDT	5.1	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Aldrin	0.0099	ND [0.0047]	ND [0.0054]	ND [0.0049]	ND [0.0048]	ND [0.0052]
8081B	mg/kg	alpha-BHC	0.0029	ND [0.0050]	ND [0.0057]	ND [0.0052]	ND [0.0050]	ND [0.0055]
8081B	mg/kg	beta-BHC	0.01	ND [0.0059]	ND [0.0068]	ND [0.0061]	ND [0.0059]	ND [0.0065]
8081B	mg/kg	cis-Chlordane	0.18	ND [0.0061]	ND [0.0069]	ND [0.0063]	ND [0.0061]	ND [0.0066]
8081B	mg/kg	delta-BHC	NE	ND [0.0059]	ND [0.0068]	ND [0.0061]	ND [0.0059]	ND [0.0065]
8081B	mg/kg	Dieldrin	0.0047	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Endosulfan I	9.3	ND [0.0040]	ND [0.0045]	ND [0.0041]	ND [0.0040]	ND [0.0043]
8081B	mg/kg	Endosulfan II	9.3	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8081B	mg/kg	Endosulfan sulfate	9.3	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Endrin	0.61	ND [0.0079]	ND [0.0090]	ND [0.0082]	ND [0.0079]	ND [0.0086]
8081B	mg/kg	Endrin aldehyde	NE	ND [0.011]	ND [0.013]	ND [0.012]	ND [0.011]	ND [0.012]
8081B	mg/kg	Endrin ketone	NE	ND [0.012]	ND [0.014]	ND [0.012]	ND [0.012]	ND [0.013]
8081B	mg/kg	gamma-BHC (Lindane)	0.016	ND [0.0040]	ND [0.0045]	ND [0.0041]	ND [0.0040]	ND [0.0043]
8081B	mg/kg	Heptachlor	0.0076	ND [0.0041]	ND [0.0047]	ND [0.0042]	ND [0.0041]	ND [0.0045]
8081B	mg/kg	Heptachlor Epoxide	0.0019	ND [0.0046]	ND [0.0053]	ND [0.0048]	ND [0.0046]	ND [0.0050]
8081B	mg/kg	Methoxychlor	13	ND [0.039]	ND [0.044]	ND [0.040]	ND [0.039]	ND [0.043]
8081B	mg/kg	Toxaphene	0.72	ND [0.18]	ND [0.21]	ND [0.19]	ND [0.18]	ND [0.20]
8081B	mg/kg	trans-Chlordane	0.18	ND [0.0040]	ND [0.0045]	ND [0.0041]	ND [0.0040]	ND [0.0043]
8082A	mg/kg	PCB-1016 (Aroclor 1016)	1	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

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Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	ADEC	MS/MSD ¹				
8082A	mg/kg	PCB-1221 (Aroclor 1221)	1	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1232 (Aroclor 1232)	1	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1242 (Aroclor 1242)	1	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1248 (Aroclor 1248)	1	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1254 (Aroclor 1254)	1	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8082A	mg/kg	PCB-1260 (Aroclor 1260)	1	ND [0.015]	ND [0.017]	ND [0.015]	ND [0.015]	ND [0.016]
8260D	mg/kg	1,1,1-Trichloroethane	32	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,1-Dichloroethane	0.092	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,1-Dichloroethene	1.2	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,1-Dichloropropene	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2,3-Trichlorobenzene	0.15	ND [0.35]	ND [0.35]	ND [0.43]	ND [0.28]	ND [0.44]
8260D	mg/kg	1,2,3-Trichloropropane	0.000031	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2,4-Trichlorobenzene	0.082	ND [0.35]	ND [0.35]	ND [0.43]	ND [0.28]	ND [0.44]
8260D	mg/kg	1,2,4-Trimethylbenzene	0.61	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2-Dibromo-3-chloropropane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2-Dichlorobenzene	2.4	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,2-Dichloropropane	0.03	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,3,5-Trimethylbenzene	0.66	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,3-Dichlorobenzene	2.3	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	1,3-Dichloropropane	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	2,2-Dichloropropane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	2-Butanone	15	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	2-Chlorotoluene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	4-Chlorotoluene	NE	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	4-Isopropyltoluene	NE	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	4-Methyl-2-pentanone	18	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Acetone	38	ND [0.70] J-	ND [0.70] J-	ND [0.85] J-	ND [0.57] J-	ND [0.88] J-
8260D	mg/kg	Bromobenzene	0.36	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	Bromochloromethane	NE	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Carbon disulfide	2.9	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Carbon tetrachloride	0.021	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Chlorobenzene	0.46	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Chloroethane	72	ND [0.17] J-	ND [0.17] J-	ND [0.21] J-	ND [0.14] J-	ND [0.22] J-
8260D	mg/kg	Chloromethane	0.61	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	cis-1,2-Dichloroethene	0.12	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

[] - Laboratory LOD

Solid shade indicates screening value exceedance

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Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	ADEC	MS/MSD ¹				
8260D	mg/kg	Dichlorodifluoromethane	3.9	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Ethylbenzene	0.13	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	Isopropylbenzene	5.6	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	Methylene chloride	0.33	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Methyl-tert-butyl ether (MTBE)	0.4	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	n-Butylbenzene	20	ND [0.22]	ND [0.22]	ND [0.27]	ND [0.18]	ND [0.27]
8260D	mg/kg	n-Propylbenzene	9.1	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	o-Xylene	1.5	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	sec-Butylbenzene	28	ND [0.17]	ND [0.17]	ND [0.21]	ND [0.14]	ND [0.22]
8260D	mg/kg	Styrene	10	ND [0.044]	ND [0.043]	ND [0.053]	ND [0.036]	ND [0.055]
8260D	mg/kg	tert-Butylbenzene	11	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Tetrachloroethene (PCE)	0.19	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Toluene	6.7	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	trans-1,2-Dichloroethene	1.3	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Trichloroethene (TCE)	0.011	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Trichlorofluoromethane	41	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Vinyl chloride	0.0008	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8260D	mg/kg	Xylene, isomers m & p	1.5	ND [0.087]	ND [0.087]	ND [0.11]	ND [0.071]	ND [0.11]
8270E	mg/kg	1,2,4-Trichlorobenzene	0.082	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1,2-Dichlorobenzene	2.4	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1,3-Dichlorobenzene	2.3	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1,4-Dichlorobenzene	0.037	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	1-Methylnaphthalene	0.41	ND [0.0088]	ND [0.0099]	ND [0.0091] JN	ND [0.0089]	ND [0.0097]
8270E	mg/kg	2,2-oxybis(1-Chloropropane)	NE	ND [0.053]	ND [0.060]	ND [0.055]	ND [0.053]	ND [0.058]
8270E	mg/kg	2,4,5-Trichlorophenol	28	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	2,4,6-Trichlorophenol	0.092	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	2,4-Dichlorophenol	0.21	ND [0.053]	ND [0.060]	ND [0.055] JN	ND [0.053]	ND [0.058]
8270E	mg/kg	2,4-Dimethylphenol	3.2	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	2,4-Dinitrophenol	0.34	ND [1.1]	ND [1.3]	R	ND [1.1]	ND [1.2]
8270E	mg/kg	2,4-Dinitrotoluene	0.024	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	2,6-Dinitrotoluene	0.005	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	2-Chloronaphthalene	26	ND [0.035]	ND [0.040]	ND [0.037]	ND [0.035]	ND [0.039]
8270E	mg/kg	2-Chlorophenol	0.71	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	2-Methyl-4,6-dinitrophenol	NE	ND [0.44]	ND [0.50]	ND [0.46] J-	ND [0.44]	ND [0.48]
8270E	mg/kg	2-Methylnaphthalene	1.3	ND [0.013]	ND [0.015]	ND [0.014] JN	ND [0.013]	ND [0.014]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

[] - Laboratory LOD

Solid shade indicates screening value exceedance

NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	ADEC	MS/MSD ¹				
8270E	mg/kg	2-Methylphenol (o-Cresol)	6.2	ND [0.053]	ND [0.060]	ND [0.055]	ND [0.053]	ND [0.058]
8270E	mg/kg	2-Nitroaniline	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	2-Nitrophenol	NE	ND [0.053]	ND [0.060]	ND [0.055] JN	ND [0.053]	ND [0.058]
8270E	mg/kg	3,3'-Dichlorobenzidine	0.056	ND [0.35]	ND [0.40]	ND [0.37] JN	ND [0.35]	ND [0.39]
8270E	mg/kg	3-Methylphenol/4-Methylphenol Coelution	6.1	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	3-Nitroaniline	NE	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	4-Bromophenyl phenyl ether	NE	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	4-Chloro-3-methylphenol	NE	ND [0.053]	ND [0.060]	ND [0.055]	ND [0.053]	ND [0.058]
8270E	mg/kg	4-Chloroaniline	0.015	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	4-Chlorophenyl phenyl ether	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	4-Nitroaniline	NE	ND [0.088]	ND [0.099]	ND [0.091]	ND [0.089]	ND [0.097]
8270E	mg/kg	4-Nitrophenol	NE	ND [0.44]	ND [0.50]	ND [0.46]	ND [0.44]	ND [0.48]
8270E	mg/kg	Acenaphthene	37	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Acenaphthylene	18	ND [0.011]	ND [0.012]	ND [0.011]	ND [0.011]	ND [0.012]
8270E	mg/kg	Anthracene	390	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(a)anthracene	0.7	0.013 [0.018] J	ND [0.020]	ND [0.018] J-	ND [0.018]	ND [0.019]
8270E	mg/kg	Benzo(a)pyrene	1.5	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(b)fluoranthene	15	0.0096 [0.0088] J	ND [0.0099]	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(g,h,i)perylene	2300	0.0069 [0.0088] J	ND [0.0099]	ND [0.0091] JN	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzo(k)fluoranthene	150	0.0061 [0.0088] J	ND [0.0099]	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Benzoic acid	200	ND [0.53]	ND [0.60]	R	ND [0.53]	ND [0.58]
8270E	mg/kg	Benzyl alcohol	5.7	ND [0.44]	ND [0.50]	ND [0.46]	ND [0.44]	ND [0.48]
8270E	mg/kg	Benzyl butyl phthalate	16	ND [0.18]	ND [0.20]	ND [0.18]	ND [0.18]	ND [0.19]
8270E	mg/kg	bis-(2-Chloroethoxy)methane	NE	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	bis-(2-Chloroethyl)ether	0.00042	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	bis-(2-Ethylhexyl)phthalate	88	ND [0.18]	ND [0.20]	ND [0.18]	ND [0.18]	ND [0.19]
8270E	mg/kg	Carbazole	NE	ND [0.044]	ND [0.050]	ND [0.046] J-	ND [0.044]	ND [0.048]
8270E	mg/kg	Chrysene	600	0.011 [0.0088] J	0.0066 [0.0099] J	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Dibenzo(a,h)anthracene	1.5	ND [0.018]	ND [0.020]	ND [0.018]	ND [0.018]	ND [0.019]
8270E	mg/kg	Dibenzofuran	0.97	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Diethyl phthalate	60	ND [0.18]	ND [0.20]	ND [0.18] J-	ND [0.18]	ND [0.19]
8270E	mg/kg	Dimethyl phthalate	48	ND [0.18]	ND [0.20]	ND [0.18] J-	ND [0.18]	ND [0.19]
8270E	mg/kg	Di-n-butyl phthalate	16	ND [0.18]	ND [0.20]	ND [0.18] JN	ND [0.18]	ND [0.19]
8270E	mg/kg	Di-n-octyl phthalate	370	ND [0.18]	ND [0.20]	ND [0.18] J-	ND [0.18]	ND [0.19]
8270E	mg/kg	Fluoranthene	590	0.019 [0.0088] J B	0.0088 [0.0099] J B	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270E	mg/kg	Fluorene	36	ND [0.0088]	ND [0.0099]	ND [0.0091]	ND [0.0089]	ND [0.0097]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

[] - Laboratory LOD

Solid shade indicates screening value exceedance

NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 23 SD 25-056 10/09/2025 00:40 580-154935-6 Sediment	26-HOM-CW 31 SD 25-056 10/10/2025 20:13 580-154935-9 Sediment	26-HOM-CW 32 SD 25-056 10/10/2025 21:06 580-154935-10 Sediment	26-HOM-CW 33 SD 25-056 10/10/2025 21:15 580-154935-11 Sediment	26-HOM-CW 81 SD 25-056 10/07/2025 15:15 580-154935-1 Sediment	
Method	Units	Analyte	ADEC	MS/MSD ¹				
8270E	mg/kg	Hexachlorobenzene	0.0082	ND [0.044]	ND [0.049]	ND [0.045]	ND [0.044]	ND [0.048]
8270E	mg/kg	Hexachlorobutadiene	0.02	ND [0.053]	ND [0.060]	ND [0.055] JN	ND [0.053]	ND [0.058]
8270E	mg/kg	Hexachlorocyclopentadiene	0.0093	R	R	R	R	R
8270E	mg/kg	Hexachloroethane	0.018	ND [0.088]	ND [0.099]	ND [0.091] J-	ND [0.089]	ND [0.097]
8270E	mg/kg	Indeno(1,2,3-cd)pyrene	15	0.0086 [0.011] J	ND [0.012]	ND [0.011]	0.0060 [0.011] J	ND [0.012]
8270E	mg/kg	Isophorone	2.7	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Naphthalene	0.038	ND [0.018]	ND [0.020]	ND [0.018] J-	ND [0.018]	ND [0.019]
8270E	mg/kg	Nitrobenzene	0.0079	ND [0.044]	ND [0.050]	ND [0.046] JN	ND [0.044]	ND [0.048]
8270E	mg/kg	n-Nitrosodimethylamine	0.0000033	ND [0.088]	ND [0.099]	ND [0.091] JN	ND [0.089]	ND [0.097]
8270E	mg/kg	n-Nitrosodi-n-propylamine	0.00068	ND [0.066]	ND [0.074]	ND [0.069] JN	ND [0.066]	ND [0.072]
8270E	mg/kg	n-Nitrosodiphenylamine	4.6	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Pentachlorophenol	0.0043	ND [0.18]	ND [0.20]	ND [0.18]	ND [0.18]	ND [0.19]
8270E	mg/kg	Phenanthrene	39	0.010 [0.011] J	ND [0.012]	ND [0.011] J-	ND [0.011]	ND [0.012]
8270E	mg/kg	Phenol	29	ND [0.044]	ND [0.050]	ND [0.046]	ND [0.044]	ND [0.048]
8270E	mg/kg	Pyrene	87	0.015 [0.0088] J B	0.011 [0.0099] J B	ND [0.0091] J-	ND [0.0089]	ND [0.0097]
8270ESIM	mg/kg	1-Methylnaphthalene	0.41	ND [0.018]	ND [0.0020]	ND [0.018] JN	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	2-Methylnaphthalene	1.3	ND [0.035]	ND [0.0040]	ND [0.037]	ND [0.0035]	ND [0.0039]
8270ESIM	mg/kg	Acenaphthene	37	ND [0.018]	ND [0.0020]	ND [0.018] JN	0.00096 [0.0018] J	ND [0.0019]
8270ESIM	mg/kg	Acenaphthylene	18	ND [0.018]	ND [0.0020]	ND [0.018] JN	0.00052 [0.0018] J	ND [0.0019]
8270ESIM	mg/kg	Anthracene	390	ND [0.018]	0.0010 [0.0020] J B	ND [0.018]	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(a)anthracene	0.7	ND [0.018]	0.0036 [0.0020] B	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(a)pyrene	1.5	ND [0.018]	0.0021 [0.0020] J B	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(b)fluoranthene	15	0.012 [0.018] J	0.0029 [0.0020] B	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Benzo(g,h,i)perylene	2300	ND [0.018]	0.0017 [0.0020] J	ND [0.018] JN	0.0010 [0.0018] J	0.00098 [0.0019] J
8270ESIM	mg/kg	Benzo(k)fluoranthene	150	ND [0.018]	0.0015 [0.0020] J	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Chrysene	600	0.0082 [0.018] J B	0.0030 [0.0020] B	ND [0.018] J-	ND [0.0018]	0.00094 [0.0019] J B
8270ESIM	mg/kg	Dibenzo(a,h)anthracene	1.5	ND [0.018]	ND [0.0020]	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Fluoranthene	590	0.017 [0.018] J	0.0068 [0.0020] B	ND [0.018] JN	0.0020 [0.0018] J B	0.0023 [0.0019] J B
8270ESIM	mg/kg	Fluorene	36	ND [0.018]	ND [0.0020]	ND [0.018]	0.00091 [0.0018] J	ND [0.0019]
8270ESIM	mg/kg	Indeno(1,2,3-cd)pyrene	15	ND [0.018]	0.0014 [0.0020] J	ND [0.018] J-	ND [0.0018]	ND [0.0019]
8270ESIM	mg/kg	Naphthalene	0.038	ND [0.035]	ND [0.0040]	ND [0.037]	ND [0.0035]	ND [0.0039]
8270ESIM	mg/kg	Phenanthrene	39	ND [0.026]	0.0023 [0.0030] J B	ND [0.027]	ND [0.0027]	0.0019 [0.0029] J B
8270ESIM	mg/kg	Pyrene	87	0.017 [0.018] J	0.0072 [0.0020] B	ND [0.018]	0.0027 [0.0018] B	0.0032 [0.0019] B
AK101	mg/kg	Gasoline Range Organics (C6-C10)	300	ND [0.75]	ND [0.75] J-	ND [0.92]	ND [0.61] J-	ND [0.95]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

[] - Laboratory LOD

Solid shade indicates screening value exceedance

NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

				Sample ID	26-HOM-CW 23 SD	26-HOM-CW 31 SD	26-HOM-CW 32 SD	26-HOM-CW 33 SD	26-HOM-CW 81 SD
				Location ID	25-056	25-056	25-056	25-056	25-056
				Collection Date	10/09/2025 00:40	10/10/2025 20:13	10/10/2025 21:06	10/10/2025 21:15	10/07/2025 15:15
				Lab Sample ID	580-154935-6	580-154935-9	580-154935-10	580-154935-11	580-154935-1
				Matrix	Sediment	Sediment	Sediment	Sediment	Sediment
Method	Units	Analyte	ADEC	MS/MSD ¹					
D2216	PERCENT	Percent Moisture	NE	24.6 [1.0]	33.6 [1.0]	27.3 [1.0]	25.0 [1.0]	31.3 [1.0]	

1 - MS/MSD was run only on 8270E and 8270ESIM methods
ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil
[] - Laboratory LOD
Solid shade indicates screening value exceedance
NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 82 SD 25-056 10/07/2025 16:15 580-154935-2 Sediment	26-HOM-CW 91 SD 25-056 10/09/2025 11:40 580-154935-8 Sediment	26-HOM-CW 90 SD 25-056 10/09/2025 11:30 580-154935-7 Sediment	TB 1 25-056 10/06/2025 00:00 580-154935-14 Soil
Method	Units	Analyte	ADEC	Dup of 91 SD		
6020B	mg/kg	Antimony	4.6	0.21 [0.16]	0.26 [0.16]	0.17 [0.15] J
6020B	mg/kg	Arsenic	0.2	7.8 [0.31]	13 [0.33]	12 [0.30]
6020B	mg/kg	Cadmium	9.1	0.10 [0.079]	0.057 [0.082] J	ND [0.075]
6020B	mg/kg	Chromium	100000	47 [0.38]	22 [0.39]	22 [0.36]
6020B	mg/kg	Copper	370	40 [0.35]	15 [0.37]	15 [0.34]
6020B	mg/kg	Lead	400	9.0 [0.16]	4.4 [0.16]	4.6 [0.15]
6020B	mg/kg	Nickel	340	36 [0.37]	16 [0.39]	18 [0.36]
6020B	mg/kg	Selenium	6.9	0.21 [0.20] J	0.14 [0.20] J	0.15 [0.19] J
6020B	mg/kg	Silver	11	0.092 [0.079] J	0.046 [0.082] J	0.038 [0.075] J
6020B	mg/kg	Zinc	4900	94 [7.9]	42 [8.2]	46 [7.5]
7471B	mg/kg	Mercury	0.36	0.11 [0.055]	0.049 [0.049] J	0.040 [0.046] J
8081B	mg/kg	4,4'-DDD	0.098	ND [0.011]	ND [0.021]	ND [0.0095]
8081B	mg/kg	4,4'-DDE	0.72	ND [0.0096]	ND [0.018]	ND [0.0083]
8081B	mg/kg	4,4'-DDT	5.1	ND [0.0082]	ND [0.016]	ND [0.0071]
8081B	mg/kg	Aldrin	0.0099	ND [0.0049]	ND [0.0093]	ND [0.0043]
8081B	mg/kg	alpha-BHC	0.0029	ND [0.0052]	ND [0.0098]	ND [0.0045]
8081B	mg/kg	beta-BHC	0.01	ND [0.0062]	ND [0.012]	ND [0.0053]
8081B	mg/kg	cis-Chlordane	0.18	ND [0.0063]	ND [0.012]	ND [0.0055]
8081B	mg/kg	delta-BHC	NE	ND [0.0062]	ND [0.012]	ND [0.0053]
8081B	mg/kg	Dieldrin	0.0047	ND [0.0082]	ND [0.016]	ND [0.0071]
8081B	mg/kg	Endosulfan I	9.3	ND [0.0041]	ND [0.0078]	ND [0.0036]
8081B	mg/kg	Endosulfan II	9.3	ND [0.015]	ND [0.028]	ND [0.013]
8081B	mg/kg	Endosulfan sulfate	9.3	ND [0.0082]	ND [0.016]	ND [0.0071]
8081B	mg/kg	Endrin	0.61	ND [0.0082]	ND [0.016]	ND [0.0071]
8081B	mg/kg	Endrin aldehyde	NE	ND [0.012]	ND [0.022]	ND [0.010]
8081B	mg/kg	Endrin ketone	NE	ND [0.012]	ND [0.023]	ND [0.011]
8081B	mg/kg	gamma-BHC (Lindane)	0.016	ND [0.0041]	ND [0.0078]	ND [0.0036]
8081B	mg/kg	Heptachlor	0.0076	ND [0.0043]	ND [0.0080]	ND [0.0037]
8081B	mg/kg	Heptachlor Epoxide	0.0019	ND [0.0048]	ND [0.0090]	ND [0.0042]
8081B	mg/kg	Methoxychlor	13	ND [0.041]	ND [0.076]	ND [0.035]
8081B	mg/kg	Toxaphene	0.72	ND [0.19]	ND [0.36]	ND [0.17]
8081B	mg/kg	trans-Chlordane	0.18	ND [0.0041]	ND [0.0078]	ND [0.0036]
8082A	mg/kg	PCB-1016 (Aroclor 1016)	1	ND [0.015]	ND [0.014]	ND [0.013]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

[] - Laboratory LOD

Solid shade indicates screening value exceedance

NE - Not Established

**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 82 SD 25-056 10/07/2025 16:15 580-154935-2 Sediment	26-HOM-CW 91 SD 25-056 10/09/2025 11:40 580-154935-8 Sediment	26-HOM-CW 90 SD 25-056 10/09/2025 11:30 580-154935-7 Sediment	TB 1 25-056 10/06/2025 00:00 580-154935-14 Soil	
Method	Units	Analyte	ADEC	Dup of 91 SD			
8082A	mg/kg	PCB-1221 (Aroclor 1221)	1	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1232 (Aroclor 1232)	1	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1242 (Aroclor 1242)	1	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1248 (Aroclor 1248)	1	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1254 (Aroclor 1254)	1	ND [0.015]	ND [0.014]	ND [0.013]	
8082A	mg/kg	PCB-1260 (Aroclor 1260)	1	ND [0.015]	ND [0.014]	ND [0.013]	
8260D	mg/kg	1,1,1-Trichloroethane	32	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,1-Dichloroethane	0.092	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,1-Dichloroethene	1.2	ND [0.082]	ND [0.070]	ND [0.065]	
8260D	mg/kg	1,1-Dichloropropene	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2,3-Trichlorobenzene	0.15	ND [0.33]	ND [0.28]	ND [0.26]	ND [0.40] J-
8260D	mg/kg	1,2,3-Trichloropropane	0.000031	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2,4-Trichlorobenzene	0.082	ND [0.33]	ND [0.28]	ND [0.26]	ND [0.40] J-
8260D	mg/kg	1,2,4-Trimethylbenzene	0.61	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2-Dibromo-3-chloropropane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2-Dichlorobenzene	2.4	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,2-Dichloropropane	0.03	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,3,5-Trimethylbenzene	0.66	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,3-Dichlorobenzene	2.3	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	1,3-Dichloropropane	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	2,2-Dichloropropane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	2-Butanone	15	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	2-Chlorotoluene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	4-Chlorotoluene	NE	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	4-Isopropyltoluene	NE	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	4-Methyl-2-pentanone	18	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	Acetone	38	ND [0.65] J-	ND [0.56] J-	ND [0.52] J-	ND [0.80] J-
8260D	mg/kg	Bromobenzene	0.36	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	Bromochloromethane	NE	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Carbon disulfide	2.9	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Carbon tetrachloride	0.021	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	Chlorobenzene	0.46	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Chloroethane	72	ND [0.16] J-	ND [0.14] J-	ND [0.13] J-	ND [0.20] J-
8260D	mg/kg	Chloromethane	0.61	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	cis-1,2-Dichloroethene	0.12	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

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**Table 3: 2025 Homer Feasibility Study
Data Table
Comparison to ADEC Criteria**

		Sample ID Location ID Collection Date Lab Sample ID Matrix	26-HOM-CW 82 SD 25-056 10/07/2025 16:15 580-154935-2 Sediment	26-HOM-CW 91 SD 25-056 10/09/2025 11:40 580-154935-8 Sediment	26-HOM-CW 90 SD 25-056 10/09/2025 11:30 580-154935-7 Sediment	TB 1 25-056 10/06/2025 00:00 580-154935-14 Soil	
Method	Units	Analyte	ADEC	Dup of 91 SD			
8260D	mg/kg	Dichlorodifluoromethane	3.9	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Ethylbenzene	0.13	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	Isopropylbenzene	5.6	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	Methylene chloride	0.33	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	Methyl-tert-butyl ether (MTBE)	0.4	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	n-Butylbenzene	20	ND [0.20]	ND [0.18]	ND [0.16]	ND [0.25] J-
8260D	mg/kg	n-Propylbenzene	9.1	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	o-Xylene	1.5	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	sec-Butylbenzene	28	ND [0.16]	ND [0.14]	ND [0.13]	ND [0.20] J-
8260D	mg/kg	Styrene	10	ND [0.041]	ND [0.035]	ND [0.032]	ND [0.050] J-
8260D	mg/kg	tert-Butylbenzene	11	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Tetrachloroethene (PCE)	0.19	ND [0.082]	ND [0.070]	ND [0.065]	
8260D	mg/kg	Toluene	6.7	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	trans-1,2-Dichloroethene	1.3	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Trichloroethene (TCE)	0.011	ND [0.082]	ND [0.070]	ND [0.065]	
8260D	mg/kg	Trichlorofluoromethane	41	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8260D	mg/kg	Vinyl chloride	0.0008	ND [0.082]	ND [0.070]	ND [0.065]	
8260D	mg/kg	Xylene, Isomers m & p	1.5	ND [0.082]	ND [0.070]	ND [0.065]	ND [0.10] J-
8270E	mg/kg	1,2,4-Trichlorobenzene	0.082	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	1,2-Dichlorobenzene	2.4	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	1,3-Dichlorobenzene	2.3	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	1,4-Dichlorobenzene	0.037	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	1-Methylnaphthalene	0.41	ND [0.0092]	ND [0.0086]	ND [0.0079]	
8270E	mg/kg	2,2-oxybis(1-Chloropropane)	NE	ND [0.055]	ND [0.052]	ND [0.047]	
8270E	mg/kg	2,4,5-Trichlorophenol	28	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	2,4,6-Trichlorophenol	0.092	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	2,4-Dichlorophenol	0.21	ND [0.055]	ND [0.052]	ND [0.047]	
8270E	mg/kg	2,4-Dimethylphenol	3.2	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	2,4-Dinitrophenol	0.34	ND [1.2]	ND [1.1]	ND [0.99]	
8270E	mg/kg	2,4-Dinitrotoluene	0.024	ND [0.092]	ND [0.086]	ND [0.079]	
8270E	mg/kg	2,6-Dinitrotoluene	0.005	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	2-Chloronaphthalene	26	ND [0.037]	ND [0.034]	ND [0.032]	
8270E	mg/kg	2-Chlorophenol	0.71	ND [0.046]	ND [0.043]	ND [0.040]	
8270E	mg/kg	2-Methyl-4,6-dinitrophenol	NE	ND [0.46]	ND [0.43]	ND [0.40]	
8270E	mg/kg	2-Methylnaphthalene	1.3	ND [0.014]	ND [0.013]	ND [0.012]	

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

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Method	Units	Analyte	ADEC	Dup of 91 SD		
8270E	mg/kg	2-Methylphenol (o-Cresol)	6.2	ND [0.055]	ND [0.052]	ND [0.047]
8270E	mg/kg	2-Nitroaniline	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	2-Nitrophenol	NE	ND [0.055]	ND [0.052]	ND [0.047]
8270E	mg/kg	3,3'-Dichlorobenzidine	0.056	ND [0.37]	ND [0.34]	ND [0.32]
8270E	mg/kg	3-Methylphenol/4-Methylphenol Coelution	6.1	ND [0.046]	0.21 [0.043]	ND [0.040]
8270E	mg/kg	3-Nitroaniline	NE	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	4-Bromophenyl phenyl ether	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	4-Chloro-3-methylphenol	NE	ND [0.055]	ND [0.052]	ND [0.047]
8270E	mg/kg	4-Chloroaniline	0.015	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	4-Chlorophenyl phenyl ether	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	4-Nitroaniline	NE	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	4-Nitrophenol	NE	ND [0.46]	ND [0.43]	ND [0.40]
8270E	mg/kg	Acenaphthene	37	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Acenaphthylene	18	ND [0.011]	ND [0.010]	ND [0.0095]
8270E	mg/kg	Anthracene	390	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(a)anthracene	0.7	ND [0.018]	ND [0.017]	ND [0.016]
8270E	mg/kg	Benzo(a)pyrene	1.5	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(b)fluoranthene	15	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(g,h,i)perylene	2300	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzo(k)fluoranthene	150	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Benzoic acid	200	ND [0.55]	ND [0.52]	ND [0.47]
8270E	mg/kg	Benzyl alcohol	5.7	ND [0.46]	ND [0.43]	ND [0.40]
8270E	mg/kg	Benzyl butyl phthalate	16	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	bis-(2-Chloroethoxy)methane	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	bis-(2-Chloroethyl)ether	0.00042	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	bis-(2-Ethylhexyl)phthalate	88	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Carbazole	NE	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	Chrysene	600	ND [0.0092]	ND [0.0086]	ND [0.0079]
8270E	mg/kg	Dibenzo(a,h)anthracene	1.5	ND [0.018]	ND [0.017]	ND [0.016]
8270E	mg/kg	Dibenzofuran	0.97	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	Diethyl phthalate	60	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Dimethyl phthalate	48	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Di-n-butyl phthalate	16	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Di-n-octyl phthalate	370	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Fluoranthene	590	ND [0.0092]	0.0053 [0.0086] J B	ND [0.0079]
8270E	mg/kg	Fluorene	36	ND [0.0092]	ND [0.0086]	ND [0.0079]

1 - MS/MSD was run only on 8270E and 8270ESIM methods

ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil

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Method	Units	Analyte	ADEC	Dup of 91 SD		
8270E	mg/kg	Hexachlorobenzene	0.0082	ND [0.045]	ND [0.043]	ND [0.039]
8270E	mg/kg	Hexachlorobutadiene	0.02	ND [0.055]	ND [0.052]	ND [0.047]
8270E	mg/kg	Hexachlorocyclopentadiene	0.0093	R	R	R
8270E	mg/kg	Hexachloroethane	0.018	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	Indeno(1,2,3-cd)pyrene	15	ND [0.011]	ND [0.010]	ND [0.0095]
8270E	mg/kg	Isophorone	2.7	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	Naphthalene	0.038	ND [0.018]	ND [0.017]	ND [0.016]
8270E	mg/kg	Nitrobenzene	0.0079	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	n-Nitrosodimethylamine	0.0000033	ND [0.092]	ND [0.086]	ND [0.079]
8270E	mg/kg	n-Nitrosodi-n-propylamine	0.00068	ND [0.069]	ND [0.064]	ND [0.059]
8270E	mg/kg	n-Nitrosodiphenylamine	4.6	ND [0.046]	ND [0.043]	ND [0.040]
8270E	mg/kg	Pentachlorophenol	0.0043	ND [0.18]	ND [0.17]	ND [0.16]
8270E	mg/kg	Phenanthrene	39	ND [0.011]	ND [0.010]	ND [0.0095]
8270E	mg/kg	Phenol	29	ND [0.046]	0.062 [0.043]	ND [0.040]
8270E	mg/kg	Pyrene	87	ND [0.0092]	0.0067 [0.0086] J B	0.0055 [0.0079] J B
8270ESIM	mg/kg	1-Methylnaphthalene	0.41	ND [0.0018]	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	2-Methylnaphthalene	1.3	ND [0.0037]	ND [0.0034]	ND [0.0032]
8270ESIM	mg/kg	Acenaphthene	37	ND [0.0018]	0.00087 [0.0017] J JN	ND [0.0016] JN
8270ESIM	mg/kg	Acenaphthylene	18	ND [0.0018]	0.00092 [0.0017] J JN	ND [0.0016] JN
8270ESIM	mg/kg	Anthracene	390	ND [0.0018]	0.0016 [0.0017] J JN	0.00088 [0.0016] J JN
8270ESIM	mg/kg	Benzo(a)anthracene	0.7	ND [0.0018]	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	Benzo(a)pyrene	1.5	ND [0.0018]	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	Benzo(b)fluoranthene	15	ND [0.0018]	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	Benzo(g,h,i)perylene	2300	0.0013 [0.0018] J	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	Benzo(k)fluoranthene	150	ND [0.0018]	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	Chrysene	600	0.00078 [0.0018] J B	0.0013 [0.0017] J B	0.00055 [0.0016] J B
8270ESIM	mg/kg	Dibenzo(a,h)anthracene	1.5	ND [0.0018]	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	Fluoranthene	590	0.0015 [0.0018] J B	0.0034 [0.0017] JN	0.0018 [0.0016] J JN
8270ESIM	mg/kg	Fluorene	36	ND [0.0018]	0.0012 [0.0017] J	ND [0.0016]
8270ESIM	mg/kg	Indeno(1,2,3-cd)pyrene	15	ND [0.0018]	ND [0.0017]	ND [0.0016]
8270ESIM	mg/kg	Naphthalene	0.038	ND [0.0037]	0.0024 [0.0034] J	ND [0.0032]
8270ESIM	mg/kg	Phenanthrene	39	0.0014 [0.0028] J B	0.0029 [0.0026] J JN	0.0015 [0.0024] J JN
8270ESIM	mg/kg	Pyrene	87	0.0023 [0.0018] B	0.0043 [0.0017] B	0.0017 [0.0016] J B
AK101	mg/kg	Gasoline Range Organics (C6-C10)	300	ND [0.70] J-	ND [0.61]	ND [0.56]
						ND [0.86]

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Data Table
Comparison to ADEC Criteria**

				Sample ID Location ID	26-HOM-CW 82 SD 25-056	26-HOM-CW 91 SD 25-056	26-HOM-CW 90 SD 25-056	TB 1 25-056
				Collection Date	10/07/2025 16:15	10/09/2025 11:40	10/09/2025 11:30	10/06/2025 00:00
				Lab Sample ID	580-154935-2	580-154935-8	580-154935-7	580-154935-14
				Matrix	Sediment	Sediment	Sediment	Soil
Method	Units	Analyte	ADEC	Dup of 91 SD				
D2216	PERCENT	Percent Moisture	NE	27.8 [1.0]	22.8 [1.0]	16.4 [1.0]		

1 - MS/MSD was run only on 8270E and 8270ESIM methods
ADEC - 18 AAC 75 Method 2 Table B1/B2 Cleanup Levels for soil
[] - Laboratory LOD
Solid shade indicates screening value exceedance
NE - Not Established

**Table 4: Homer Feasibility Study
DMMP LOD Summary**

Method	Analyte	DMMP PAL	Number of Failures	Total Samples
8081B	4,4'-DDD	0.016	1	13
8081B	4,4'-DDE	0.009	12	13
8081B	4,4'-DDT	0.012	1	13
8081B	cis-Chlordane	0.0028	13	13
8081B	Dieldrin	0.0019	13	13
8081B	Heptachlor	0.0015	13	13
8081B	trans-Chlordane	0.0028	13	13
8260D	1,2,4-Trichlorobenzene	0.031	14	14
8260D	1,2-Dichlorobenzene	0.035	14	14
8270E	1,2,4-Trichlorobenzene	0.031	13	13
8270E	1,2-Dichlorobenzene	0.035	13	13
8270E	2,4-Dimethylphenol	0.029	13	13
8270E	2-Methylphenol (o-Cresol)	0.063	1	13
8270E	Benzoic acid	0.65	1	13
8270E	Benzyl alcohol	0.057	13	13
8270E	Benzyl butyl phthalate	0.063	13	13
8270E	Diethyl phthalate	0.2	2	13
8270E	Dimethyl phthalate	0.071	13	13
8270E	Hexachlorobenzene	0.022	13	13
8270E	Hexachlorobutadiene	0.011	13	13
8270E	n-Nitrosodiphenylamine	0.028	13	13

**Table 5: Homer Feasibility Study
ADEC LOD Summary**

Method	Analyte	ADEC PAL	Number of Failures	Total Samples
8081B	alpha-BHC	0.0029	13	13
8081B	beta-BHC	0.01	1	13
8081B	Dieldrin	0.0047	13	13
8081B	Heptachlor	0.0076	1	13
8081B	Heptachlor Epoxide	0.0019	13	13
8260D	1,1-Dichloroethane	0.092	5	14
8260D	1,1-Dichloroethene	1.2	1	13
8260D	1,2,3-Trichlorobenzene	0.15	14	14
8260D	1,2,3-Trichloropropane	0.000031	14	14
8260D	1,2,4-Trichlorobenzene	0.082	14	14
8260D	1,2,4-Trimethylbenzene	0.61	1	14
8260D	1,2-Dichloropropane	0.03	14	14
8260D	1,3,5-Trimethylbenzene	0.66	1	14
8260D	Bromobenzene	0.36	1	14
8260D	Carbon tetrachloride	0.021	14	14
8260D	Chlorobenzene	0.46	1	14
8260D	Chloromethane	0.61	1	14
8260D	cis-1,2-Dichloroethene	0.12	2	14
8260D	Ethylbenzene	0.13	1	14
8260D	Methylene chloride	0.33	1	14
8260D	Methyl-tert-butyl ether (MTBE)	0.4	1	14
8260D	Tetrachloroethene (PCE)	0.19	1	13
8260D	trans-1,2-Dichloroethene	1.3	1	14
8260D	Trichloroethene (TCE)	0.011	13	13
8260D	Vinyl chloride	0.0008	13	13
8270E	1,4-Dichlorobenzene	0.037	13	13
8270E	2,4-Dinitrophenol	0.34	13	13
8270E	2,4-Dinitrotoluene	0.024	13	13
8270E	2,6-Dinitrotoluene	0.005	13	13
8270E	3,3'-Dichlorobenzidine	0.056	13	13
8270E	4-Chloroaniline	0.015	13	13
8270E	bis-(2-Chloroethyl)ether	0.00042	13	13
8270E	Hexachlorobenzene	0.0082	13	13
8270E	Hexachlorobutadiene	0.02	13	13
8270E	Hexachlorocyclopentadiene	0.0093	13	13
8270E	Hexachloroethane	0.018	13	13
8270E	Nitrobenzene	0.0079	13	13
8270E	n-Nitrosodimethylamine	0.0000033	13	13
8270E	n-Nitrosodi-n-propylamine	0.00068	13	13
8270E	Pentachlorophenol	0.0043	13	13

Data Flag Explanations

ND - Analyte is not detected; [] - Laboratory Limit of Detection (LOD)

	Analyte LOD is greater than the screening criteria
	Analyte was detected at a concentration greater than the screening criteria.
Qualifier	Definition
J	Analyte result is considered an estimated value because the level is below the laboratory LOQ but above the DL
B	Analyte result is considered a high estimated value due to contamination present in the method, trip, or equipment blank.
J+, J-, JN	Analyte result is considered an estimated value biased (high, low, uncertain) due to a quality control failure
R	Analyte result is rejected - result is not usable for project.

**Attachment 1:
ADEC Data Review Checklist
SDG: 580-154935-1/2**

ADEC Contaminated Sites Program Laboratory Data Review Checklist

Completed By:	Mike Utley	CS Site Name:	Homer Harbor	Lab Name:	Eurofins
Title:	Homer Harbor Dredged Material Characterization	ADEC File No.:	N/A	Lab Report No.:	580-154935
Consulting Firm:	USACE	Hazard ID No.:	N/A	Lab Report Date:	01-Dec-2025

Note: Any N/A or No box checked must have an explanation in the comments box.

1. Laboratory

- a. Did an ADEC Contaminated Sites Laboratory Approval Program (CS-LAP) approved laboratory receive and perform all of the submitted sample analyses?
 Yes No N/A

Comments: *The Eurofins Anchorage Service Center (ASR) received all samples. They submitted samples to EURO Lancaster who is approved for all analyses (6020B, 7471B, 8081B, 8082A, 8260D, 8270E, 8270ESIM, and AK101). They also submitted samples for Organotins SIM (Tributyl Tin via GC/MS SIM) to EURO CAL Science who is **not approved** by ADEC for that method/analyte.*

- b. If the samples were transferred to another “network” laboratory or sub-contracted to an alternate laboratory, was the laboratory performing the analyses CS-LAP approved?
 Yes No N/A
 Comments: See above.

2. Chain of Custody (CoC)

- a. Is the CoC information completed, signed, and dated (including released/received by)?
 Yes No N/A
 Comments: First page of CoC was signed by all parties, but following pages were not.

- b. Were the correct analyses requested?
 Yes No N/A
 Analyses requested: 6020B, 7471B, 8081B, 8082A, 8260D, 8270E, 8270ESIM, AK101, D2216

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Comments: Anchorage Receiving Station (ASR) received all samples in one cooler. As noted above, ASR sent samples to EURO Lancaster and EURO Cal Science. However, the CoC for the transfer to EURO Lancaster only indicated that samples for 6020B, 7471B, PRE-SCREEN, and D2216 were being transferred (detail regarding the other methods were not documented). As per the original CoC, three jars per sample were collected and provided to ASR. Lab Transfer CoC to Lancaster indicates that two jars per sample were provided, and Lab Transfer CoC to Cal Science indicates that one jar per sample was provided. Given that the laboratory documentation and EDD indicate that Lancaster analyzed the samples for all methods (except Tributyl Tin), it can be safely assumed that each sample jar is accounted for, in spite of the lack of documentation of shipment for each method.

3. Laboratory Sample Receipt Documentation

- a. Is the sample/cooler temperature documented and within range at receipt (0° to 6° C)?

Yes No N/A

Cooler temperature(s): Samples were received at the Anchorage Service Center in one cooler and received at 1.3° C. The Service Center submitted samples to two different network labs: EURO Lancaster (rec'd at 1.5° C); and EURO Cal Science (rec'd at 3.5° C).

Sample temperature(s): Not applicable

Comments: Click or tap here to enter text.

- b. Is the sample preservation acceptable – acidified waters, methanol preserved soil (GRO, BTEX, VOCs, etc.)?

Yes No N/A

Comments: Click or tap here to enter text.

- c. Is the sample condition documented – broken, leaking, zero headspace (VOA vials); canister vacuum/pressure checked and no open valves, etc.?

Yes No N/A

Comments: No deviations were noted on the case narrative or in sample receiving documentation.

- d. If there were any discrepancies, were they documented? For example, incorrect sample containers/preservation, sample temperature outside of acceptable range, insufficient or missing samples, canister not holding a vacuum, etc.?

Yes No N/A

Comments: No deviations were noted on the case narrative or in sample receiving documentation.

- e. Is the data quality or usability affected?

Yes No N/A

Comments: No deviations were noted.

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4. Case Narrative

- a. Is the case narrative present and understandable?

Yes No N/A

Comments: Click or tap here to enter text.

- b. Are there discrepancies, errors, or QC failures identified by the lab?

Yes No N/A

Comments: Click or tap here to enter text.

- c. Were all the corrective actions documented?

Yes No N/A

Comments: Click or tap here to enter text.

- d. What is the effect on data quality/usability according to the case narrative?

Comments: Case narrative documents deficiencies but does not address usability.

5. Sample Results

- a. Are the correct analyses performed/reported as requested on CoC?

Yes No N/A

Comments: Click or tap here to enter text.

- b. Are all applicable holding times met?

Yes No N/A

Comments: Click or tap here to enter text.

- c. Are all soils reported on a dry weight basis?

Yes No N/A

Comments: Click or tap here to enter text.

- d. Are the reported limits of quantitation (LoQ) or limits of detections (LOD), or reporting limits (RL) less than the Cleanup Level or the action level for the project?

Yes No N/A

Comments: A total of 21 nondetect analytes exceeded the DMMP Screening Values in one or more samples. These results are highlighted yellow in the DMMP data table. A comparison of reported LODs to the most conservative ADEC Cleanup Limits specified in 18 AAC 75 was also completed; a total of 40 nondetect analytes exceeded the ADEC levels in at least one sample. These are also highlighted yellow in the ADEC data table.

- e. Is the data quality or usability affected?

Yes No N/A

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Comments: When a sample LOD for any given analyte exceeds the applicable screening limit, the presence (or absence) of that analyte at that limit cannot be accurately determined.

6. QC Samples

a. Method Blank

- i. Was one method blank reported per matrix, analysis, and 20 samples?

Yes No N/A

Comments: Click or tap here to enter text.

- ii. Are all method blank results less than LOQ (or RL)?

Yes No

Comments: The following analytes were detected in an associated method blank: Fluoranthene (8270 and 8270SIM), Pyrene (8270 and 8270SIM), Anthracene, Benzo(a)anthracene, Benzo(a)pyrene, Benzo(b)fluoranthene, Chrysene, and Phenanthrene (8270SIM only).

- iii. If above LoQ or RL, what samples are affected?

Comments: The following samples have at least one analyte listed above that was detected in an associated method blank: 26-HOM-CW 11 SD, 26-HOM-CW 12 SD, 26-HOM-CW 20 SD, 26-HOM-CW 21 SD, 26-HOM-CW 22 SD, 26-HOM-CW 23 SD, 26-HOM-CW 31 SD, 26-HOM-CW 33 SD, 26-HOM-CW 81 SD, 26-HOM-CW 82 SD, 26-HOM-CW 90 SD, and 26-HOM-CW 91 SD

- iv. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?

Yes No N/A

Comments: Affected sample results are qualified "B" in the associated data tables.

- v. Data quality or usability affected?

Yes No N/A

Comments: All detections are significantly below (i.e. less than 20%) of the applicable DMMP screening criterion.

b. Laboratory Control Sample/Duplicate (LCS/LCSD)

- i. Organics – Are one LCS/LCSD reported per matrix, analysis and 20 samples? (LCS/LCSD required per AK methods, LCS required per SW846)

Yes No N/A

Comments: Click or tap here to enter text.

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- ii. Metals/Inorganics – Are one LCS and one sample duplicate reported per matrix, analysis and 20 samples?
Yes No N/A
Comments: A duplicate was not run for D2216, 6020B, and 7471B.
- iii. Accuracy – Are all percent recoveries (%R) reported and within method or laboratory limits and project specified objectives, if applicable? (AK Petroleum methods: AK101 60%-120%, AK102 75%-125%, AK103 60%-120%; all other analyses see the laboratory QC pages)
Yes No N/A
Comments: Click or tap here to enter text.
- iv. Precision – Are all relative percent differences (RPD) reported and less than method or laboratory limits and project specified objectives, if applicable? Was the RPD reported from LCS/LCSD, and or sample/sample duplicate? (AK Petroleum methods 20%; all other analyses see the laboratory QC pages)
Yes No N/A
Comments: Where LCSDs or duplicates were run, all RPDs met criteria.
- v. If %R or RPD is outside of acceptable limits, what samples are affected?
Comments: No deviations were noted.
- vi. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?
Yes No N/A
Comments: No deviations were noted.
- vii. Is the data quality or usability affected?
Yes No N/A
Comments: Precision cannot be evaluated for all data for methods 6020B, 7471B, D2216, and 8082A (no duplicate, LCSD, or MSD). Likewise, precision cannot be evaluated for 8081B batch 410-717133, 8270ESIM batch 410-719765, and 8270ESIM batch 410-721118. However, all impacted results are below the applicable DMMP PAL, and data usability is not significantly impacted.
- c. Matrix Spike/Matrix Spike Duplicate (MS/MSD)
- i. Organics – Are one MS/MSD reported per matrix, analysis and 20 samples?
Yes No N/A
Comments: MS/MSD request was not indicated on chain of custody. However, the laboratory ran a MS/MSD on 8270E and one of three batches for 8270ESIM.

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- ii. Metals/Inorganics – Are one MS/MSD reported per matrix, analysis and 20 samples?
Yes No N/A
Comments: MS/MSDs were not run for methods 6020B and 7471B.
- iii. Accuracy – Are all percent recoveries (%R) reported and within method or laboratory limits and project specified objectives, if applicable?
Yes No N/A
Comments: MS/MSD was run on only 8270E and 8270ESIM, on sample 26-HOM-CW 32 SD. Several analytes for both methods failed low in either the MS, the MSD, or both. Analytes which failed low but recovered above 10% are flagged “J-“, and analytes which recovered below 10% are flagged “R” (unusable). Only the associated sample results (26-HOM-CW 32 SD) are flagged.
- iv. Precision – Are all relative percent differences (RPD) reported and less than method or laboratory limits and project specified objectives, if applicable? RPD reported from MS/MSD, and or sample/sample duplicate.
Yes No N/A
Comments: Several analytes also failed RPD for the reported MS/MSD. If the analyte failed MS and/or MSD recoveries (but above 10%), the “J-“ flag was changed to “JN” to indicate potential bias of unknown direction. R flags due to lack of MS/MSD recoveries were not changed. Again, this pertains only to the impacted sample 26-HOM-CW 32 SD.
- v. If %R or RPD is outside of acceptable limits, what samples are affected?
Comments: Only sample 26-HOM-CW 32 SD, methods 8270E and 8270ESIM, were impacted and flagged.
- vi. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?
Yes No N/A
Comments: See above.
- vii. Is the data quality or usability affected?
Yes No N/A
Comments: Results for 2,4-Dinitrophenol, Benzoic Acid, and Hexachlorocyclopentadiene in sample 26-HOM-CW 32 SD are rejected due to low biased MS/MSD recoveries (less than 10%). Usability for low biased results or RPD failures is not significantly impacted.

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d. Surrogates – Organics Only or Isotope Dilution Analytes (IDA) – Isotope Dilution Methods Only

- i. Are surrogate/IDA recoveries reported for organic analyses – field, QC, and laboratory samples?

Yes No N/A

Comments: Click or tap here to enter text.

- ii. Accuracy – Are all percent recoveries (%R) reported and within method or laboratory limits and project specified objectives, if applicable? (AK Petroleum methods 50-150 %R for field samples and 60-120 %R for QC samples; all other analyses see the laboratory report pages)

Yes No N/A

Comments: One of four surrogates failed low in samples 26-HOM-CW 12 SD and TB 1 for 8260D but recovered greater than 10%. Consequently, all analytes for these samples are flagged “J-“. One of one surrogate failed low in multiple samples for AK101 where results indicate the samples were diluted significantly (i.e. at least 25X). Results are not further flagged.

- iii. Do the sample results with failed surrogate/IDA recoveries have data flags? If so, are the data flags clearly defined?

Yes No N/A

Comments: Results are flagged as noted above in the associated data tables.

- iv. Is the data quality or usability affected?

Yes No N/A

Comments: impacted 8260 analytes are all nondetect and are not significantly impacted by a slightly low surrogate recovery; AK101 results are all nondetect at a level that is significantly below the PAL (ADEC); there is no PAL for the DMMP criteria.

e. Trip Blanks

- i. Is one trip blank reported per matrix, analysis, and for each cooler containing volatile samples? Yes No N/A

Comments: Click or tap here to enter text.

- ii. Are all results less than LoQ or RL?

Yes No N/A

Comments: **Not all reported analytes are reported in trip blank. PCE, TCE, and VC are not reported; the laboratory has been asked for a revised data package. This text will be updated once a revision or explanation has been received.**

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- iii. If above LoQ or RL, what samples are affected?

Comments: Click or tap here to enter text.

- iv. Is the data quality or usability affected?

Yes No N/A

Comments: It is assumed that PCE, TCE, and VC will be reports as nondetect in the trip blank, so no impacts are anticipated.

f. Field Duplicate

- i. Are one field duplicate submitted per matrix, analysis, and 10 project samples?

Yes No N/A

Comments: Two field duplicates were collected: 26-HOM-CW 20 SD is a duplicate of 26-HOM-CW 21 SD and 26-HOM-CW 90 SD is a duplicate of 26-HOM-CW 91 SD

- ii. Was the duplicate submitted blind to lab?

Yes No N/A

Comments: Click or tap here to enter text.

- iii. Precision – All relative percent differences (RPD) less than specified project objectives? (Recommended: 30% water or air, 50% soil)

$$RPD (\%) = \left| \frac{R_1 - R_2}{\left(\frac{R_1 + R_2}{2}\right)} \right| \times 100$$

Where R_1 = Sample Concentration

R_2 = Field Duplicate Concentration

Yes No N/A

Comments: For primary/duplicate pair -21 SD/-20 SD, the following methods/analytes did not meeting ADEC requirements: 8270E: Chrysene and Fluoranthene; 8270ESIM - Acenaphthene, Acenaphthylene, Anthracene, Benzo(a)anthracene, Benzo(a)pyrene, Benzo(b)fluoranthene, Benzo(g,h,i)perylene, Chrysene, Dibenzo(a,h)anthracene, and Fluoranthene.

For primary duplicate pair -91 SD/-90 SD, the following methods/analytes did not meet ADEC requirements: 8081, nearly all analytes; 8270ESIM - Phenanthrene, Fluoranthene, Acenaphthene, Anthracene, Acenaphthylene.

- iv. Is the data quality or usability affected? (Explain)

Yes No N/A

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Comments: -21 SD/-20 SD – Impacted analytes are all flagged “JN”; results are either nondetect or well below the DMMP PAL; thus, usability is not impacted by this deviation.

-91 SD/-90 SD – for the 8081 analysis, the primary sample was analyzed at a five cut and the duplicate was analyzed at a ten cut. All results are nondetect; the difference is due to differing dilutions. Therefore, no flags were applied, and usability is not impacted. For the 8270ESIM method, the impacted analytes are flagged “JN” (note that some “B” flags were changed to “JN” to indicate increased uncertainty of reported results). However, all results are nondetect and well below the DMMP PAL. Usability is not impacted.

g. Decontamination or Equipment Blanks

- i. Were decontamination or equipment blanks collected?

Yes No N/A

Comments: Click or tap here to enter text.

- ii. Are all results less than LoQ or RL?

Yes No N/A

Comments: Click or tap here to enter text.

- iii. If above LoQ or RL, specify what samples are affected.

Comments: No equipment blank was provided.

- iv. Are data quality or usability affected?

Yes No N/A

Comments: No equipment blank was provided.

7. Other Data Flags/Qualifiers (ACOE, AFCEE, Lab Specific, etc.)

- a. Are they defined and appropriate?

Yes No N/A

Comments: Acetone and chloroethane failed low (76%/76%) in the CCV associated with batch 410-716956, impacting all samples. Impacted results are flagged “J-“. All impacted results are nondetect and are well below ADEC PALs (DMMP PAL is not established for these compounds). Results are considered usable.

Lab indicates that 8270E closing CCV for Hexachlorocyclopentadiene recovered low, that CCV and samples were rerun and the closing CCV again recovered low. However, there is no indication in the data package that the samples were rerun. The original sample that was spiked was run first (26-HOM-CW 32 SD), followed by the MS and MSD, both of which did not recover (0%). The rest of the samples were analyzed next followed by the closing CCV, which recovered at

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21% (acceptable DoD Limit is 50%). For these reasons, this compound is considered rejected in all samples.